

Supporting Information

Massive Parallel Catalyst Screening: Towards Asymmetric MCRs

Ulrike Kusebauch,¹ Barbara Beck,¹ Kim Messer,¹ Eberhardt Herdtweck,² Alexander Dömling^{1*}

¹*Morphochem AG, Gmunderstr.37-37a, 81379 München, Germany.*

²*Anorganisch Chemisches Institut der Technischen Universität München, Lichtenbergstr.4, 85747 Garching, Germany.*

alexander.doemling@morphochem.de

General. Melting points were determined in open capillaries on an electrothermal IA 9100 melting point apparatus and are reported uncorrected. Proton and Carbon nmr spectra were obtained in CDCl₃ and determined with a Mercury 400 spectrometer. Chemical shifts are expressed in ppm (δ) with respect to TMS as an internal standard. NMR peak assignments were made on the basis of 2-D experiments. Electrospray ionisation mass spectra (ESI) were obtained using a MSD (Hewlett-Packards HPLC 1100 driven electrospray MS instrument). Products were purified either by crystallization with the named solvent or by preparative chromatography with silica gel and a mixture of ethylacetate (EA) and petrolether (PE) as eluent. Thin layer chromatography (TLC) and preparative thin layer chromatography (PTLC) was performed on 60 F₂₅₄ aluminum sheets and glass plates from Merck using UV detection. The identity was determined utilizing a Hewlett-Packard LC 1100 system using a YMC ODS-A column (2.1 mm x 50 mm, 3 μ m), detection wavelength at 220 nm and 254 nm; flow: 0.6 ml/min, with a 2.5 min gradient from 90 % H₂O to 10 % H₂O (0.5 % CH₃COOH) vs. CH₃CN (0.5 % CH₃COOH). Preparative HPLC was performed on a Labomatic System, using a Grom Sil 120 ODS-4 HE column (10 μ m, 30 x 100 mm) with a 30 min methanol/water gradient (40-100 % MeOH + 0.5 % CH₃COOH) with single wavelength detection at 254 nm, flow: 50 ml/min. IR spectra were recorded on a Nexus FT-IR apparatus from Thermo-Nicolet. The intensity of the absorption bands are characterized as s = strong, m = medium, w = weak and br = broad.

Experimental Procedures

General procedure for the preparation of benzoic acid 1-benzylcarbamoyl-2-methyl-propyl ester (**16**) and analogous Passerini products:

5 Mmol (610 mg) benzoic acid (**15**) are diluted in 10 ml diethylether. To the solution 5 mmol (360 mg) *iso*-butyric aldehyde (**14**) and 5 mmol (586 mg) benzyl isocyanide (**13**) are added. The reaction mixture is allowed to stir over night at room temperature. The crystallized product is filtered off, washed with cold diethylether and dried under high vacuum to give 1.30 g (84 %) as a solid. All described Passerini reactions were performed in scheduled quantities and were crystallized or purified by preparative HPLC.

General screening procedure for Passerini reaction with chiral catalyst:

0.25 mmol chiral ligand are diluted in 125 μ L THF (dry) in a 2 ml reaction flask with septum and magnetic stirrer under nitrogen atmosphere. Via a Hamilton syringe 100 μ L of the lewis acid (**A**) are added and stirred for 30 minutes at the respective temperature (25°C, 0°C, -18°C or -78°C). A 2.5 M stock solution of every starting material is prepared in THF, in case of catalyst to reactant ratio of 1/1 and Lewis acid to ligand ratio of 1/1, and 100 μ L are added in the ranking aldehyde, acid and isocyanide with a time delay of 30 min after each compound. The reaction mixture is stirred over night at room temperature.

For the work up 50 μ L of the reaction mixture are added to a mixture of 200 μ L THF, 200 μ L *iso*-propanol and 200 μ L saturated NaHCO₃ solution and shook vigorously. After complete separation of the two layers the upper organic layer is separated and subject to a silica based solid phase extraction column (equilibrated with 200 μ L THF) and eluted with THF.

Purification of screening reaction mixtures with chiral catalyst using HPLC:

The purification was performed on a Hewlett-Packard LC 1100 system with a HP Zorbax Eclipse XDB-C8 column (4.6 x 150 mm, 5 μ m) using a linear 14 min gradient from 10 % to 50 % MeCN (0.1 % HCOOH) vs. H₂O (0.1 % HCOOH). The corresponding UV active fraction is collected and completely evaporated under high vacuum. The residue is dissolved in *n*-hexane/*iso*-propanol (1/1) and measured on chiral stationary HPLC phase.

General procedure for Passerini reaction with chiral catalyst combination A7:

1 Mmol (466 mg) (4S,5S)-4,5-bis-(diphenylhydroxymethyl)-2,2-dimethyldioxolane (**7**) are diluted in 400 μ L THF (dry) in a 5 ml reaction flask with septum and magnetic stirrer under nitrogen atmosphere. Via a syringe 400 μ L titanisopropoxide (**A**) are added and stirred for 1h at -78°C. A 2.5 M stock solution of every starting material is prepared in 400 μ L THF are added in the ranking aldehyde, acid and isocyanide with a time delay of 60 min after each compound. The reaction mixture is stirred over night.

For the work up the reaction mixture is completely transferred with a capillary to a preparative thin layer chromatography plate and developed with a solvent mixture of petrol ether/ethyl acetate. The corresponding band is scratched out and the product is unhinged with ethyl acetate. The silica gel is filtered off and the resulting filtrate is evaporated and dried on high vacuum.

Determination of enantiomeric excess by means of chiral stationary phases:

The enantiomeric excess is determined using an Agilent HPLC 1100 and an UV-detector at 254 nm or 220 nm. The products were separated on a Chiralcel-ODH (4.6 x 250 mm, 5 μ m) stationary phase with hexane and *iso*-propanol as eluents in LiChrosolve[®] quality with a flow of 0.5 ml/min. The separations have been performed in an isocratic mode using *n*-hexane/*iso*-propanol in a ratio of 85/15, in case of product (**17**) of 90/10. Stationary phases weren't equipped with a thermostat, so that a variation in temperatures may cause differences in retention times of the enantiomers. For that reason all supposed enantiomers have been proved by mass-spectrometry. Other chiral phases used (4.6 x 50 mm): Chirex 3020, 3010, 3002 from Phenomenex.

Experimental data for the described substances:

Benzoic acid 1-benzylcarbamoyl-2-methyl-propyl ester (**16**): scheduled quantity 5 mmol gave 1.300 g (84 %) C₁₉H₂₁NO₃ MW: 311.38 g/mol; **M.p.** 112.5°C - 113.3°C; **R_f**: 0.70 (PE/EA, 1/1, v/v); **HPLC-MS (ESI)**: *t_R* = 3.65 min; *m/z* = 312 [M+H]⁺; 334 [M+Na]⁺; **¹H-NMR (CDCl₃, 400 MHz)**: δ = 1.06 (dd, 6H, ³J=11.6Hz, ⁴J=6.9Hz, 2 $\times$ CH₃); 2.47-2.54 (m, 1H, CHCH(CH₃)₂); 4.40-4.57 (m, 2H, ²J_{ab}=14.95Hz, NHCH₂), 5.37 (d, 1H, ³J=4.1Hz, CHCH(CH₃)₂); 6.44 (br, 1H, NH); 7.24-7.33 (m, 5H, 5 $\times$ CH-Ar); 7.45-7.49 (m, 2H, 2 $\times$ CH-Ar), 7.59-7.62 (m, 1H, 1 $\times$ CH-Ar), 8.06-8.08 (m, 2H, 2 $\times$ CH-Ar); **¹³C-NMR (CDCl₃, 100 MHz)**: δ = 17.32 (CH₃); 19.26 (CH₃); 31.09 (CH(CH₃)₂); 43.36 (NHCH₂); 78.87 (CHCH(CH₃)₂); 127.80 (CH-Ar); 128.93 (CH-Ar); 129.51 (CH-Ar); 129.99 (CH-Ar); 133.86 (CH-Ar); 138.15 (CH-Ar); 165.76 (CO); 169.62 (CO); **IR**: ? (cm⁻¹) = 3264 (m); 3088 (w); 2971 (m); 2359 (w); 1723 (s); 1656 (s); 1561 (m); 1292 (s), 1258 (s); 708 (s). Separation of enantiomers: *t_R*(A): 18.72 min (50 %), *t_R*(B): 21.89 min (50 %); with

chiral induction of catalyst A7: scheduled quantity 1 mmol gave 677 mg (46 %): t_R (A): 18.73 min (32 %), t_R (B): 21.90 min (68 %); ee = 36 %.

2-Hydroxy-benzoic acid 1-benzylcarbamoyl-2-methyl-propyl ester (**17**): scheduled quantity 5 mmol gave 1.516 g (93 %) $C_{19}H_{21}NO_4$ MW: 327.38 g/mol; R_f : 0.36 (PE/EA, 5/1, v/v); **HPLC-MS (ESI)**: t_R = 3.68 min; m/z = 350 $[M+Na]^+$; **1H -NMR ($CDCl_3$, 400 MHz)**: δ = 1.06 (dd, 6H, $^3J=9.3$ Hz, $^4J=6.9$ Hz, $2\times CH_3$); 2.48 (m, 1H, $CH(CH_3)_2$); 4.40-4.57 (m, 2H, $^2J_{ab}=14.9$ Hz, $^3J=5.6$ Hz, $NHCH_2$); 5.31 (d, 1H, $^3J=4.4$ Hz, $CHCH(CH_3)_2$); 6.43 (t, 1H, $^3J=5.6$ Hz, NH); 6.89-6.93 (m, 1H, $1\times CH-Ar$); 7.00-7.02 (m, 1H, $1\times CH-Ar$); 7.24-7.34 (m, 5H, $5\times CH-Ar$); 7.47-7.52 (m, 1H, $1\times CH-Ar$); 7.86-7.88 (m, 1H, $1\times CH-Ar$); 10.53 (s, 1H, OH); **^{13}C -NMR ($CDCl_3$, 100 MHz)**: δ = 17.05 (CH_3); 18.88 (CH_3); 30.73 ($CH(CH_3)_2$); 43.13 ($NHCH_2$); 78.98 ($CHCH(CH_3)_2$); 111.60 (C-Ar); 117.88 (CH-Ar); 119.40 (CH-Ar); 127.49 (CH-Ar); 128.67 (CH-Ar); 129.50 (CH-Ar); 136.34 (CH-Ar); 137.69 (C-Ar); 161.90 (ArC-OH); 168.76 (CO); 169.06 (CO). Separation of enantiomers: t_R (A): 19.88 min (50 %), t_R (B): 21.79 min (50 %); with chiral induction of catalyst A7: scheduled quantity 1 mmol gave 39 mg (12 %): t_R (A): 21.02 min (33 %), t_R (B): 23.17 min (67 %); ee = 32 %. The identity has been proved by coelution with the enantiomeric 1:1 mixture of compound **17**.

Benzoic acid 1-(3,5-dimethyl-benzylcarbamoyl)-2-methyl-propyl ester (**18**): scheduled quantity 1 mmol gave 160 mg (47 %) $C_{21}H_{25}NO_3$ MW: 339.44 g/mol; R_f : 0.60 (PE/EA, 2/1, v/v); **HPLC-MS (ESI)**: t_R = 3.84 min; m/z = 340 $[M+H]^+$; 362 $[M+Na]^+$; **1H -NMR ($CDCl_3$, 400 MHz)**: δ = 1.05-1.10 (dd, 6H, $^3J=13.9$ Hz, $^4J=6.8$ Hz, $2\times CH_3$); 2.56 (s, 6H, $2\times CH_3-Ar$); 2.46-2.54 (m, 1H, $CHCH(CH_3)_2$); 4.35-4.47 (m, 2H; $NHCH_2$); 5.35-5.37 (d, 1H, $^3J=4.3$ Hz, $CHCH(CH_3)_2$); 6.47 (m, 1H, $NHCH_2$); 6.85-6.88 (m, 3H, $3\times CH-Ar$); 7.45-7.49 (m, 2H, $2\times CH-Ar$); 7.58-7.62 (m, 1H, $1\times CH-Ar$); 8.07-8.09 (m, 2H, $2\times CH-Ar$); **^{13}C -NMR ($CDCl_3$, 100 MHz)**: δ = 16.96 (CH_3); 18.86 (CH_3); 21.05 (CH_3); 30.66 ($CHCH(CH_3)_2$); 42.85 ($NHCH_2$); 78.55 ($CHCH(CH_3)_2$); 125.17 (CH-Ar); 128.49 (CH-Ar); 128.92 (CH-Ar); 129.16 (CH-Ar); 129.59 (CH-Ar); 133.43 (CH-Ar); 137.67 (C-Ar); 138.09 (C-Ar); 165.37 (CO); 169.161 (CO). Separation of enantiomers: t_R (A): 12.05 min (50 %), t_R (B): 20.17 min (50 %). In contrast to the other performed separations, here the retention times of the two enantiomers differs in a range of 8 min. To prove these data every peak was collected and reinjected into HPLC-MS which approved the separation of the enantiomers; with chiral induction of catalyst A7: scheduled quantity 1 mmol gave 105 mg (31 %): t_R (A): 12.21 min (34 %), t_R (B): 20.15 min (66 %); ee = 32 % (for this example only few amounts were purified which leads to low concentration and low intensity in the UV spectra).

Benzoic acid 2-methyl-1-[(pyridin-3-ylmethyl)-carbamoyl]-propyl ester (**19**): scheduled quantity 1 mmol gave 237 mg (76 %) $C_{18}H_{20}N_2O_3$ MW: 312,37 g/mol; R_f : 0.22 (PE/EA, 2/1, v/v); **HPLC-MS (ESI)**: t_R = 2.88 min; m/z = 313 $[M+H]^+$; **1H -NMR ($CDCl_3$, 400 MHz)**: δ = 0.99-1.09 (m, 6H, 2x CH_3); 2.40-2.48 (m, 1H, $CHCH(CH_3)_2$); 4.36-4.52 (m, 2H, $NHCH_2$); 5.28-5.30 (m, 1H, $CHCH(CH_3)_2$); 6.69 (m, 1H, $NHCH_2$); 7.18-7.24 (m, 1H, 1x CH -Ar); 7.41-7.44 (m, 2H, 2x CH -Ar); 7.55-7.59 (m, 2H, 2x CH -Ar); 8.01-8.03 (m, 2H, 2x CH -Ar); 8.42 (m, 2H, 2x CH -Ar); **^{13}C -NMR ($CDCl_3$, 100 MHz)**: δ = 17.03 (CH_3); 18.90 (CH_3); 30.74 ($CH(CH_3)_2$); 40.50 ($NHCH_2$); 78.55 ($CHCH(CH_3)_2$); 123.59 (CH -Ar); 128.62 (CH -Ar); 129.70 (CH -Ar); 133.64 (CH -Ar); 135.47 (CH -Ar); 148.86 (CH -Ar); 165.54 (CO); 169.65 (CO). Separation of enantiomers: t_R (A): 25.20 min (33 %), t_R (B): 29.89 min (33 %), t_R (C): 34.25 min (33 %) side product, to confirm that the peaks of enantiomers A and B are correctly assigned the fractions were collected and product peaks were reinjected into HPLC-MS which approved the separation of the enantiomers; with chiral induction of catalyst A7: scheduled quantity 1 mmol gave 87 mg (28 %): t_R (A): 25.36 min (29 %), t_R (B): 29.67 min (71 %); ee = 42 %.

Benzoic acid 1-benzylcarbamoyl-2,2-dimethyl-propyl ester (**20**): scheduled quantity 1 mmol gave 263 mg (81 %) $C_{20}H_{23}NO_3$ MW: 325,41 g/mol; R_f : 0.50 (PE/EA, 3/1, v/v); **HPLC-MS (ESI)**: t_R = 3.72 min; m/z = 326 $[M+H]^+$; 348 $[M+Na]^+$; **1H -NMR ($CDCl_3$, 400 MHz)**: δ = 1.07 (s, 9H, 3x CH_3); 4.32-4.47 (m, 2H, $^2J_{ab}$ =14.92Hz, $NHCH_2$); 5.04 (s, 1H, ($CHC(CH_3)_3$)); 6.20 (m, 1H, $NHCH_2$); 7.16-7.24 (m, 5H, 5x CH -Ar); 7.38-7.42 (m, 2H, 2x CH -Ar); 7.51-7.55 (m, 1H, 1x CH -Ar); 7.99-8.01 (m, 2H, 2x CH -Ar); **^{13}C -NMR ($CDCl_3$, 100 MHz)**: δ = 26.39 (CH_3); 34.50 ($C(CH_3)_3$); 43.03 ($NHCH_2$); 81.41 ($CHC(CH_3)_3$); 127.38 (CH -Ar); 127.55 (CH -Ar); 126.57 (CH -Ar); 129.29 (C -Ar); 129.64 (CH -Ar); 133.48 (CH -Ar); 137.93 (C -Ar); 165.33 (CO); 168.31 (CO). Separation of enantiomers: t_R (A): 11.72 min (50 %), t_R (B): 14.19 min (50 %); with chiral induction of catalyst A7: scheduled quantity 1 mmol gave 156 mg (48 %): t_R (A): 11.81 min (33 %), t_R (B): 14.13 min (67 %); ee = 34 %.

Biphenyl-4-carboxylic acid 1-benzylcarbamoyl-2,2-dimethyl-propyl ester (**21**): scheduled quantity 1 mmol gave 383 mg (95 %) $C_{26}H_{27}NO_3$ MW: 401,51 g/mol; R_f : 0.40 (PE/EA, 4/1, v/v); **HPLC-MS (ESI)**: t_R = 4.08 min; m/z = 402 $[M+H]^+$; 424 $[M+Na]^+$; **1H -NMR ($CDCl_3$, 400 MHz)**: δ = 1.09 (s, 9H, 3x CH_3); 4.33-4.48 (m, 2H, $^2J_{ab}$ =14.92Hz, $NHCH_2$); 5.06 (s, 1H, $CHC(CH_3)_3$); 6.22 (t, 1H, 3J =5.5Hz, $NHCH_2$); 7.16-7.24 (m, 5H, 5x CH -Ar); 7.31-7.42 (m, 3H,

3×CH-Ar); 7.53-7.55 (m, 2H, 2×CH-Ar); 7.59-7.62 (m, 2H, 2×CH-Ar); 8.05-8.07 (m, 2H, 2×CH-Ar); ¹³C-NMR (CDCl₃, 100 MHz): δ = 26.44 (CH₃); 34.56 (C(CH₃)₃); 43.08 (NHCH₂); 81.44 (CHC(CH₃)₃); 127.23 (CH-Ar); 127.58 (CH-Ar); 127.97 (CH-Ar); 128.29 (CH-Ar); 128.62 (CH-Ar); 128.94 (CH-Ar); 130.20 (CH-Ar); 137.97 (C-Ar); 139.68 (C-Ar); 146.27 (C-Ar); 165.25 (CO); 168.36 (CO). Separation of enantiomers: t_R(A): 16.18 min (50 %), t_R(B): 26.23 min (50 %); with chiral induction of catalyst A7: scheduled quantity 1 mmol gave 185 mg (46 %): t_R(A): 16.15 min (32 %), t_R(B): 26.30 min (68 %); ee = 36 %.

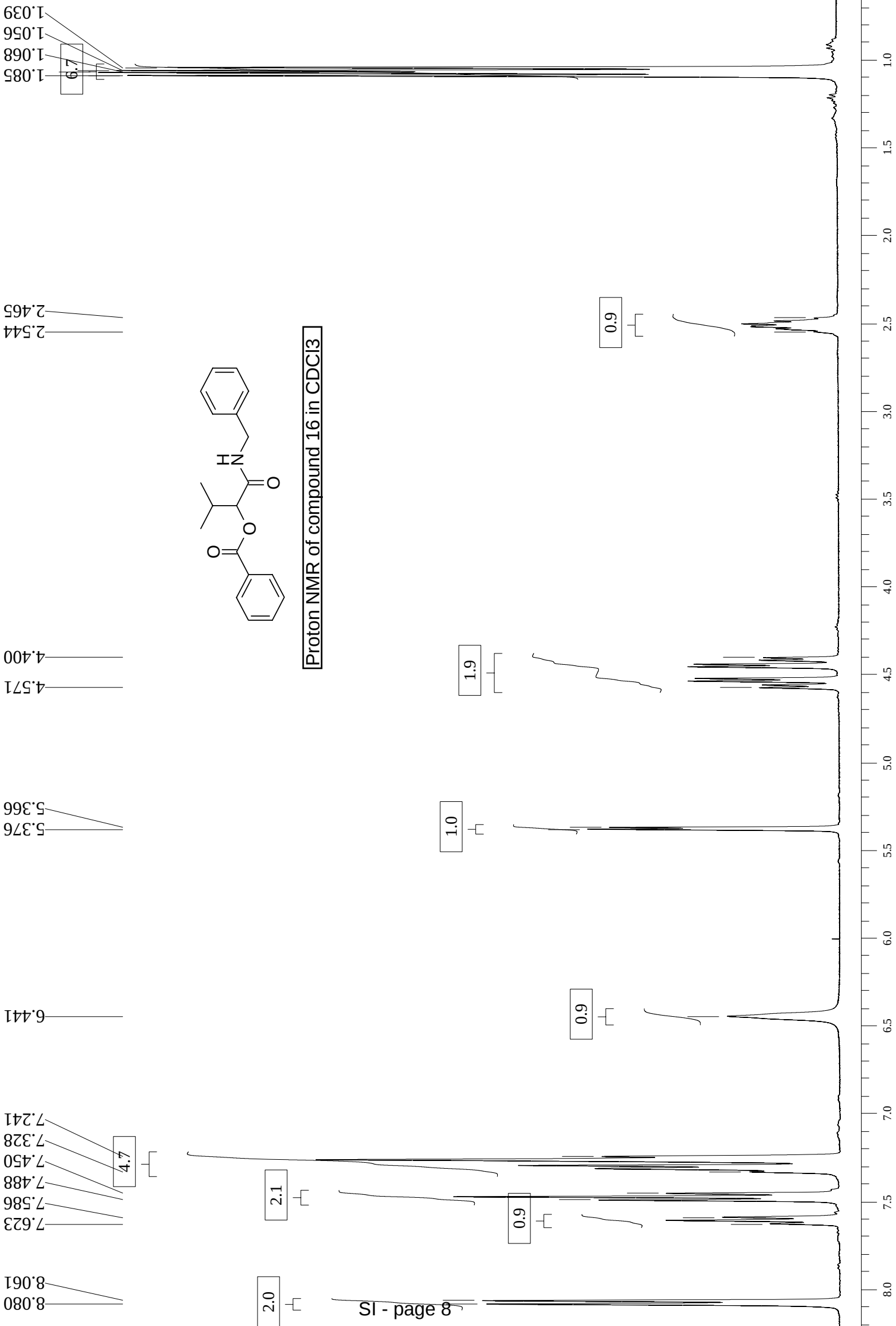
2-tert-Butoxycarbonylamino-3-phenyl-propionic acid 1-benzylcarbamoyl-2-methyl-propyl ester (**22**): scheduled quantity 5 mmol gave 1.264 g (56 %) C₂₆H₃₄N₂O₅ MW: 454.57g/mol; **HPLC-MS (ESI)**: t_R = 3.93 min; m/z = 477 [M+Na]⁺; ¹H-NMR (CDCl₃, 400 MHz): δ = 0.59-0.86 (m, 6H, 2×CH₃); 1.26-1.30 (m, 9H, 3×CH₃); 2.30-2.41 (m, 1H, CH(CH₃)₂); 3.01-3.20 (m, 2H, CCH₂CH); 4.22-4.67 (m, 2H, NHCH₂); 4.92-4.94 (br, 1H, NH); 5.06-5.07 (m, 1H, NHCHCO); 5.14 (m, 1H, CHCH(CH₃)₂); 7.16-7.31 (m, 10H, 10×CH-Ar); 7.60 (br, 1H, NH); ¹³C-NMR (CDCl₃, 100 MHz) The product is obtained as a diastereomeric mixture, all signals occur doubled: δ = 16.08 & 16.28 (1×CH₃); 18.53 & 18.95 (1×CH₃); 28.07 (3×CH₃); 29.88 & 30.23 (CH(CH₃)₂); 36.93 & 37.24 (1×CCH₂CH); 42.89 & 43.00 (1×NHCH₂); 54.82 & 55.79 (NHCH); 78.73 & 78.96 (CHCH(CH₃)₂); 80.59 & 80.64 (CCH₃); 126.94-129.06 (CH-Ar; C); 135.35 & 135.46 (C-Ar); 138.18 & 138.43 (C-Ar); 155.79 & 156.26 (CO); 169.05 (CO); 170.95 (CO); 172.11 (CO). Separation of diastereomers: injection of the mother liquor t_R(A): 11.73 min (44 %), t_R(B): 17.64 min (56 %), de = 12 %; injection of the second part of the crystal used for single crystal X-ray determination: t_R(A): 11.66 min (100 %); coelution of diastereomeric mixture and crystal t_R(A): 11.79 min (57 %), t_R(B): 17.89 min (43 %).

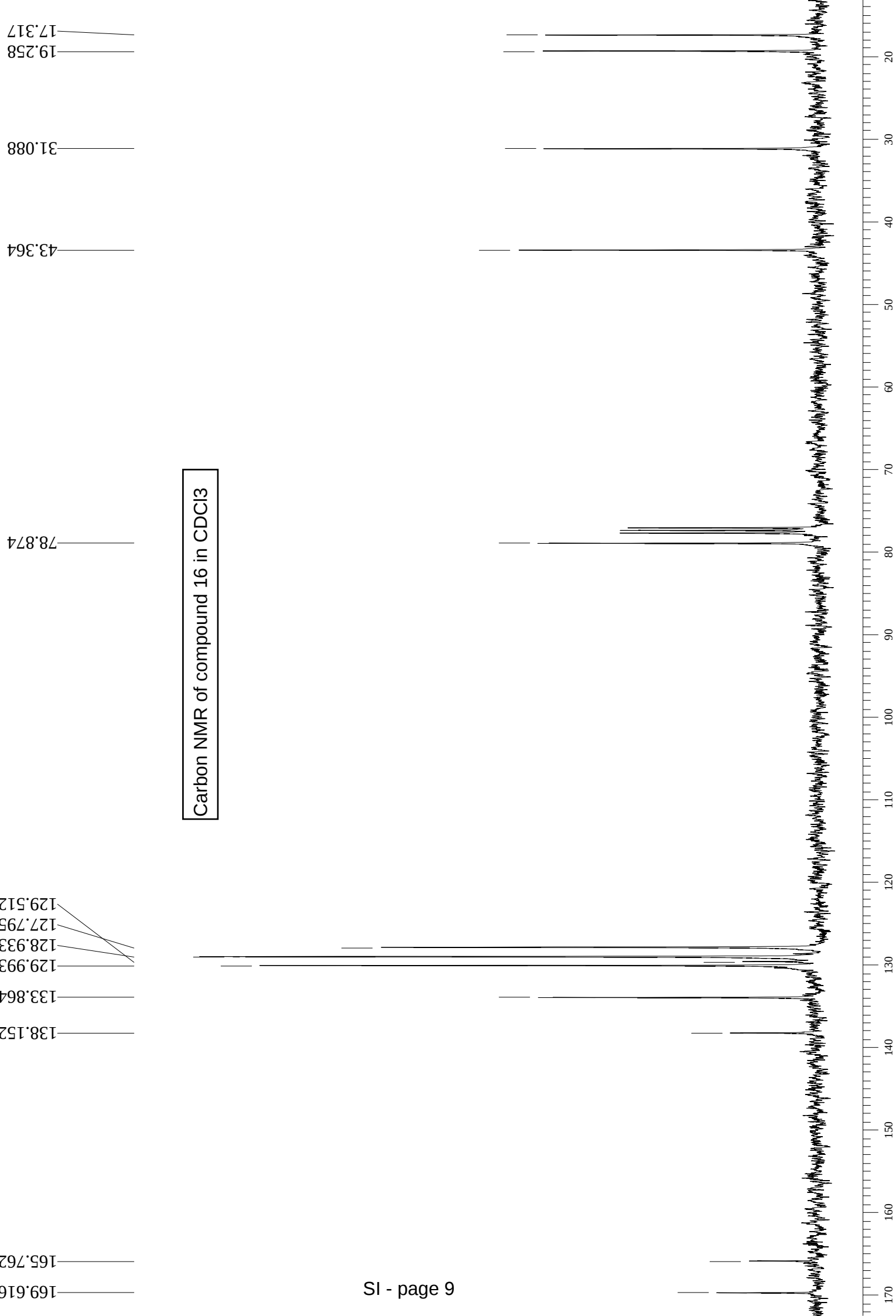
X-ray single crystal structure determination of **22**: *Crystal data*: C₂₆H₃₄N₂O₅, M_r = 454.55, colourless crystal, 0.76 × 0.64 × 0.38 mm³, orthorhombic, space group P2₁2₁2₁ (No. 19), a = 11.2376(1), b = 12.3949(1), c = 18.3839(1) Å, V = 2560.67(3) Å³, Z = 4, r_{calcd} = 1.179 g cm⁻³, F(000) = 976, m (Mo K_α) = 0.082 mm⁻¹, T = 173 K. *Data collection*: Nonius MACH3 κ-axis CCD-diffractometer, l = 0.71073 Å, unit cell parameters by least-squares for 2669 reflections. A total number of 57025 reflections were integrated. After merging, 4666 reflections [all data; R_{int} = 0.037] remained and these were used for all further calculations. *Structure solution and refinement*: Direct methods (SIR92), full-matrix least-squares refinements on F² (SHELXL-97) converged with R1 = 0.0277 [4314 data; I_o > 2σ(I_o)], wR2 =

0.0680 [all data], $GOF = 1.045$ and stopped at a maximum shift/error < 0.001. Crystallographic data (excluding structure factors) for the structure reported in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC-xxxxxx (22). Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK [Fax: (internat.) +44-1223/336-033; E-mail: deposit@ccdc.cam.ac.uk]. b) Altomare, A.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli, M. *SIR92; J. Appl. Cryst.* **1994**, 27, 435. c) Sheldrick G. M. *SHELXL-97*; University of Göttingen: Germany, **1998**.

Preparation of *N*-Benzyl-2-hydroxy-3-methyl-butyramide (**23**): 10 mg of the corresponding ester {saponification of 2-*tert*-Butoxycarbonylamino-3-phenyl-propionic acid 1-benzylcarbamoyl-2-methyl-propyl ester (**22**) in the following refers to experiment A or saponification of Benzoic acid 1-benzylcarbamoyl-2-methyl-propyl ester (**16**) in the following refers to experiment B} is dissolved in a mixture of *n*-hexane/*iso*-propanol (1/1) 40 μ L. To this solution 40 μ L of a 2N aq. NaOH solution are added and stirred for 30 min at room temperature. The formed alcohol **23** is isolated on a HPLC (HP Zorbax Eclipse XDB-C8 column) The solvent is completely evaporated under high vacuum and the residue is dissolved in *n*-hexane/*iso*-propanol (1/1) and measured on chiral HPLC phase.

N-Benzyl-2-hydroxy-3-methyl-butyramide (**23**): $C_{12}H_{17}NO_2$ MW: 207.27g/mol; **HPLC-MS (ESI)**: $t_R = 2.95$ min; $m/z = 208 [M+H]^+$; 230 $[M+Na]^+$; Experiment A: saponification of the crystal **22** leads to alcohol **23**; HPLC on Chiralcel-ODH (4.6 x 250): $t_R(A)$: 14.57 min as a single peak; Experiment B: saponification of the ester **16** with chiral induction of catalyst A7: $t_R(A)$: 14.70 min, $t_R(B)$: 15.55 min these peaks show no baseline separation; peak A which corresponds to the alcohol **23** in *R*-configuration from experiment A shows less intensity than peak B. The co-injection approved the induction of the *S*-enantiomer: $t_R(A)$: 14.60 min, $t_R(B)$: 15.46 min.

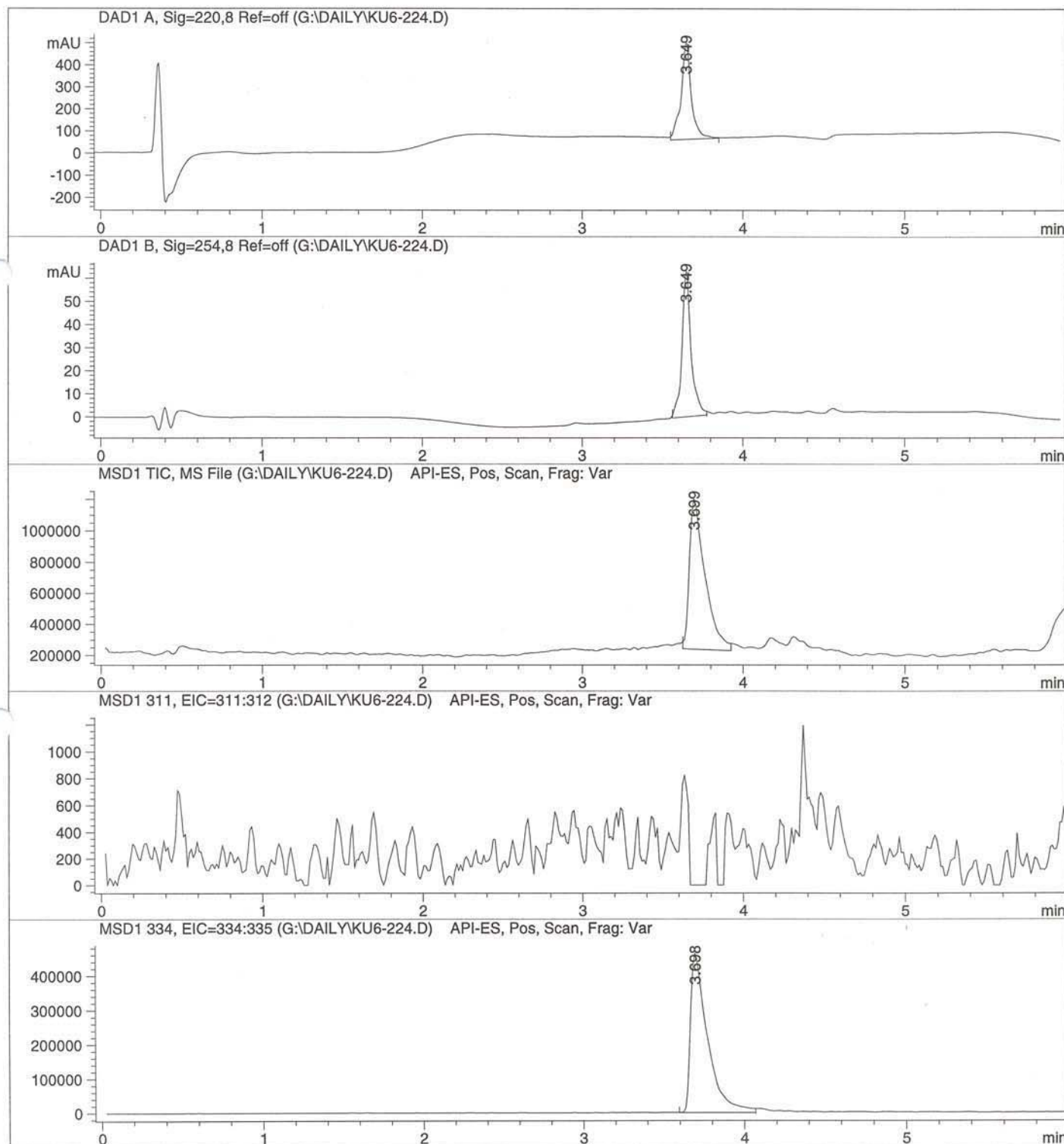




```
=====
Injection Date   : 8/12/02 2:44:07 PM      Seq. Line :   44
Sample Name     : KU6B100-2_RK            Location  : Vial 21
Acq. Operator   : admin                   Inj       :    1
                                           Inj Volume: 5 µl

Acq. Method     : D:\HPCHEM\1\METHODS\ESI_LC6.M
Last changed    : 8/12/02 2:40:46 PM by admin
                  (modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\ESI_LC6.M
Last changed    : 6/18/02 3:31:34 PM
YMC ODS-A 5cm 2µ
ESI positiv
```

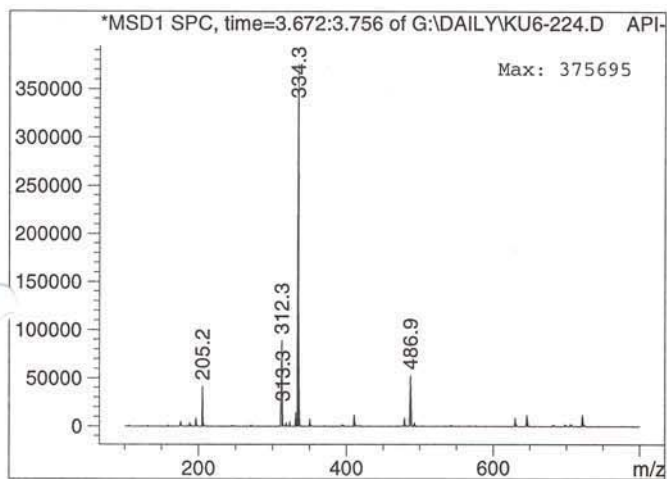
HPLC-MS Spectra of compound 16



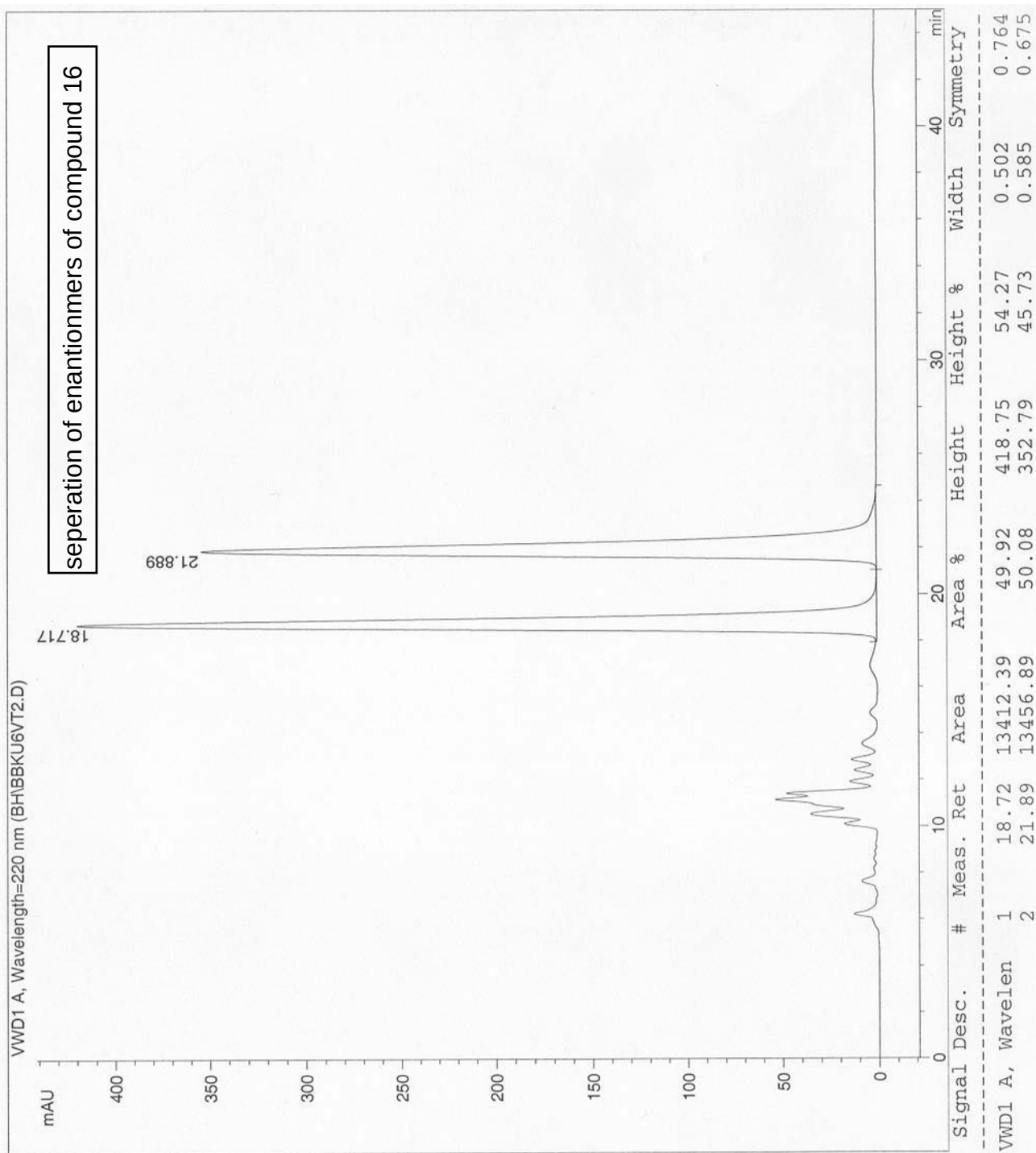
product KU6B100 after purification with HPLC

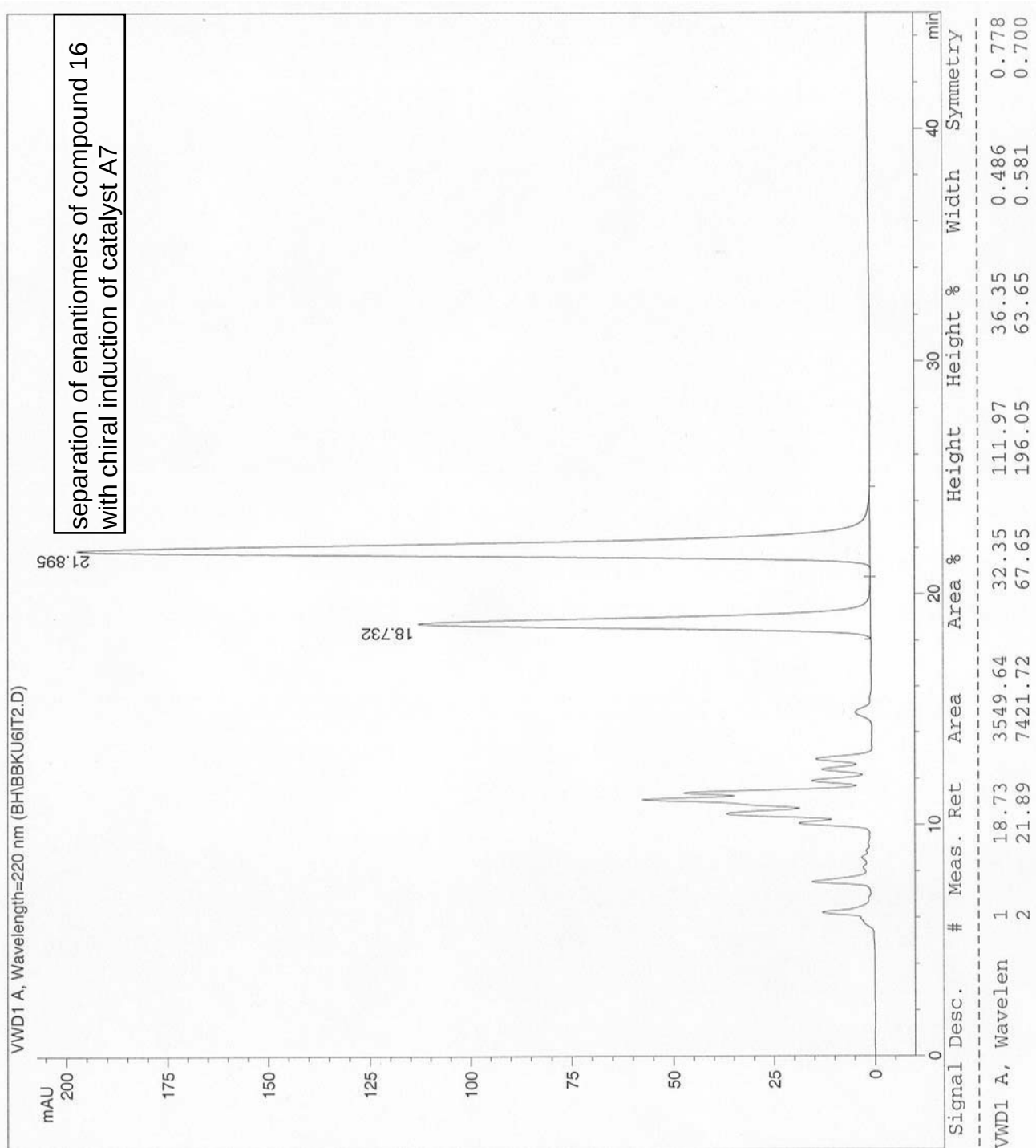
MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

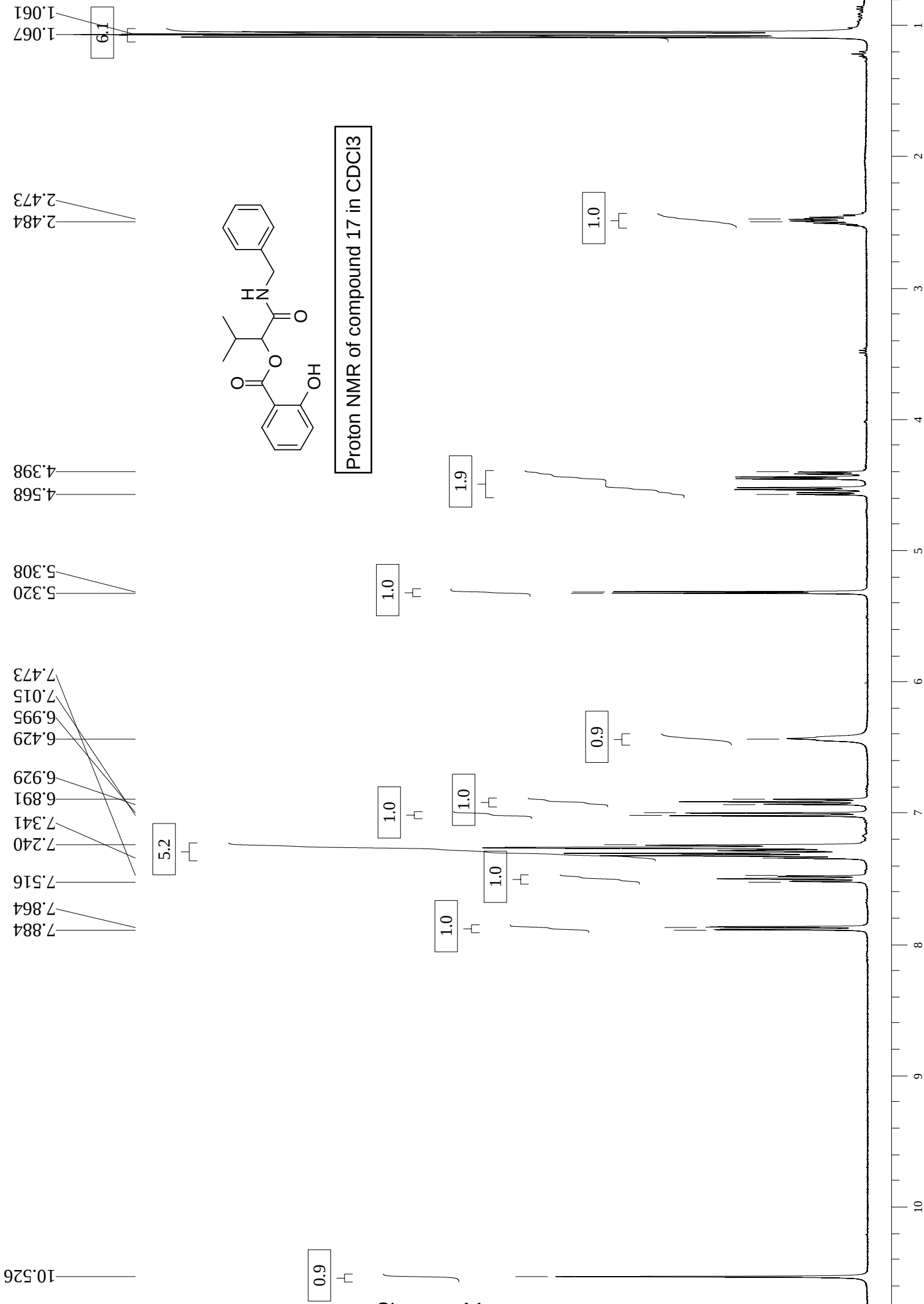
Retention Time (MS)	MS Area	Mol. Weight or Ion
3.699	6540607	486.90 I
		335.30 I
		334.30 I
		312.30 I
		205.20 I

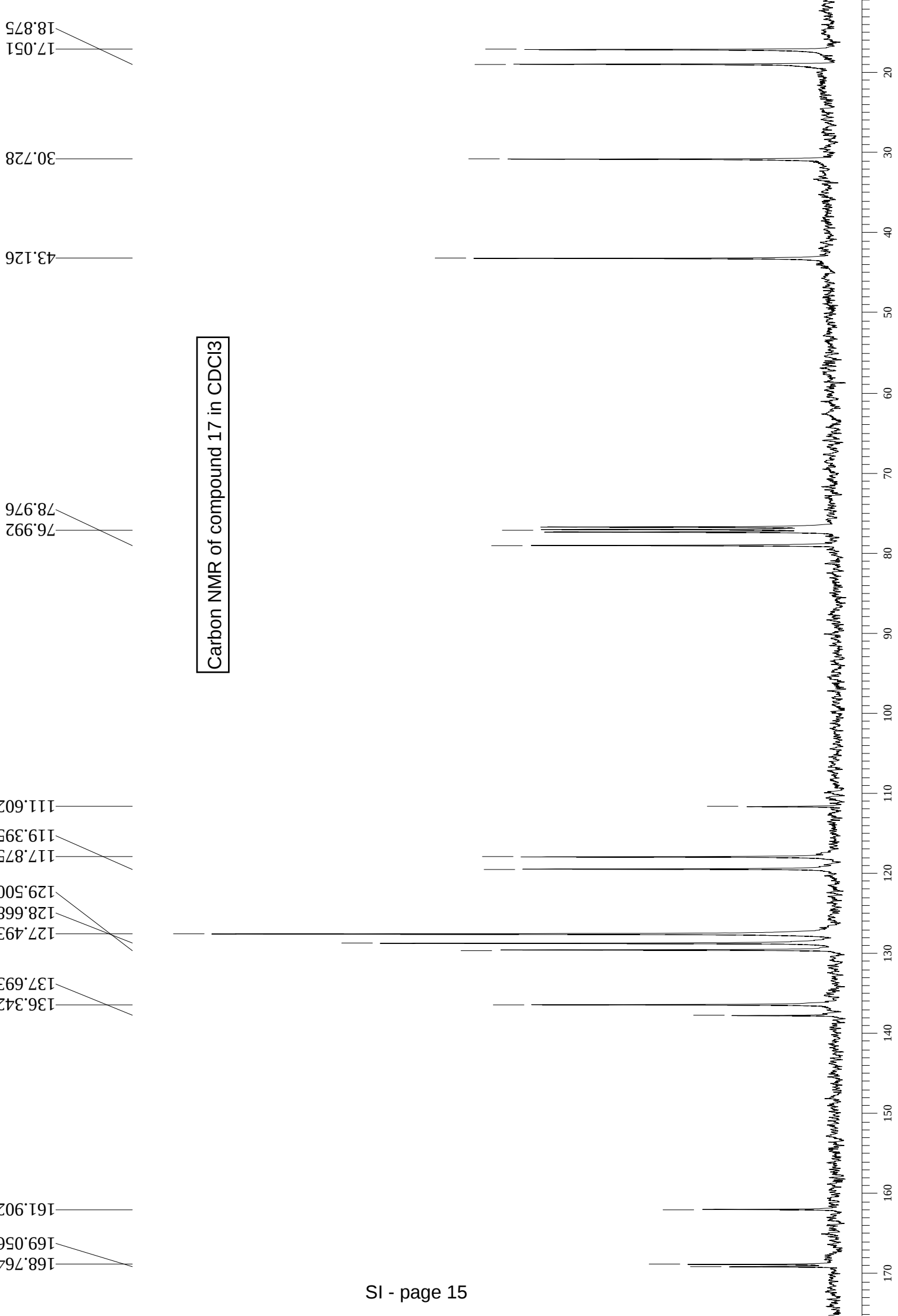


*** End of Report ***









=====

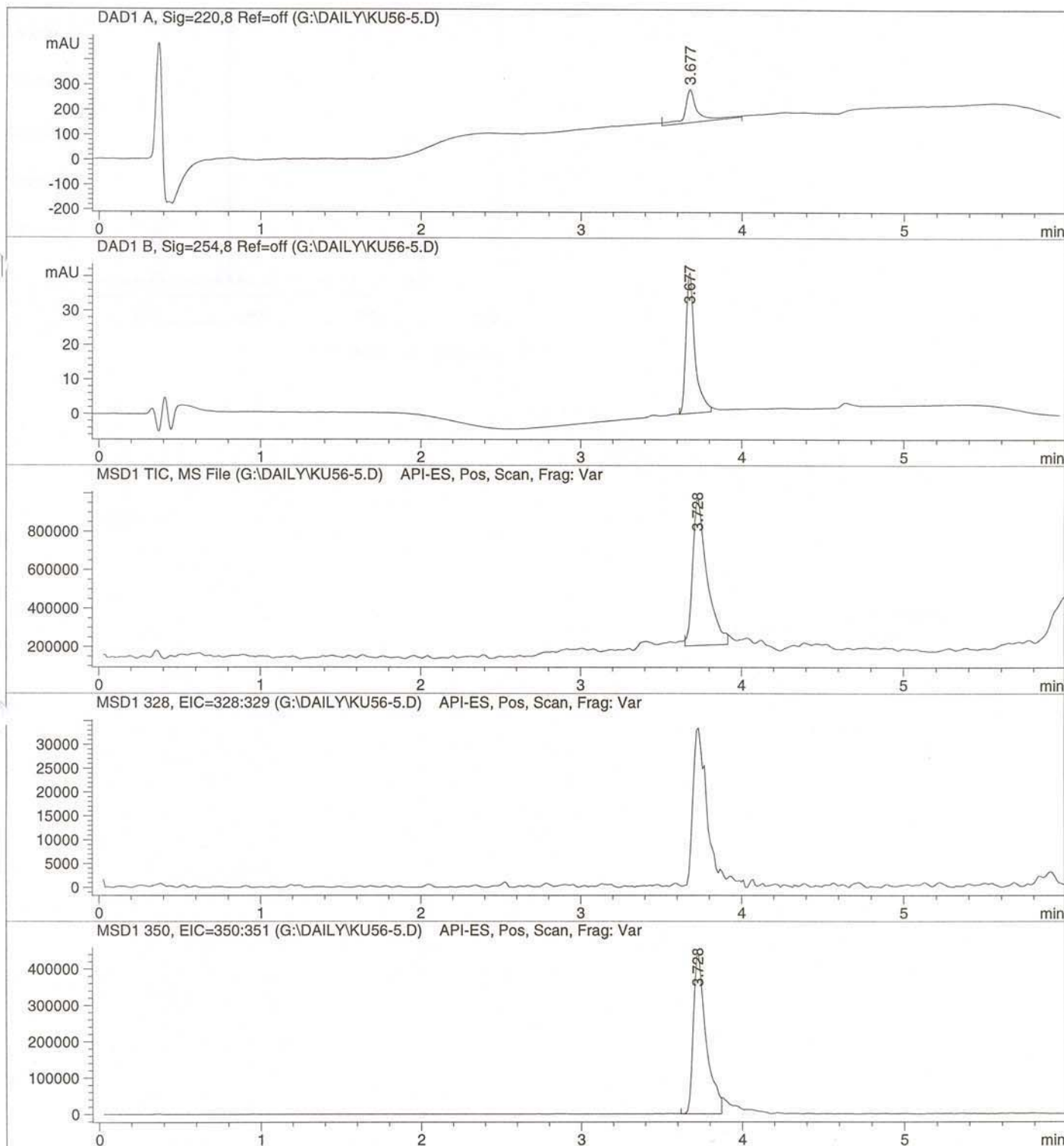
Injection Date : 10/31/02 4:55:09 AM
Sample Name : KU56-5
Acq. Operator : user

Seq. Line : 150
Location : Vial 94
Inj : 1
Inj Volume : 5 µl

Acq. Method : D:\HPCHEM\1\METHODS\ESI_LC6.M
Last changed : 10/31/02 4:53:12 AM by user
(modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\ESI_LC6.M
Last changed : 10/24/02 2:01:04 PM
(modified after loading)

YMC ODS-A 5cm 2µ
ESI positiv

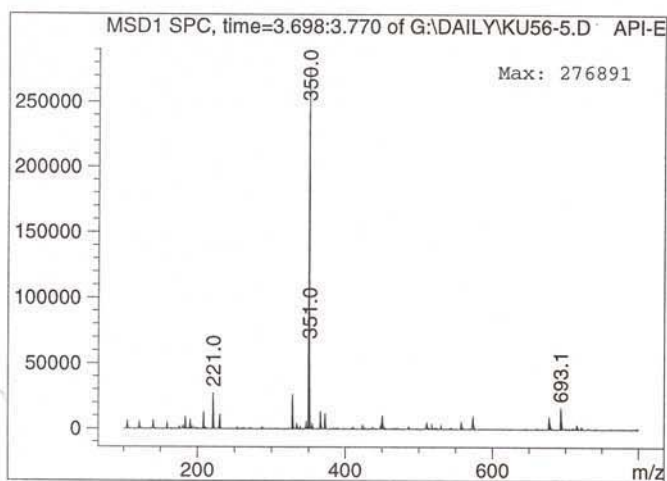
HPLC-MS Spectrum of compound 17



*KU56 - hplc-ms of product after
purification with hplc*

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
3.728	4747170	351.00 I
		350.00 I



*** End of Report ***

Standard Report

Data File name: I:\ULRIKE\HPLC4\300702\KU19R5-3.D
 Sample name: KU19r5
 Act. Inj. Vol.: Actual Injection Volume same as Method

Acquis. Meth.: POS1_E7.M

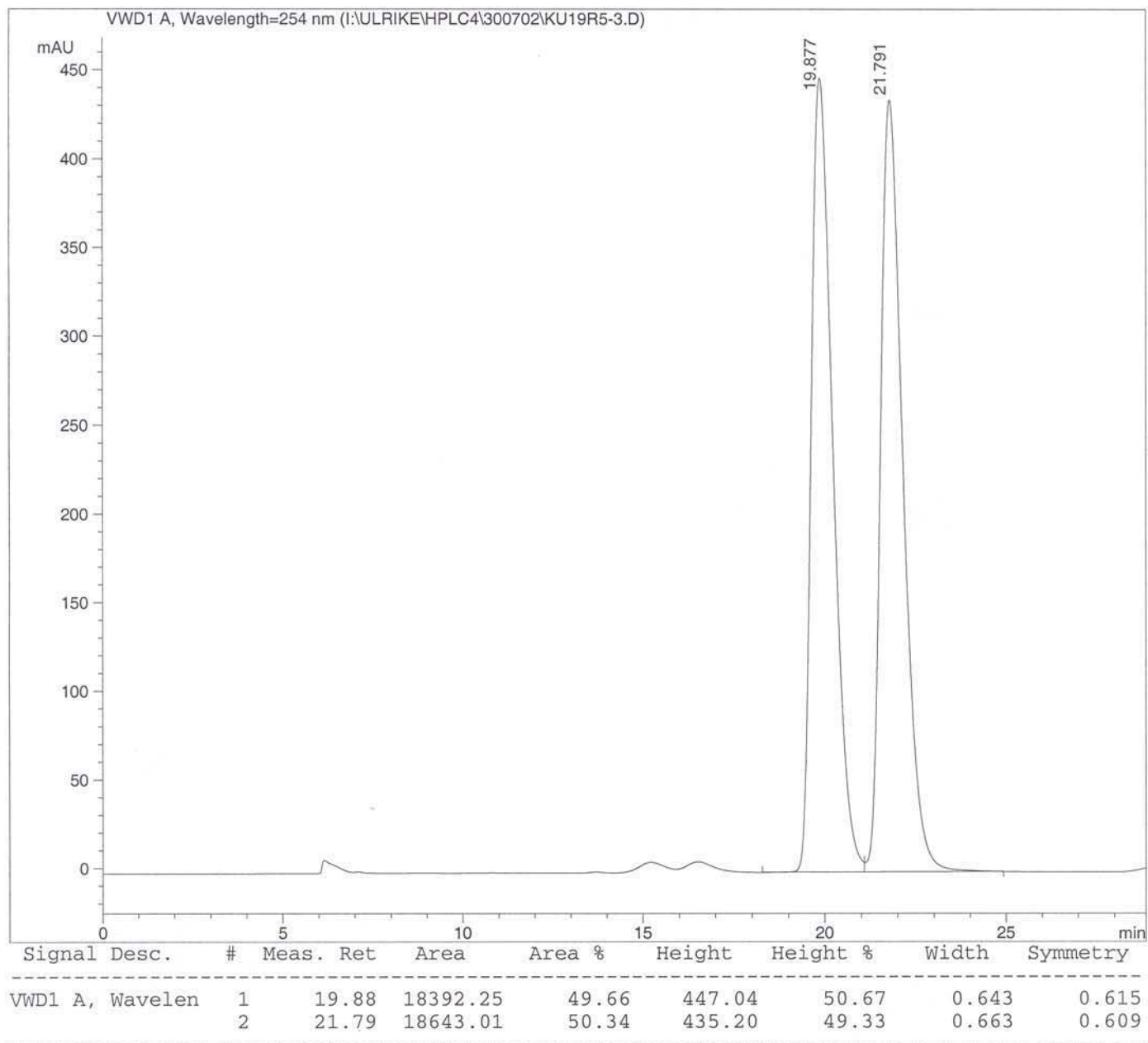
Integr. Method: D:\HPCHEM\1\METHODS\POS2_E13.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:
 Position 2:
 Phenomenex Chirex R-NGLY & DNB
 4.6 x 50

Injection date: 7/30/02 3:47:48 PM
 Report date: 12/2/02 1:50:48 PM

data acquired by:
 vial: 75

separation of enantiomers of compound 17



ku 19 - separation of enantiomers

Standard Report

Data File name: D:\HPCHEM\1\DATA\311002\KU56-5.D

Sample name: KU56-5

Act. Inj. Vol.: 15.00

uL

Acquis. Meth.: POS1_7.M

Integr. Method: D:\HPCHEM\1\METHODS\POS1_7.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 10/31/02

12:52:29 PM

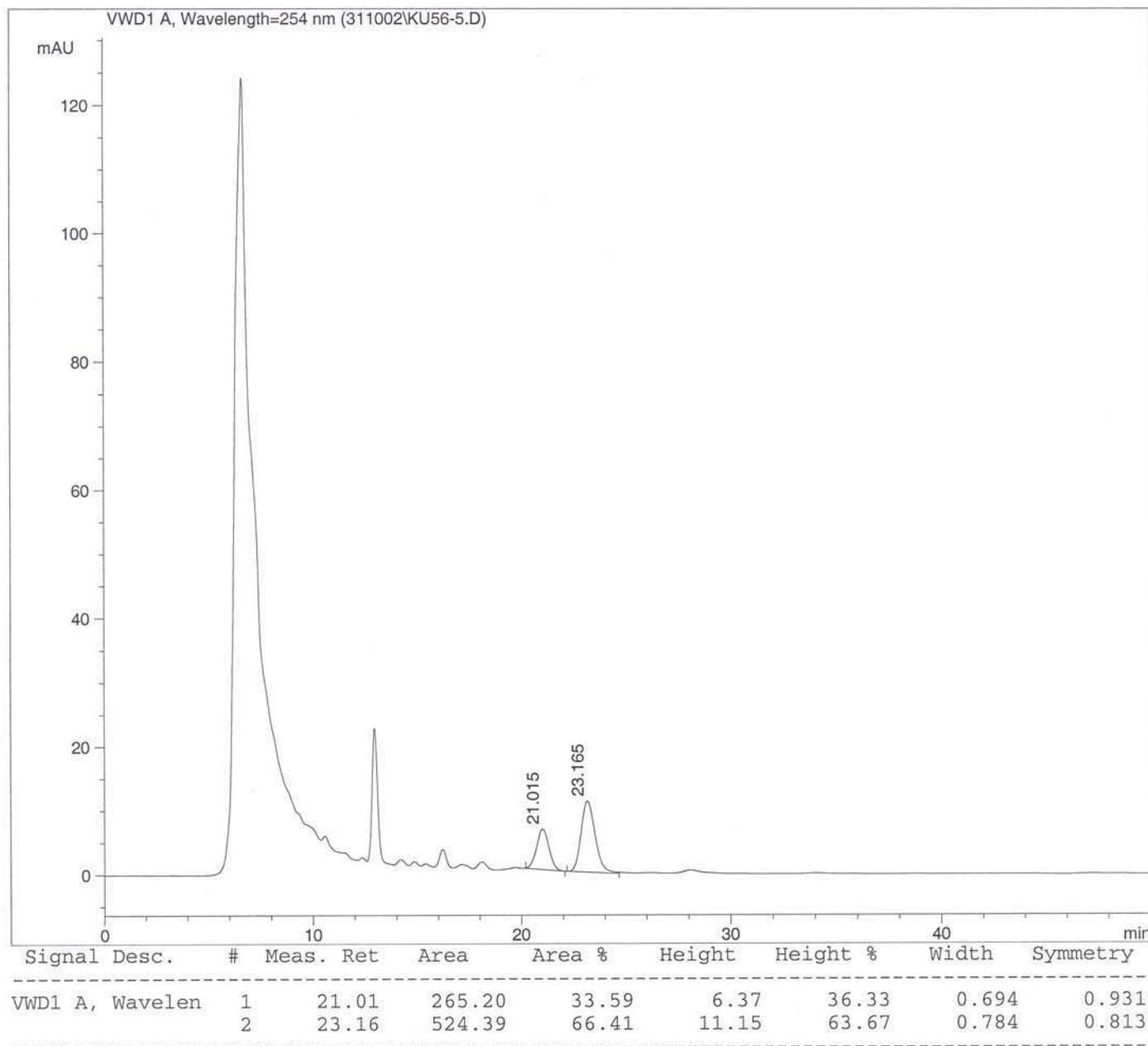
Report date: 10/31/02

1:46:05 PM

data acquired by:

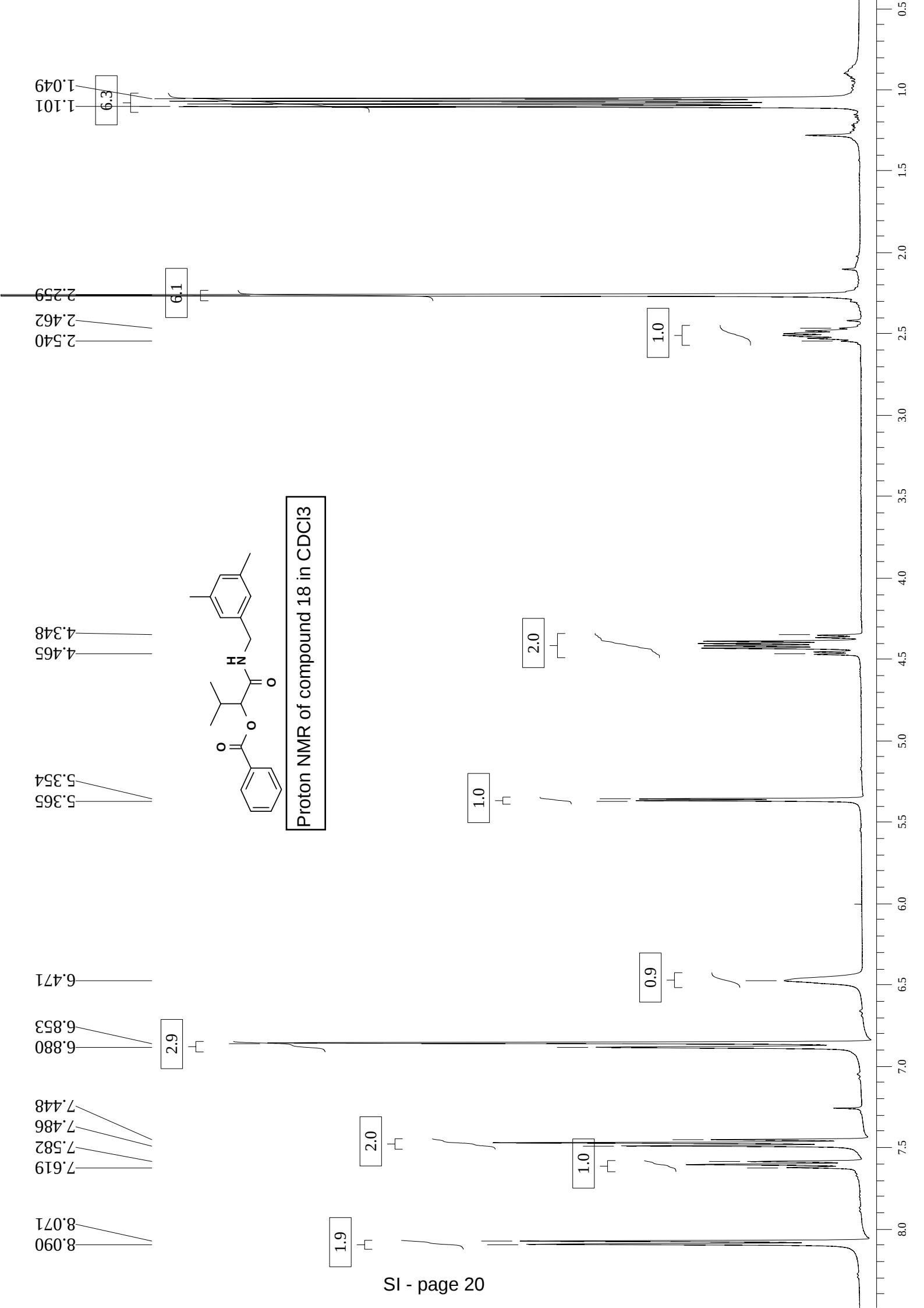
vial: 71

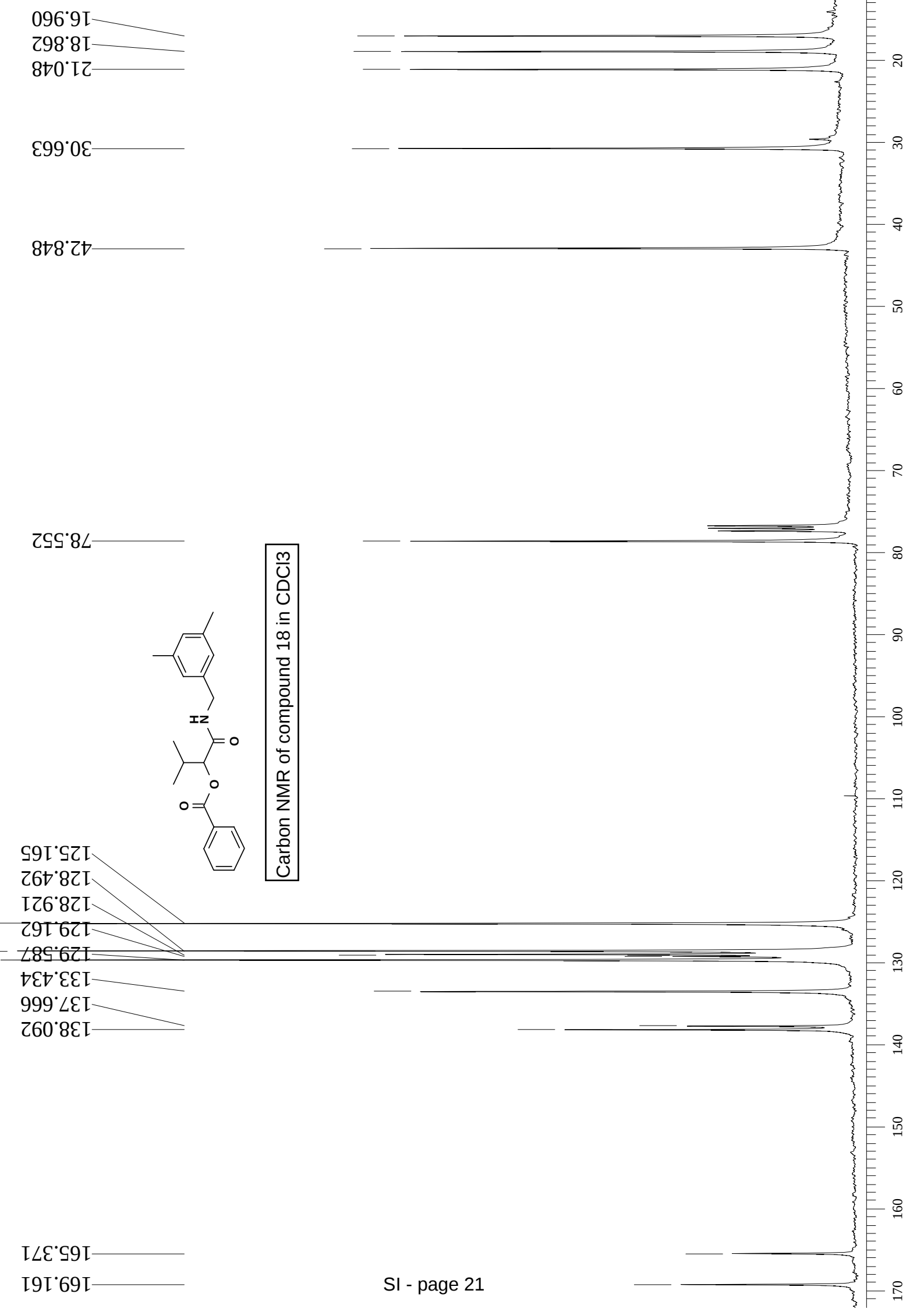
separation of enantiomers of compound 17
with chiral induction of catalyst A7



chiral HPLC of KU56 → 32:1 ee

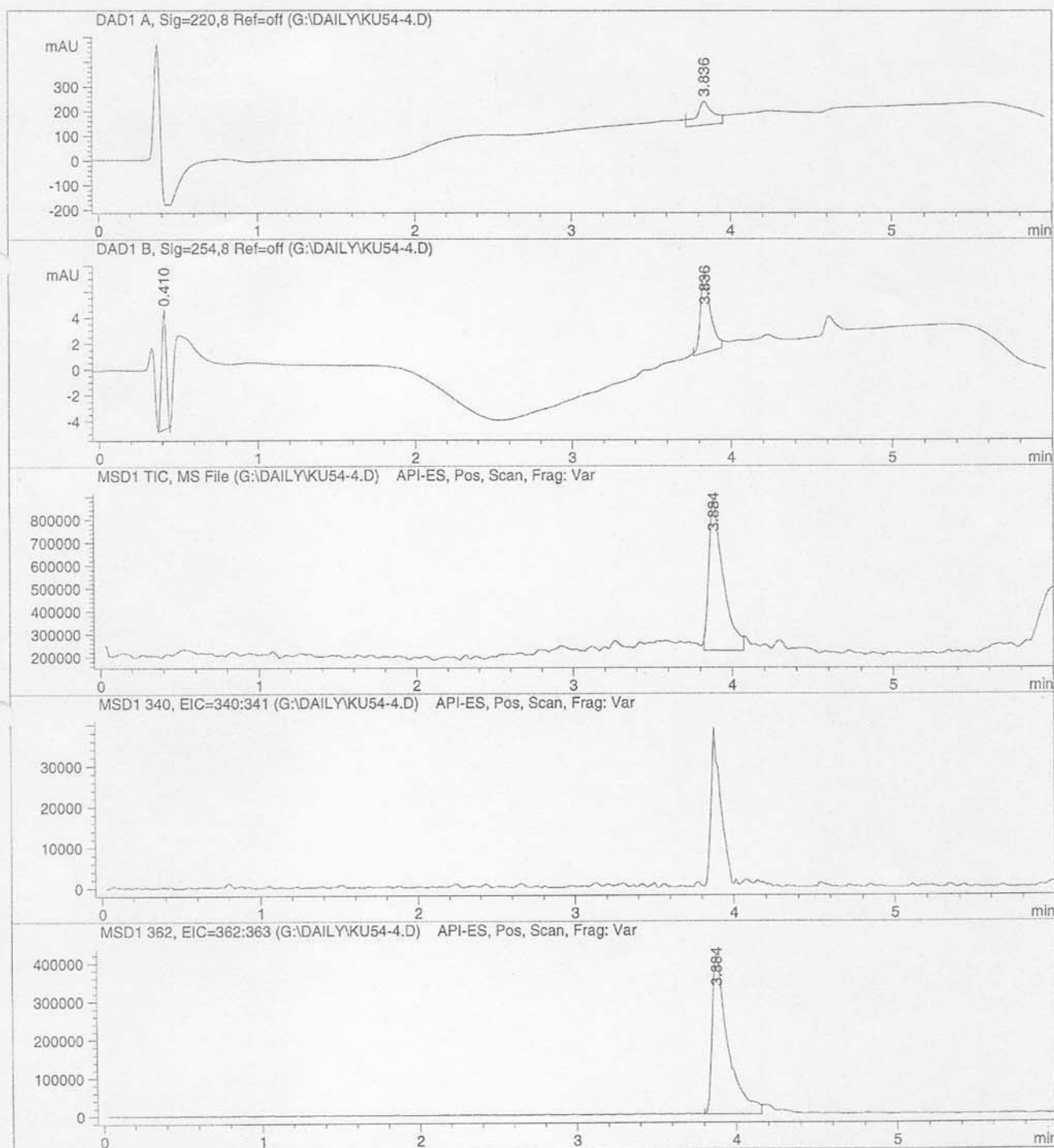
(enantiomers have been proved by co-injection
with KU19)





=====

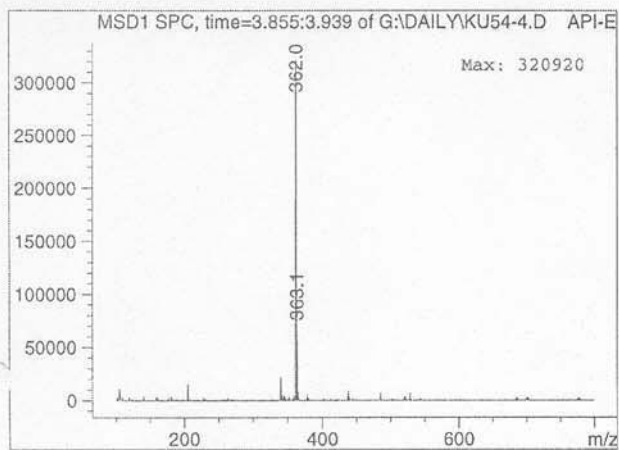
Injection Date	: 10/29/02 4:40:52 PM	Seq. Line	: 33
Sample Name	: KU54-4_RK	Location	: Vial 2
Acq. Operator	: user	Inj	: 1
		Inj Volume	: 5 µl
Acq. Method	: D:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 10/29/02 4:39:01 PM by user (modified after loading)		
Analysis Method	: C:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 10/24/02 2:01:04 PM (modified after loading)		
YMC ODS-A 5cm 2µ			
ESI positiv			

HPLC-MS Spectrum of compound 18

KU54 hplc-ms of product after purification with hplc

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
3.884	4135192	363.10 I
		362.05 I



*** End of Report ***

Standard Report

Data File name: D:\HPCHEM\1\DATA\201102\KU47-13.D
 Sample name: KU47-13
 Act. Inj. Vol.: 20.00

uL

Acquis. Meth.: POS1_3.M

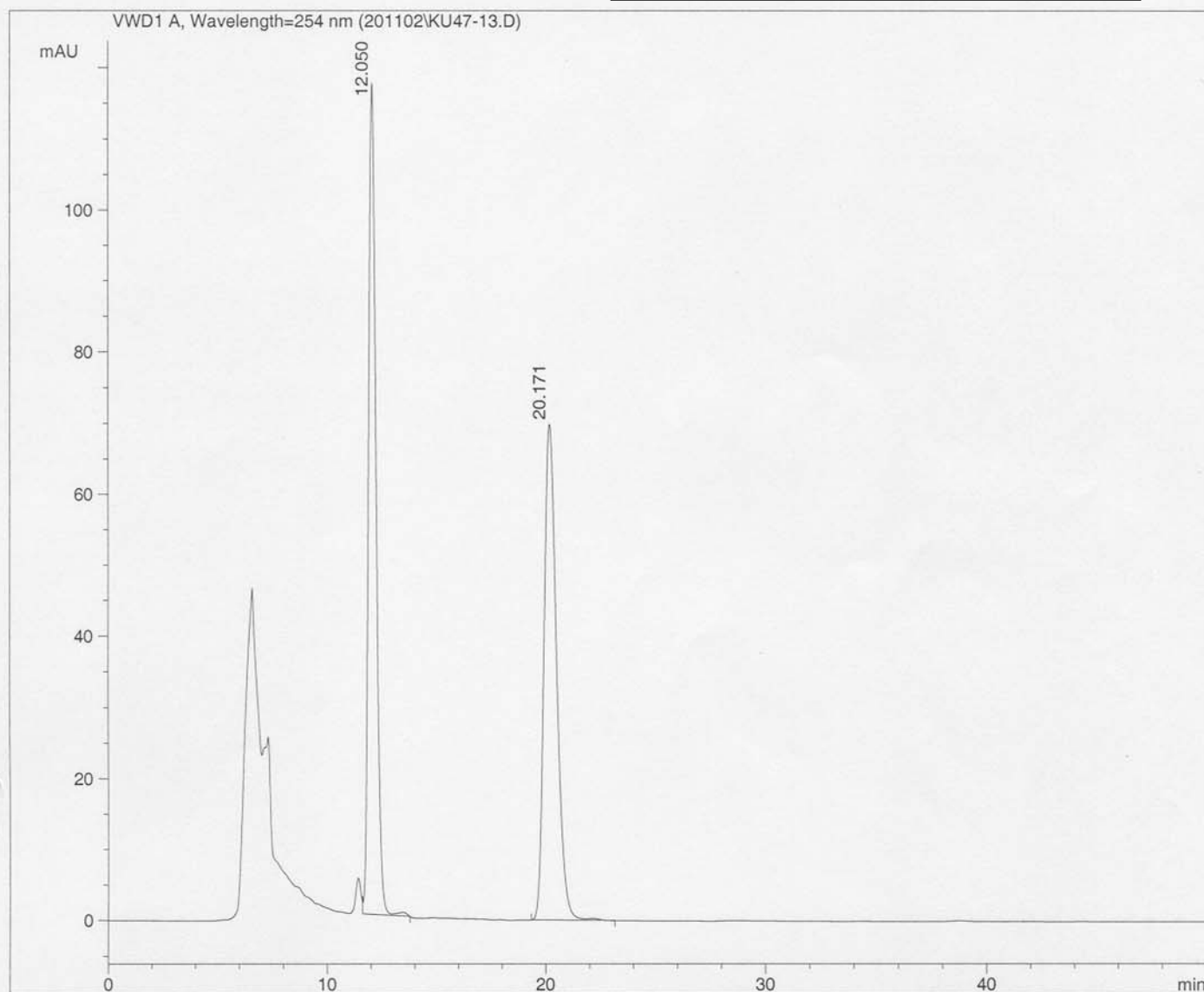
Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:
 Position 1:
 Daicel Chiralcel OD-H
 4.6 x 250

Injection date: 11/20/02 11:17:40 AM
 Report date: 11/20/02 12:18:18 PM

data acquired by:
 vial: 81

separation of enantiomers of compound 18



Signal Desc.	#	Meas. Ret	Area	Area %	Height	Height %	Width	Symmetry
VWD1 A, Wavelen	1	12.05	2785.56	49.90	116.87	62.61	0.377	0.764
	2	20.17	2796.68	50.10	69.78	37.39	0.619	0.726

KU47- fraction 13 after purification with chromatohon
 chiral hplc

Handwritten notes:
 Fraction 13 is KU47
 under 254nm light chromatohon

Standard Report

Data File name: D:\HPCHEM\1\DATA\291002\KU54-4W.D
 Sample name: KU54-4
 Act. Inj. Vol.: 40.00 uL

Acquis. Meth.: POS1_3.M

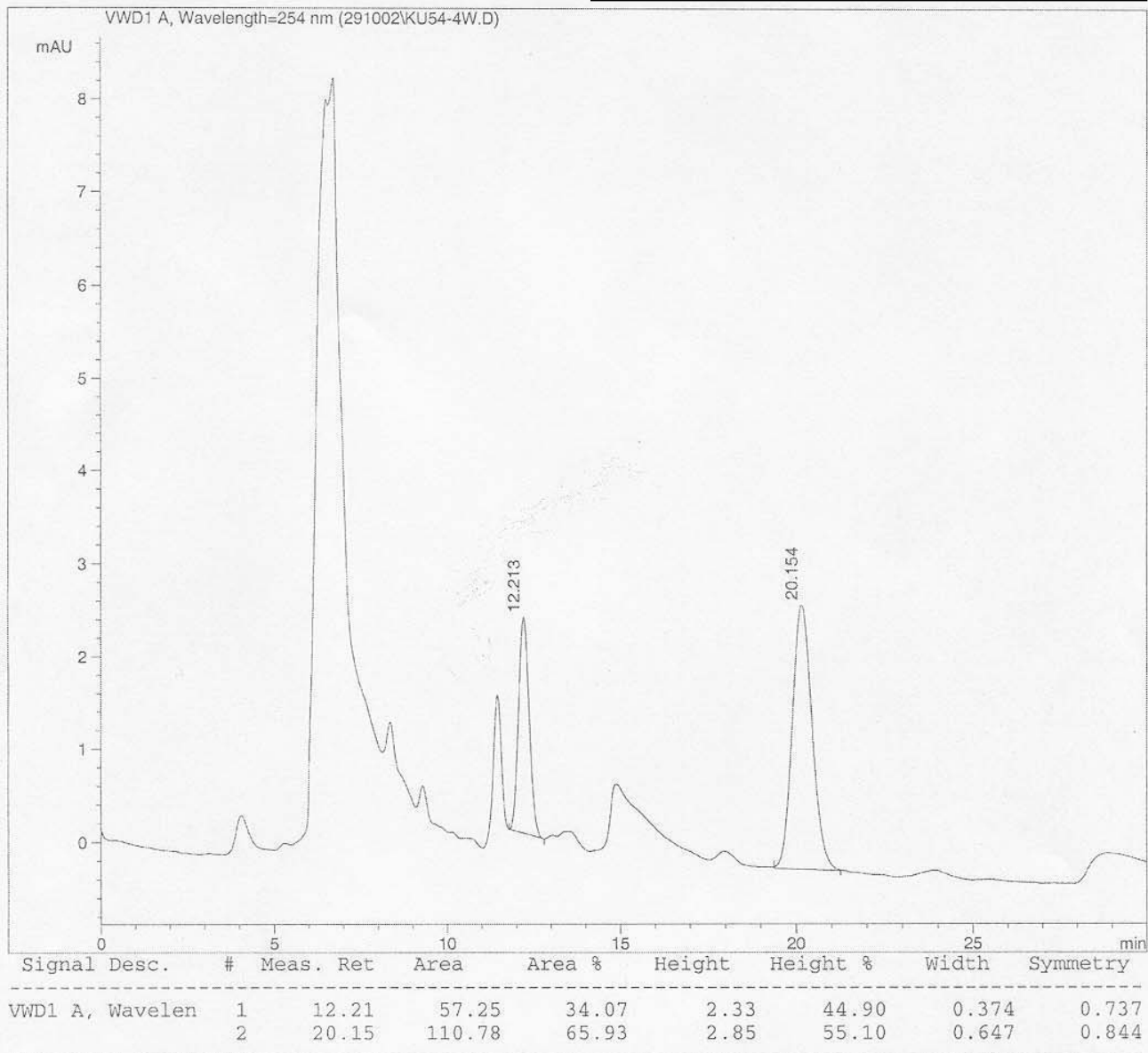
Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:
 Position 1:
 Daicel Chiralcel OD-H
 4.6 x 250

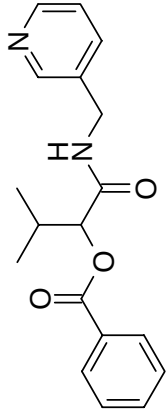
Injection date: 10/29/02 5:55:47 PM
 Report date: 10/30/02 10:10:39 AM

data acquired by:
 vial: 81

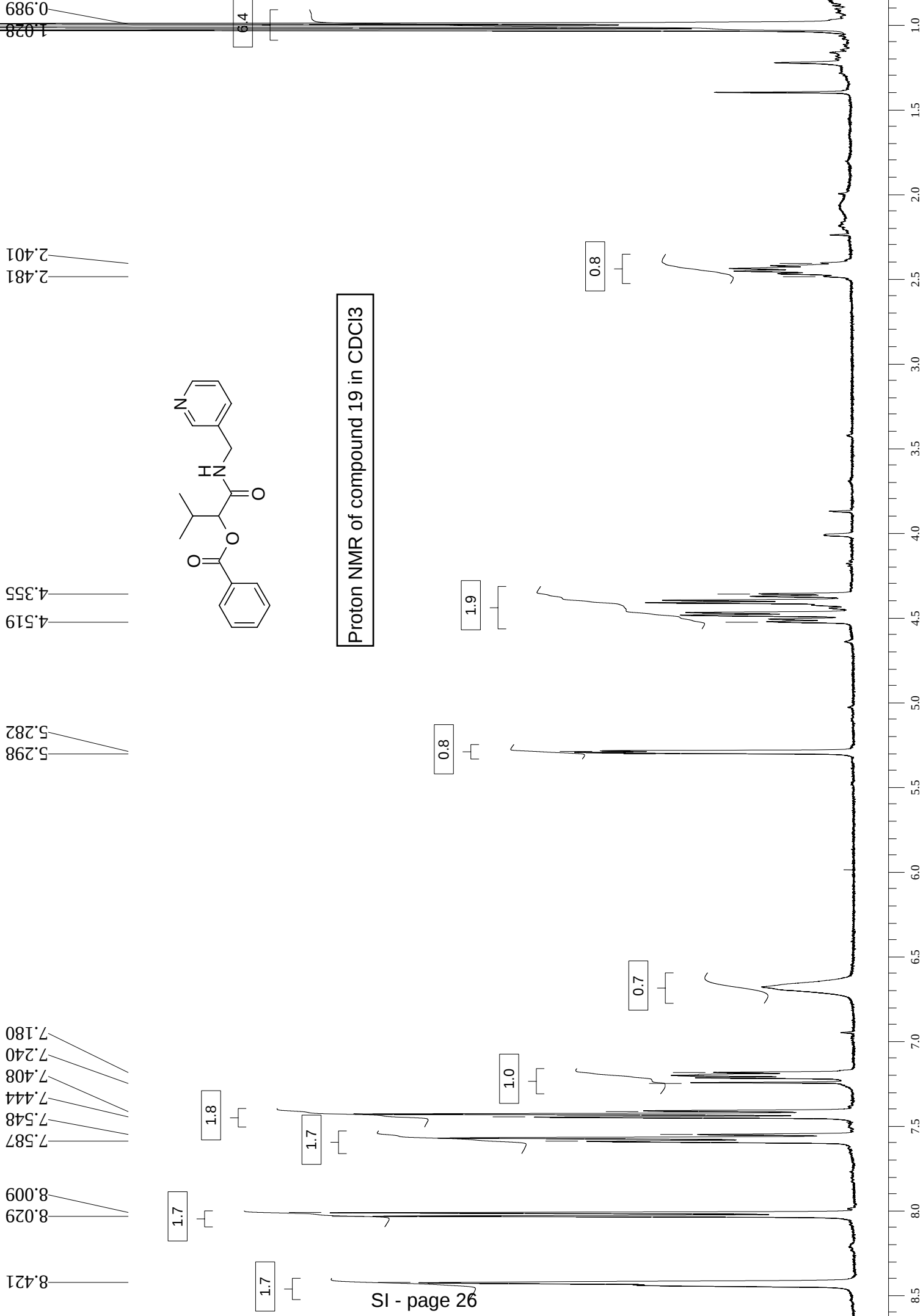
separation of enantiomers of compound 18
 with chiral induction of catalyst A7

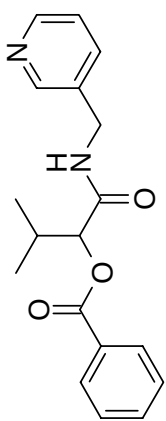


KU54 - chiral hplc → 32% ee
 peaks of enantiomers have been proved by co-injection
 with KU47

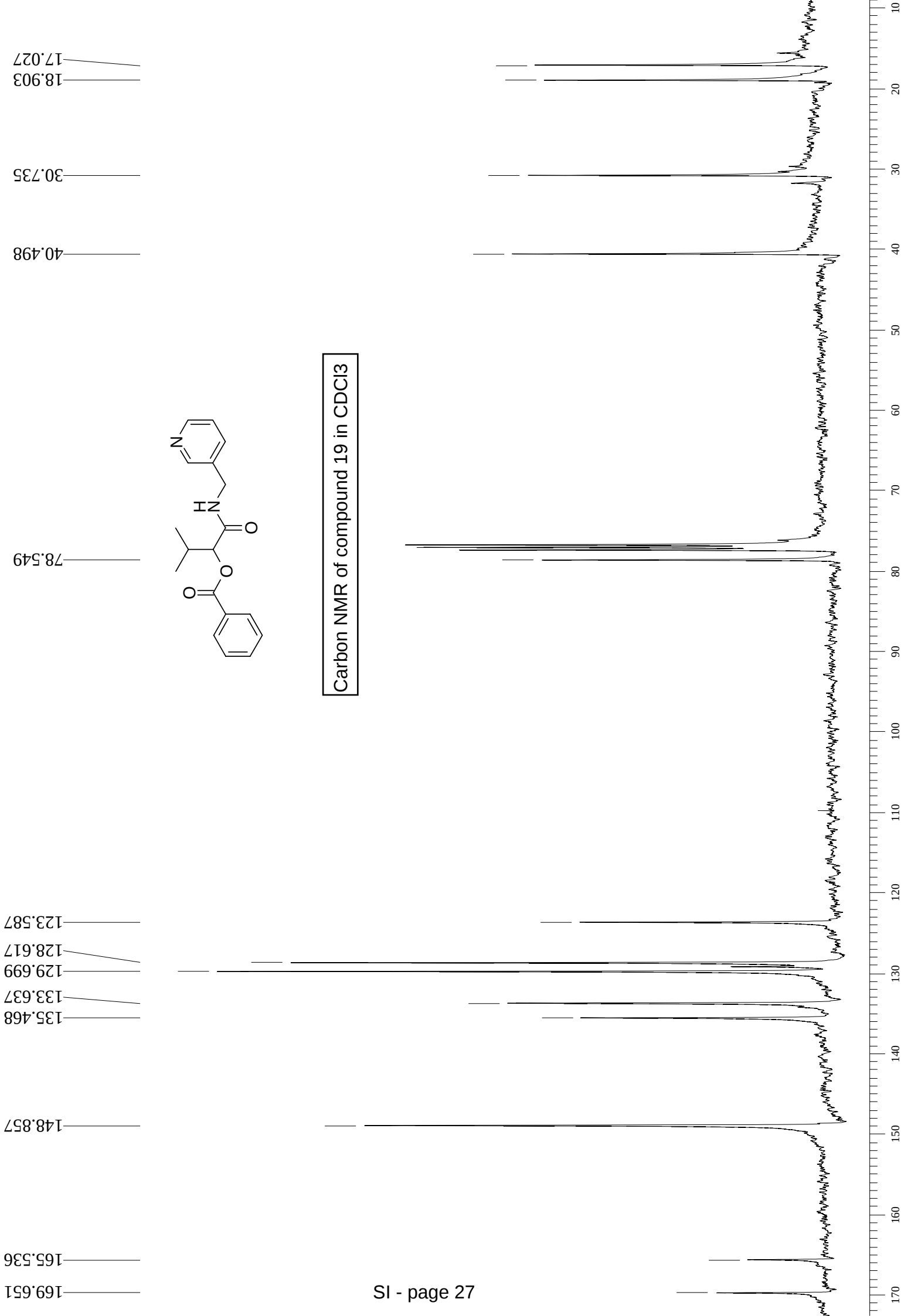


Proton NMR of compound 19 in CDCl₃



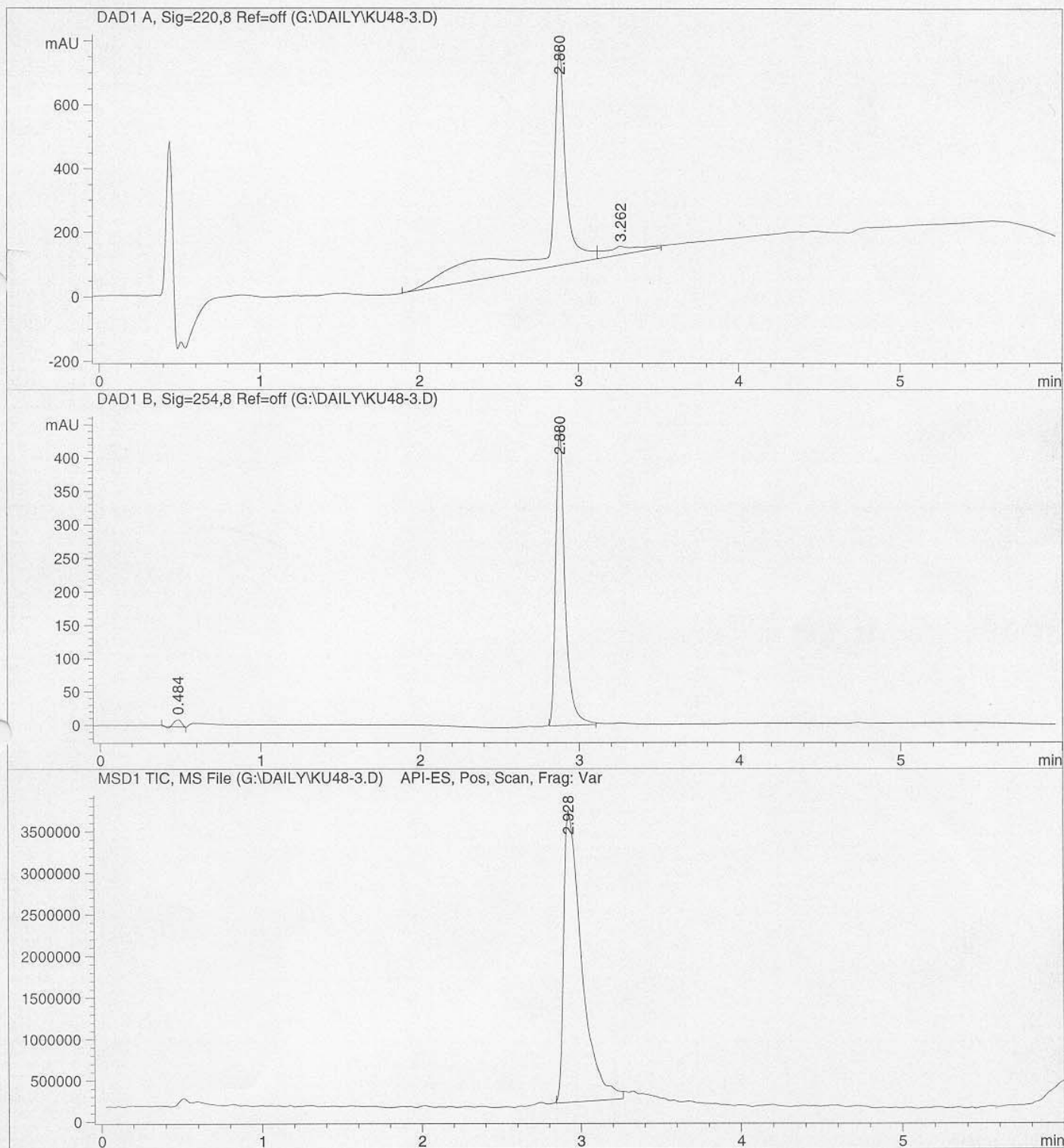


Carbon NMR of compound 19 in CDCl₃



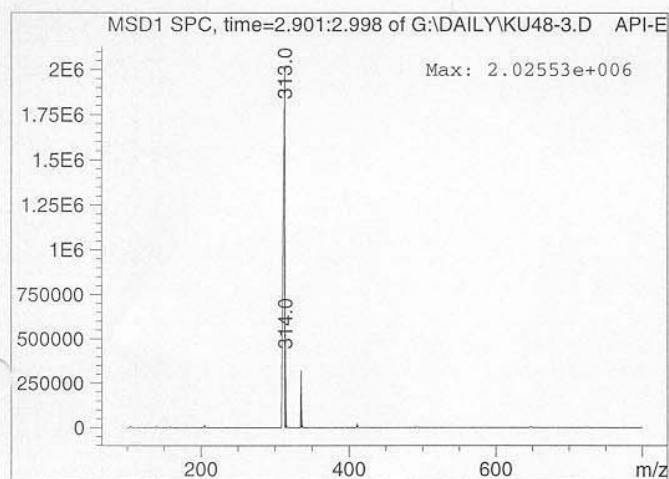
=====

Injection Date	: 11/21/02 3:54:24 PM	Seq. Line	: 53
Sample Name	: KU48-3-RK	Location	: Vial 2
Acq. Operator	: user	Inj	: 1
		Inj Volume	: 5 µl
Acq. Method	: D:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 11/21/02 3:51:31 PM by user (modified after loading)		
Analysis Method	: C:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 6/18/02 3:31:34 PM		
YMC ODS-A 5cm 2µ			
ESI positiv			

HPLC-MS Spectrum of compound 19

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
2.928	27623556	335.00 I
		314.00 I
		313.00 I



*** End of Report ***

Standard Report

Data File name: D:\HPCHEM\1\DATA\241002\KU48-2.D

Sample name: KU48

Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 10/24/02

10:44:25 AM

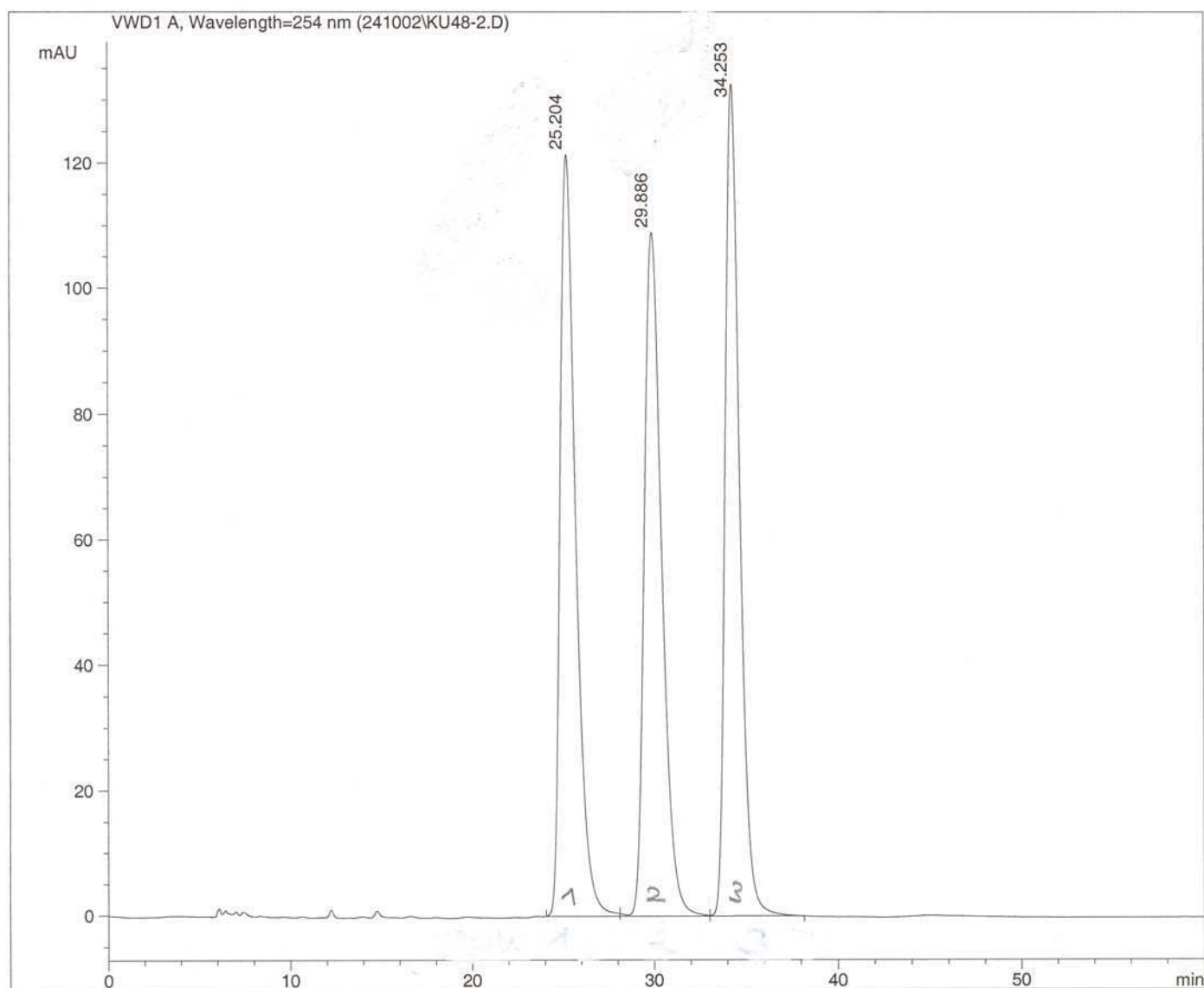
Report date: 10/24/02

11:45:37 AM

data acquired by:

vial: 21

separation of the enantiomers of compound 19



Signal Desc.	#	Meas. Ret	Area	Area %	Height	Height %	Width	Symmetry
VWD1 A, Wavelen	1	25.20	7265.04	33.32	121.53	33.46	0.913	0.614
	2	29.89	7270.34	33.35	109.03	30.02	1.028	0.708
	3	34.25	7267.83	33.33	132.65	36.52	0.843	0.730

ku48 separation of enantiomers
before purification
peak 1 and 2 are the product
→ see hplc-ms-chromatogram

Injection Date : 10/24/02 4:28:39 PM

Seq. Line : 12

Sample Name : KU48P1_RK

Location : Vial 1

Acq. Operator : user

Inj : 1

Inj Volume : 5 µl

Acq. Method : D:\HPCHEM\1\METHODS\ESI_LC6.M

Last changed : 10/24/02 4:25:27 PM by user

(modified after loading)

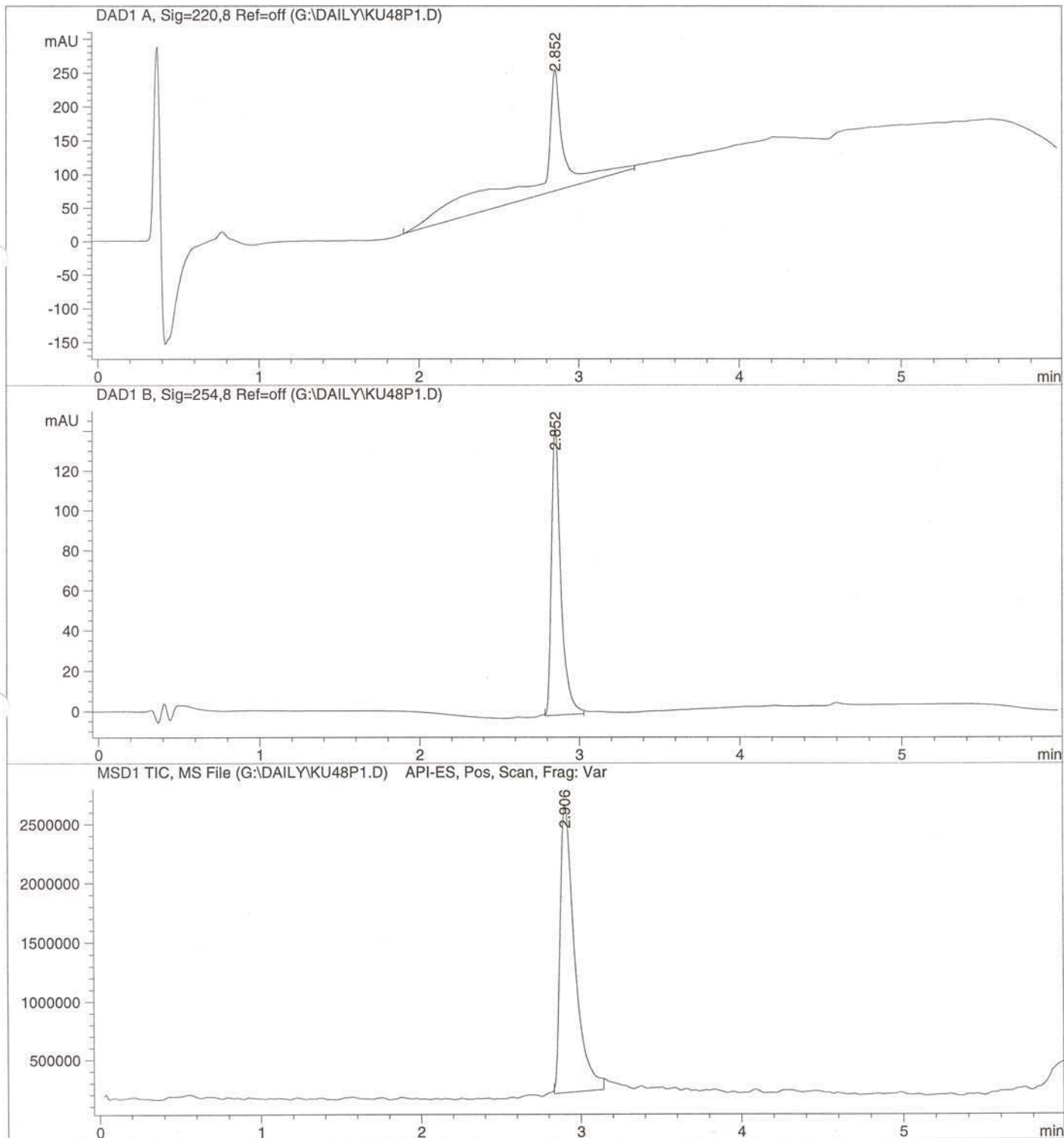
Analysis Method : C:\HPCHEM\1\METHODS\ESI_LC6.M

Last changed : 10/24/02 2:01:04 PM

(modified after loading)

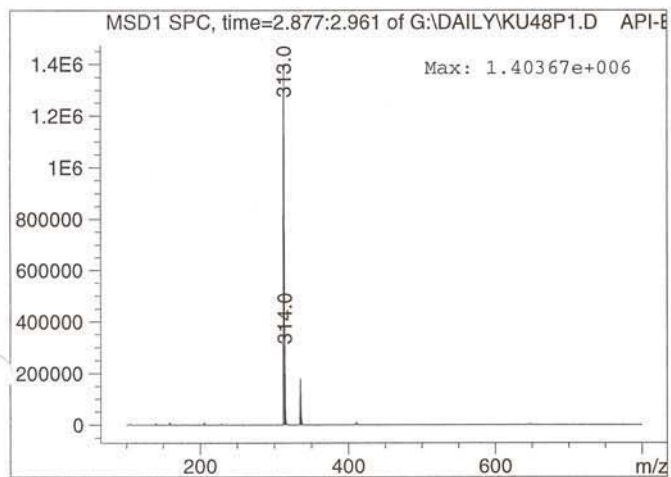
YMC ODS-A 5cm 2µ

ESI positiv

**HPLC-MS of peak 1 from separation of compound 19
on Chiracel OD-H***hplc-ms of enantiomere 1*

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
2.906	15603306	335.00 I
		314.00 I
		313.00 I



*** End of Report ***

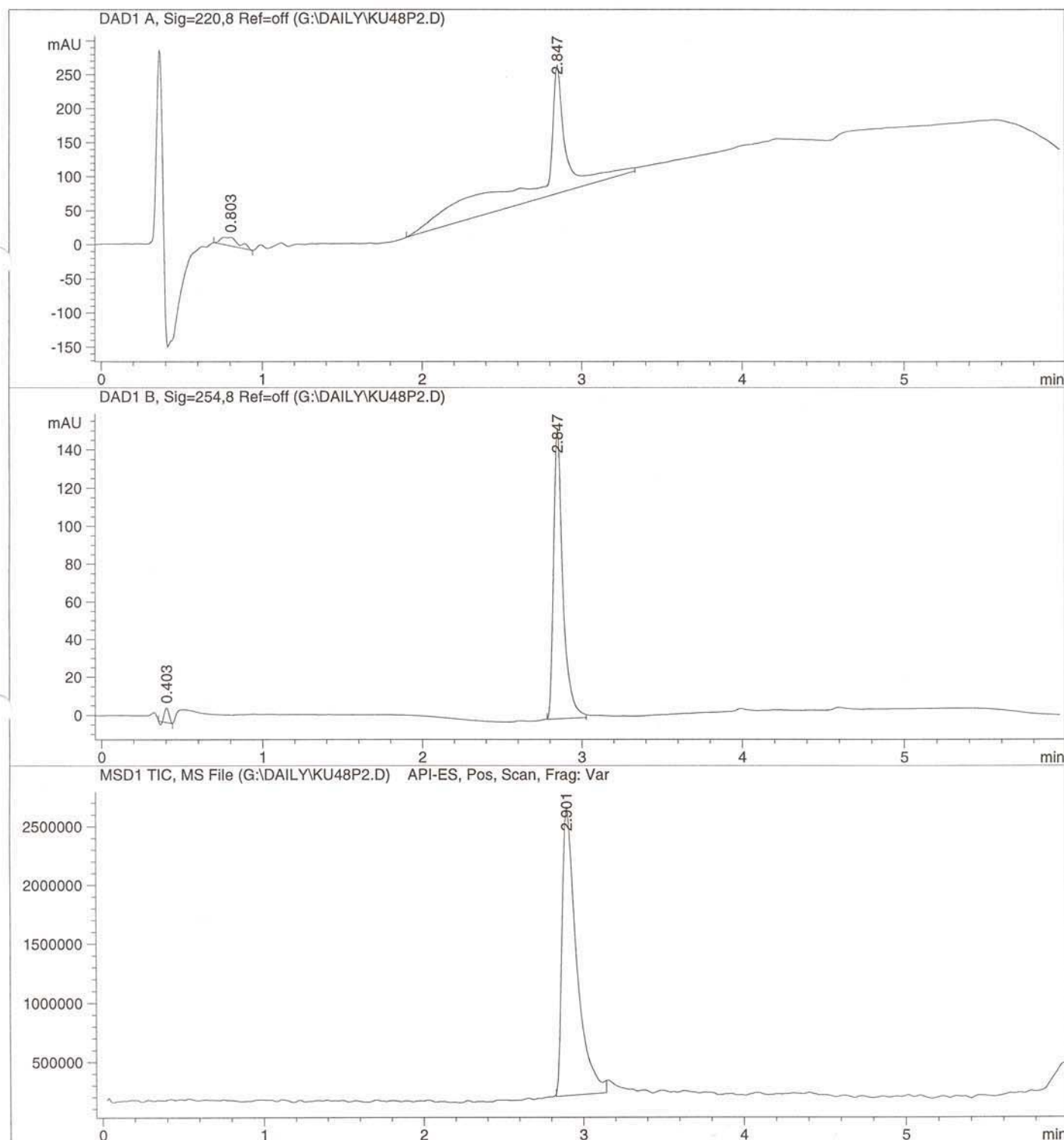
=====

Injection Date	: 10/24/02 4:37:04 PM	Seq. Line	: 13
Sample Name	: KU48P2_RK	Location	: Vial 2
Acq. Operator	: user	Inj	: 1
		Inj Volume	: 5 µl

Acq. Method : D:\HPCHEM\1\METHODS\ESI_LC6.M
 Last changed : 10/24/02 4:33:54 PM by user
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\ESI_LC6.M
 Last changed : 10/24/02 2:01:04 PM
 (modified after loading)

YMC ODS-A 5cm 2µ
 ESI positiv

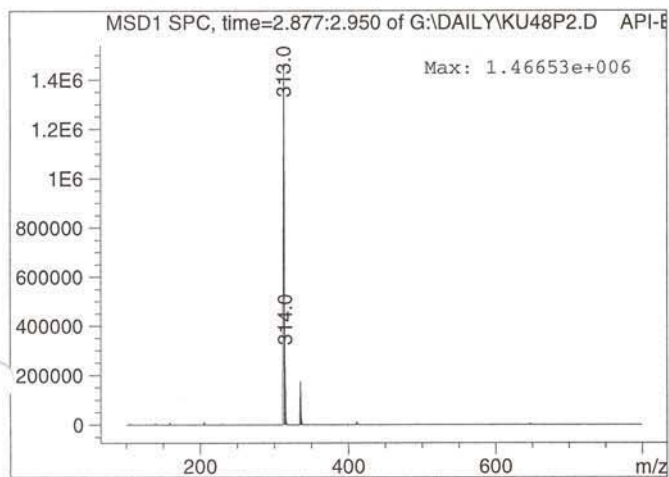
HPLC-MS of peak 2 from separation of compound 19
 on Chiracel OD-H



hplc -ms of enantiomers 2

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
2.901	15557765	335.00 I
		314.05 I
		313.05 I



*** End of Report ***

Standard Report

Data File name: D:\HPCHEM\1\DATA\061102\KU50F5.D

Sample name: KU50f5

Act. Inj. Vol.: 10.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 11/6/02

2:18:36 PM

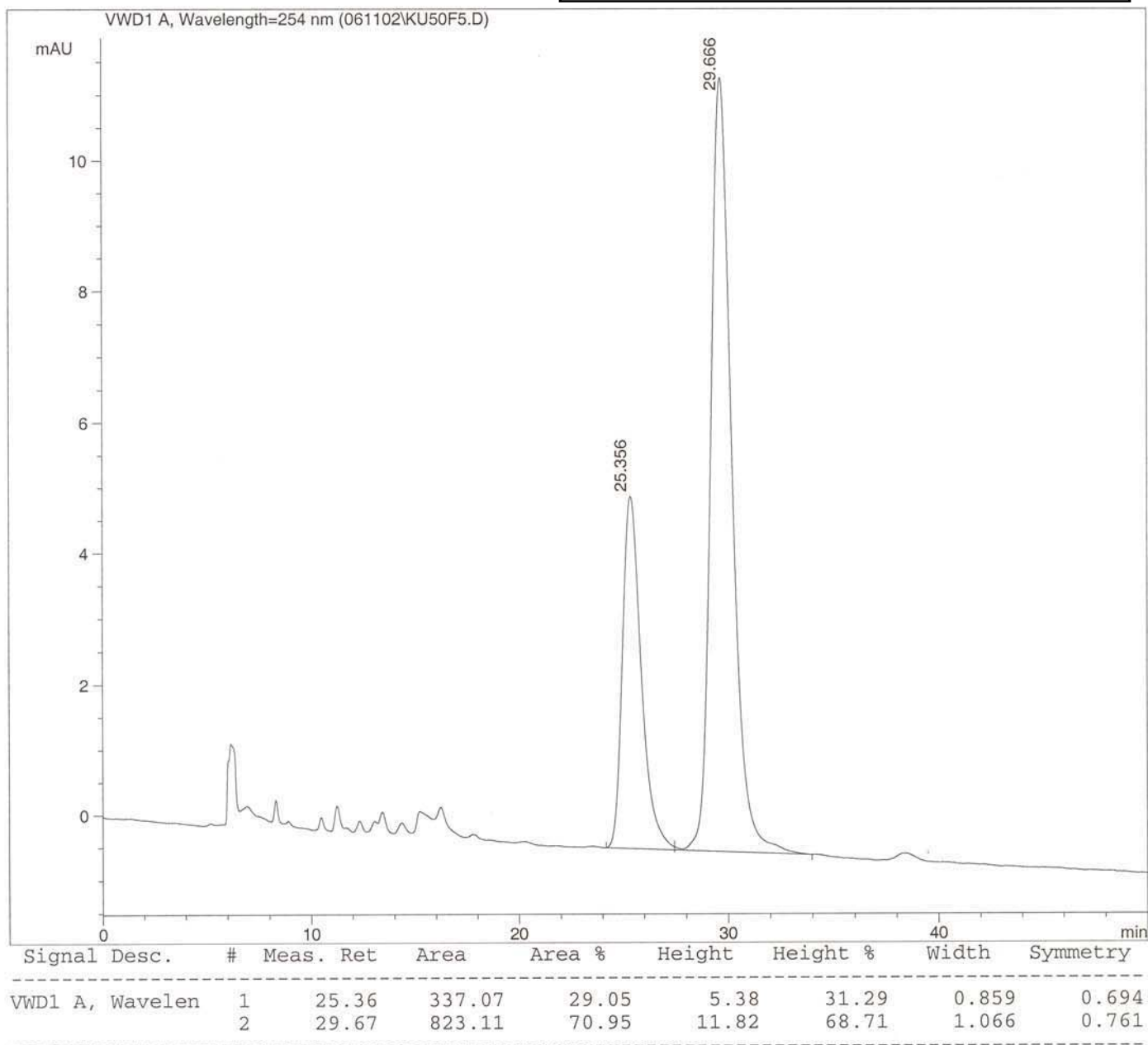
Report date: 11/6/02

3:10:47 PM

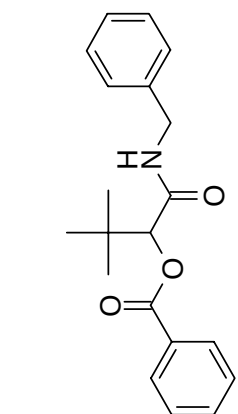
data acquired by:

vial: 81

separation of the enantiomers of compound 19
with chiral induction of catalyst A7



KU50 chiral hplc → 42% ee



Proton NMR of compound 20 in CDCl₃

4.321
4.467

5.035

6.204

7.157
7.240
7.378
7.416
7.511
7.548

7.992
8.010

9.5
1.072

4.7

1.9

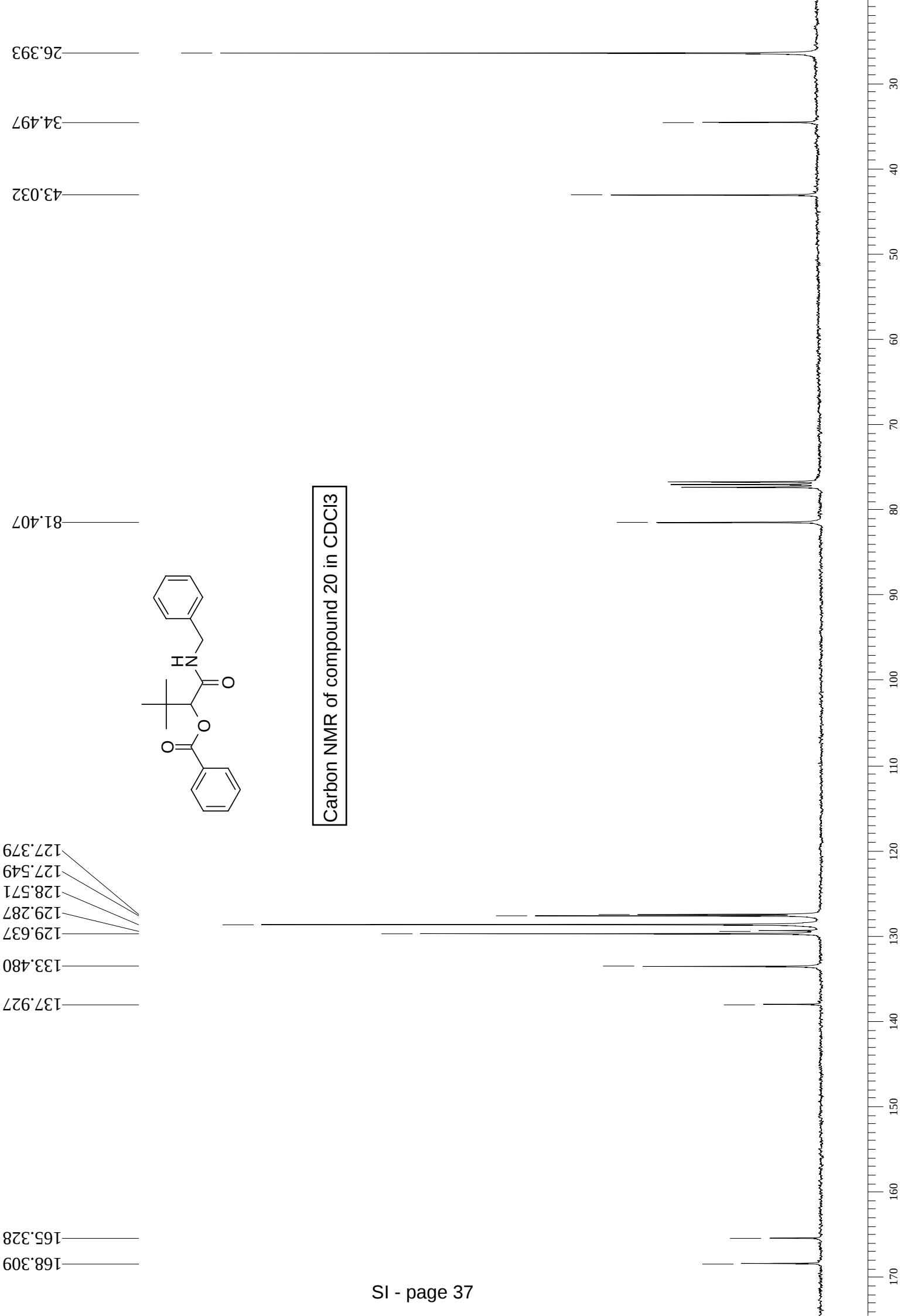
1.8

1.8

0.8

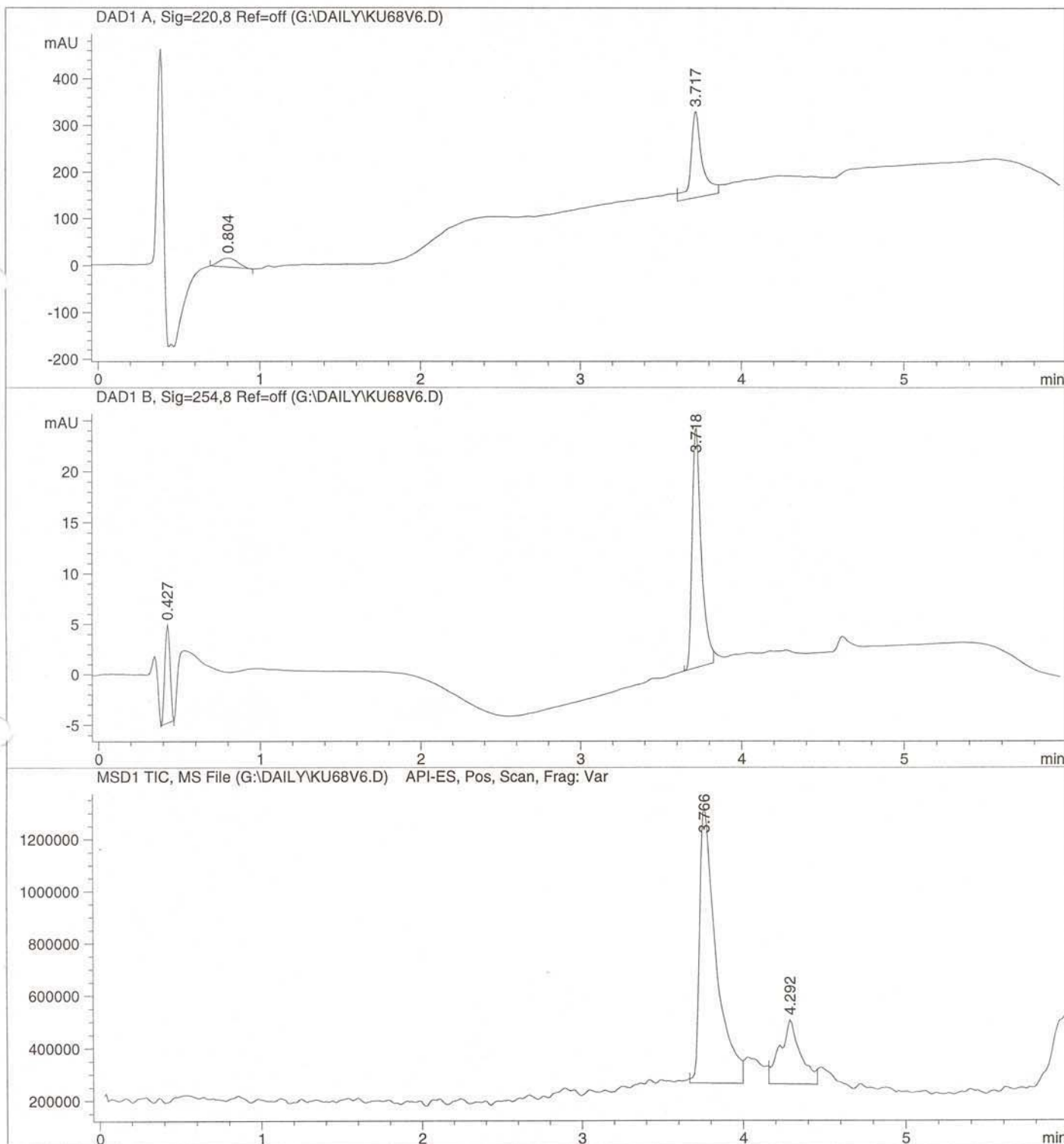
0.8

0.9



=====

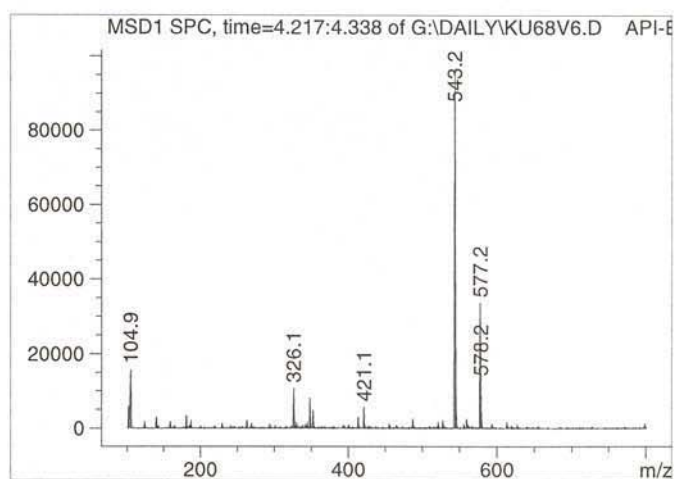
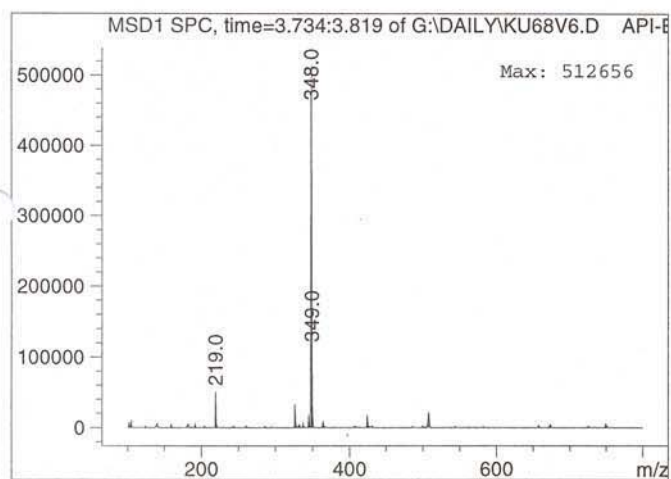
Injection Date	: 11/8/02 11:56:12 AM	Seq. Line	: 27
Sample Name	: KU68v6_RK	Location	: Vial 23
Acq. Operator	: user	Inj	: 1
		Inj Volume	: 5 µl
Acq. Method	: D:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 11/8/02 11:54:37 AM by user (modified after loading)		
Analysis Method	: C:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 6/18/02 3:31:34 PM		
YMC ODS-A 5cm 2µ			
ESI positiv			

HPLC-MS Spectrum of compound 20

hplc-ms of product after purification with hplc

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
3.766	7503952	349.00 I 348.00 I
4.292	2185946	578.20 I 577.15 I 544.20 I 543.20 I 326.10 I 104.95 I



*** End of Report ***

Standard Report

Data File name: D:\HPCHEM\1\DATA\051102\KU58.D

Sample name: KU58

Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 11/5/02

11:52:04 AM

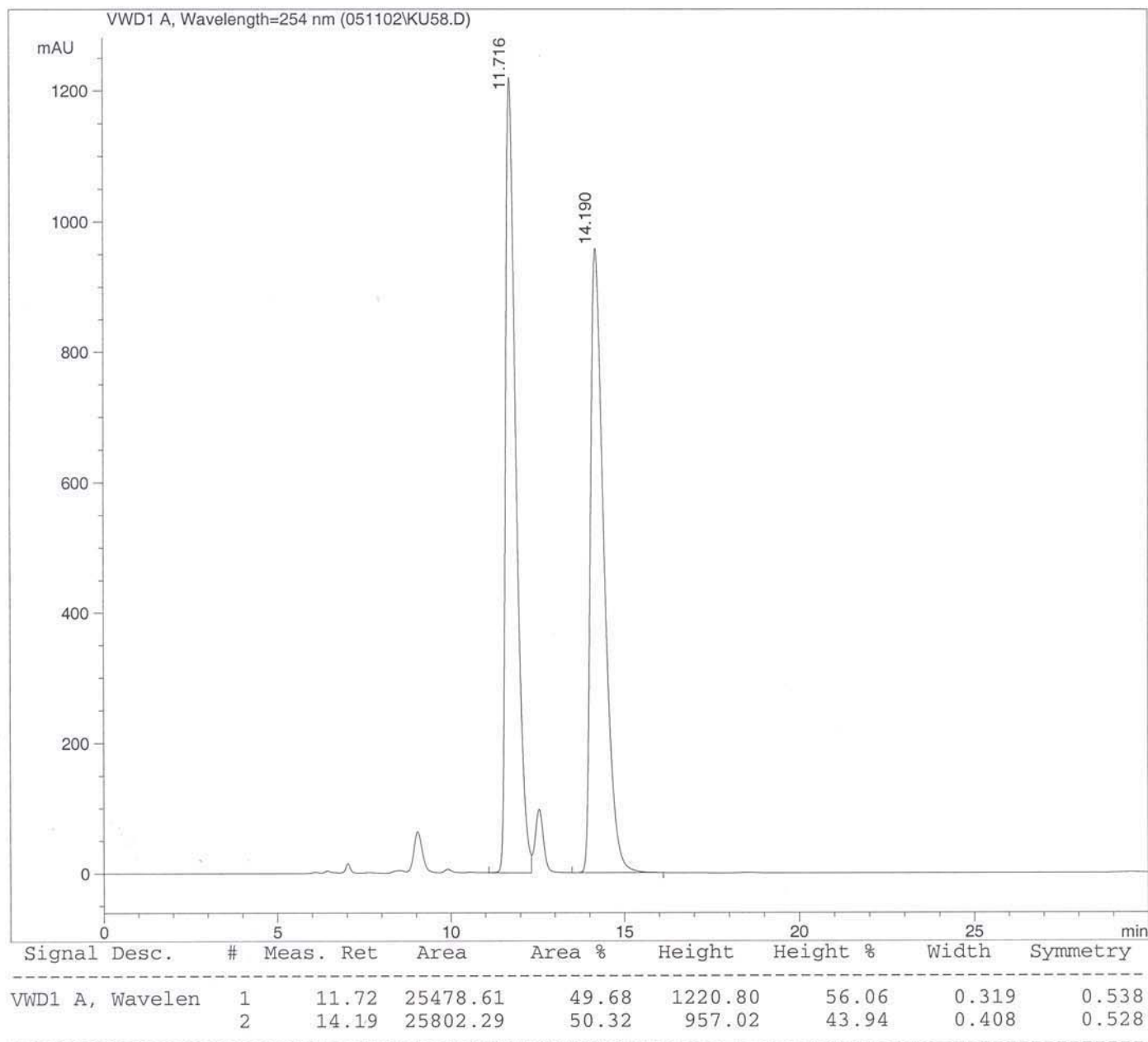
Report date: 11/5/02

12:36:53 PM

data acquired by:

vial: 71

separation of enantiomers of compound 20



KU58-separation of enantiomers

Standard Report

Data File name: D:\HPCHEM\1\DATA\081102\KU68V6.D

Sample name: KU68v6

Act. Inj. Vol.: 25.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 11/8/02

1:37:09 PM

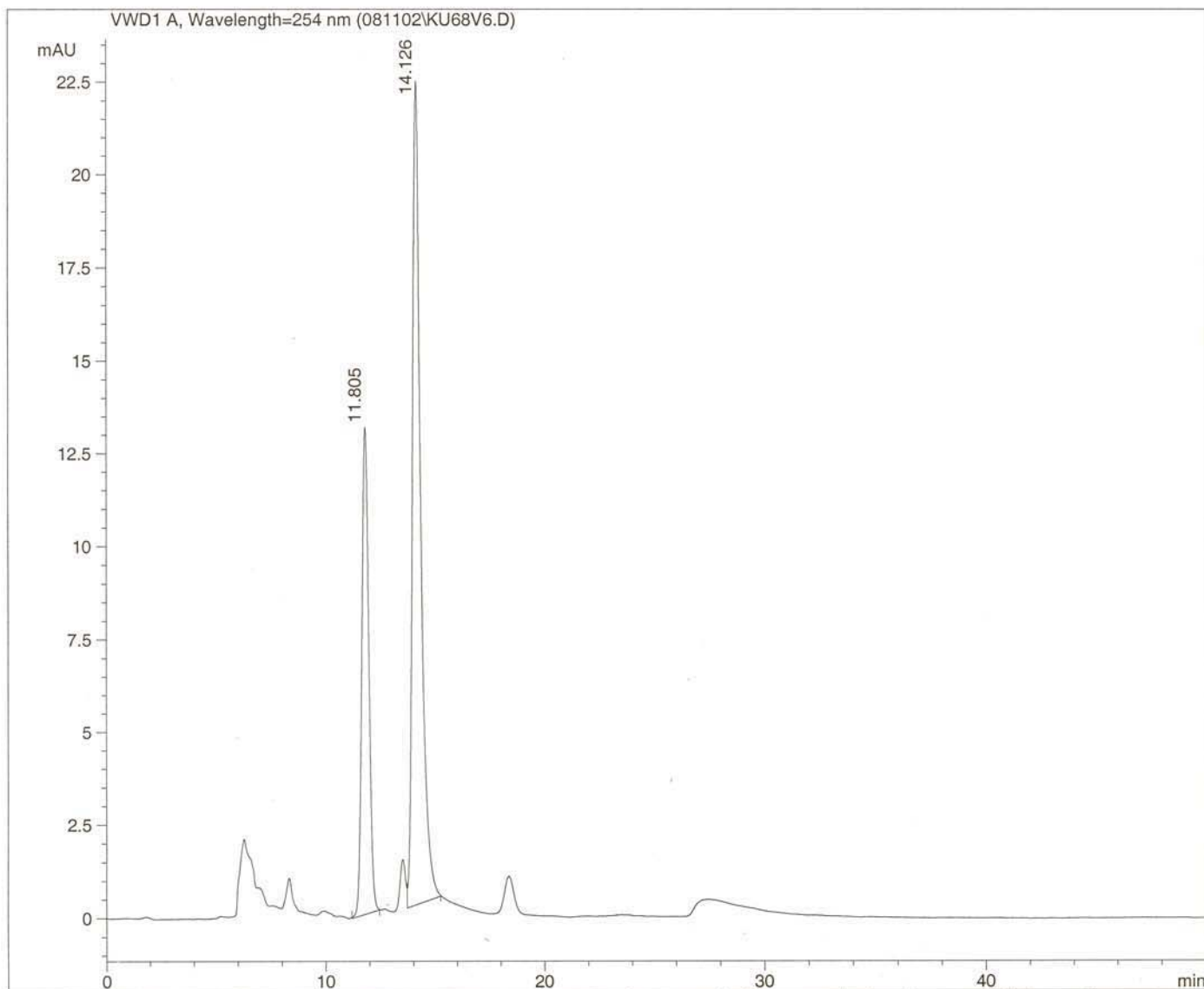
Report date: 11/8/02

2:52:37 PM

data acquired by:

vial: 82

separation of enantiomers of compound 20
with chiral induction of catalyst A7



Signal Desc.	#	Meas. Ret	Area	Area %	Height	Height %	Width	Symmetry
VWD1 A, Wavelen	1	11.80	288.27	33.26	13.11	37.16	0.367	0.907
	2	14.13	578.34	66.74	22.17	62.84	0.435	0.671

chiral hplc → 34% ee

8.069

8.049

7.615

7.594

7.547

7.528

7.418

7.314

7.240

7.164

6.221

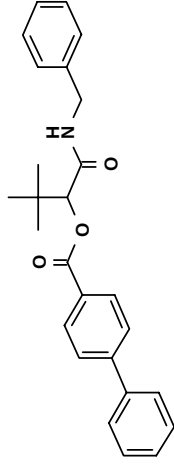
5.058

4.476

4.327

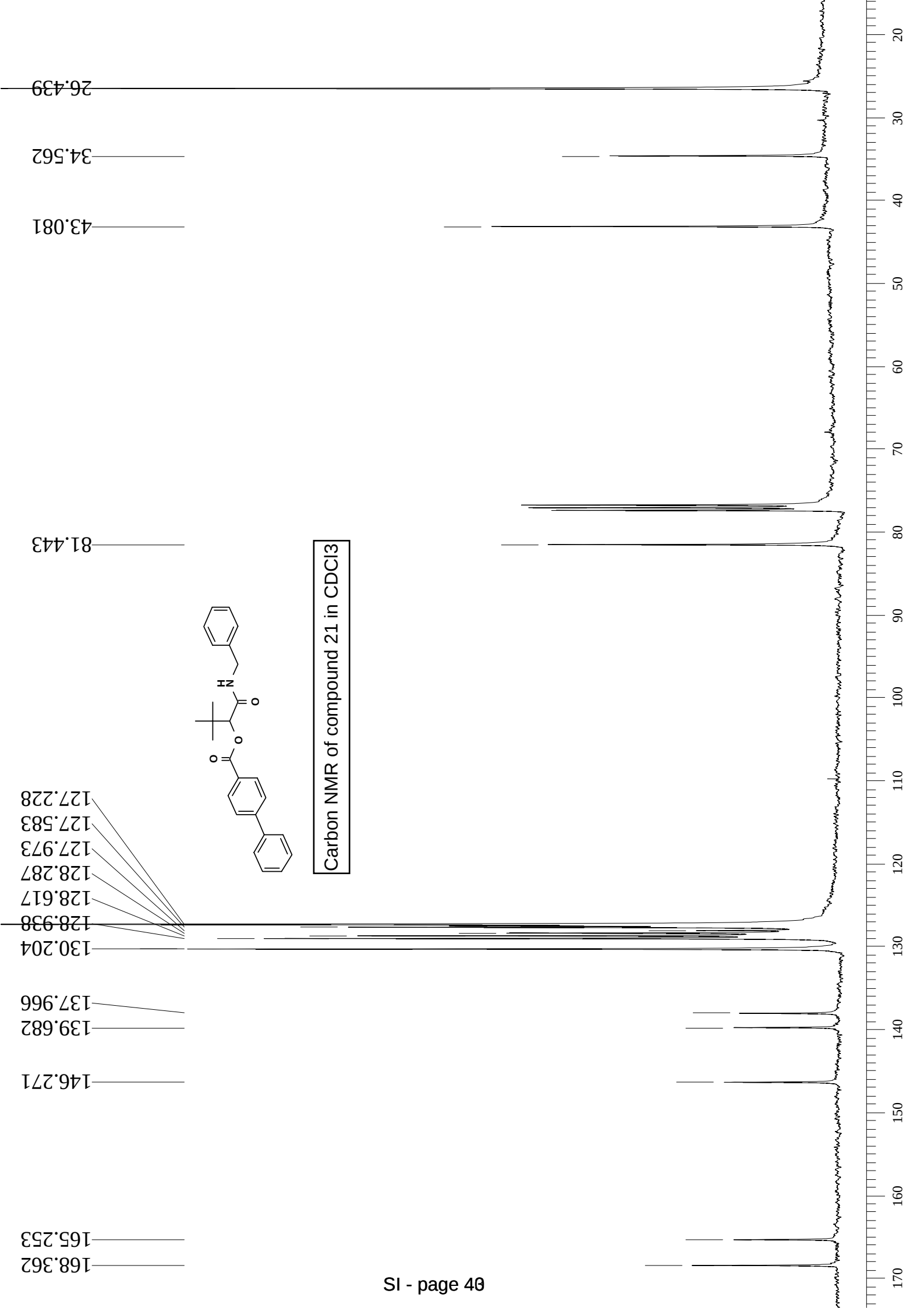
1.085

9.7



Proton NMR of compound 21 in CDCl₃

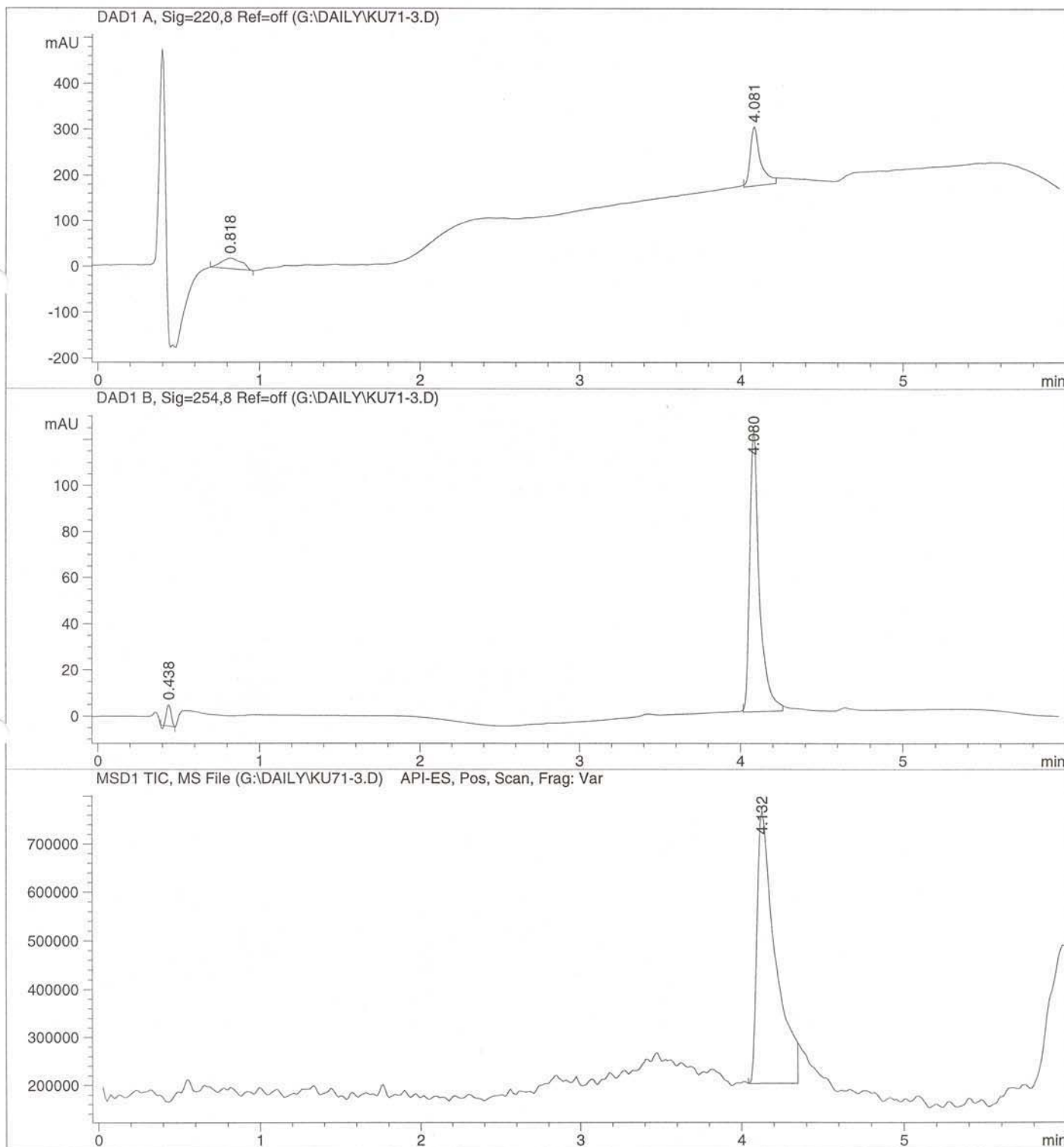
SI - page 42



```
=====
Injection Date   : 11/8/02 2:20:17 PM      Seq. Line :   44
Sample Name     : KU71-3_RK              Location  : Vial 31
Acq. Operator   : user                   Inj       :    1
                                           Inj Volume: 5 µl

Acq. Method     : D:\HPCHEM\1\METHODS\ESI_LC6.M
Last changed    : 11/8/02 2:18:40 PM by user
                  (modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\ESI_LC6.M
Last changed    : 6/18/02 3:31:34 PM
YMC ODS-A 5cm 2µ
ESI positiv
```

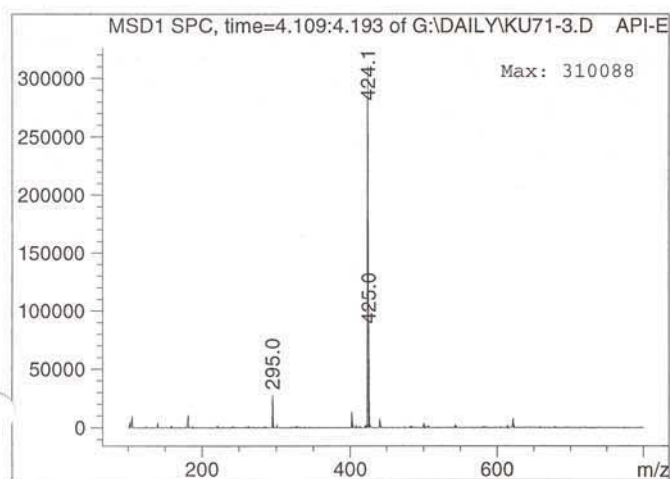
HPLC-MS Spectrum of compound 21



ku71 - after purification with hplc

MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var
Spectra averaged over upper half of peaks.
Noise Cutoff: 1000 counts.
Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
4.132	4424279	425.00 I
		424.10 I



*** End of Report ***

Standard Report

Data File name: D:\HPCHEM\1\DATA\061102\KU65VERD.D
 Sample name: KU65verd
 Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: POS1_3.M

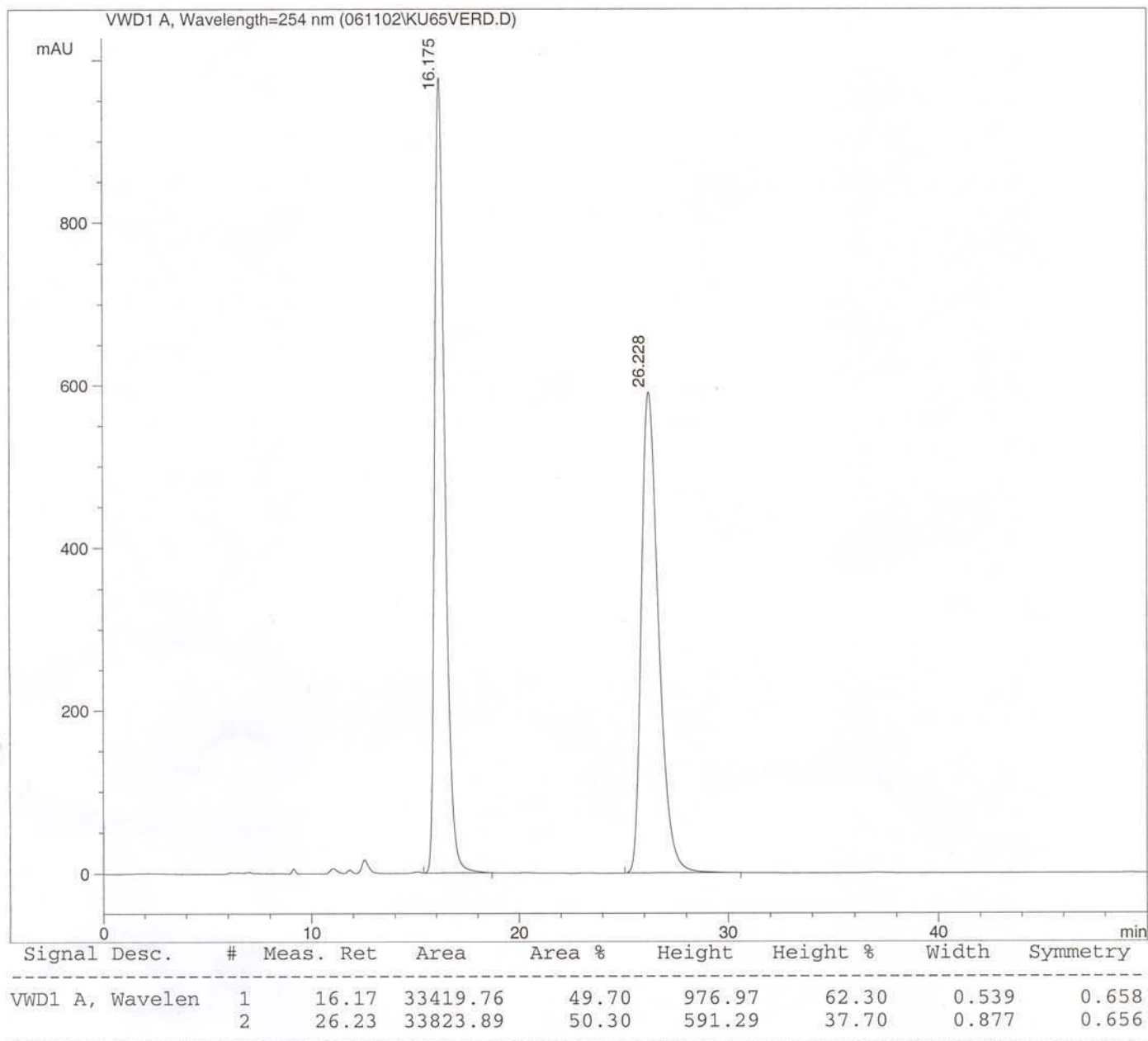
Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:
 Position 1:
 Daicel Chiralcel OD-H
 4.6 x 250

Injection date: 11/6/02 12:40:23 PM
 Report date: 11/6/02 2:14:11 PM

data acquired by:
 vial: 53

separation of enantiomers of compound 21



KU65-separation of enantiomers

Standard Report

Data File name: D:\HPCHEM\1\DATA\081102\KU71-3.D

Sample name: KU71-3

Act. Inj. Vol.: 20.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\POS1_3.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 11/8/02

3:28:56 PM

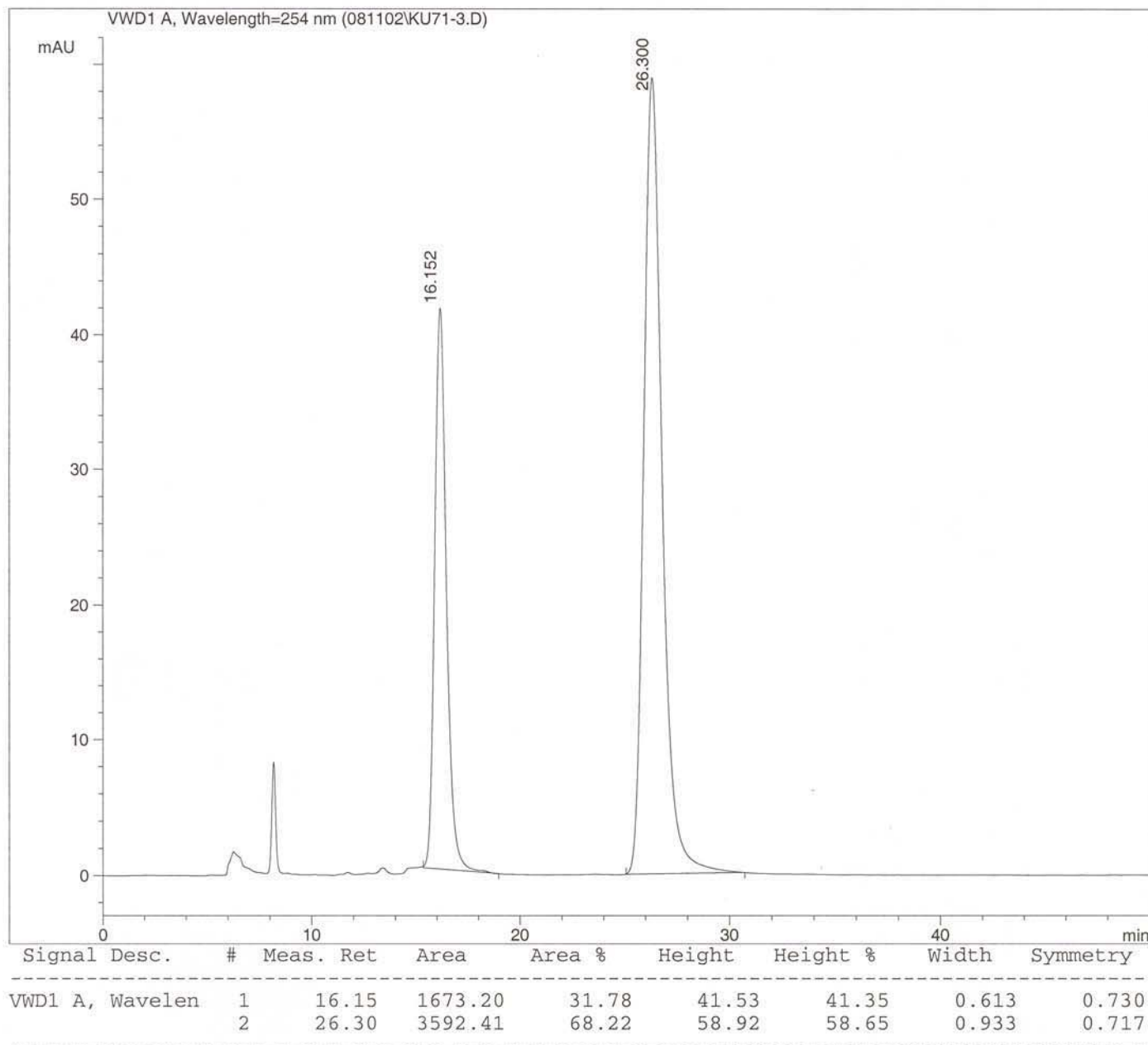
Report date: 11/8/02

4:22:10 PM

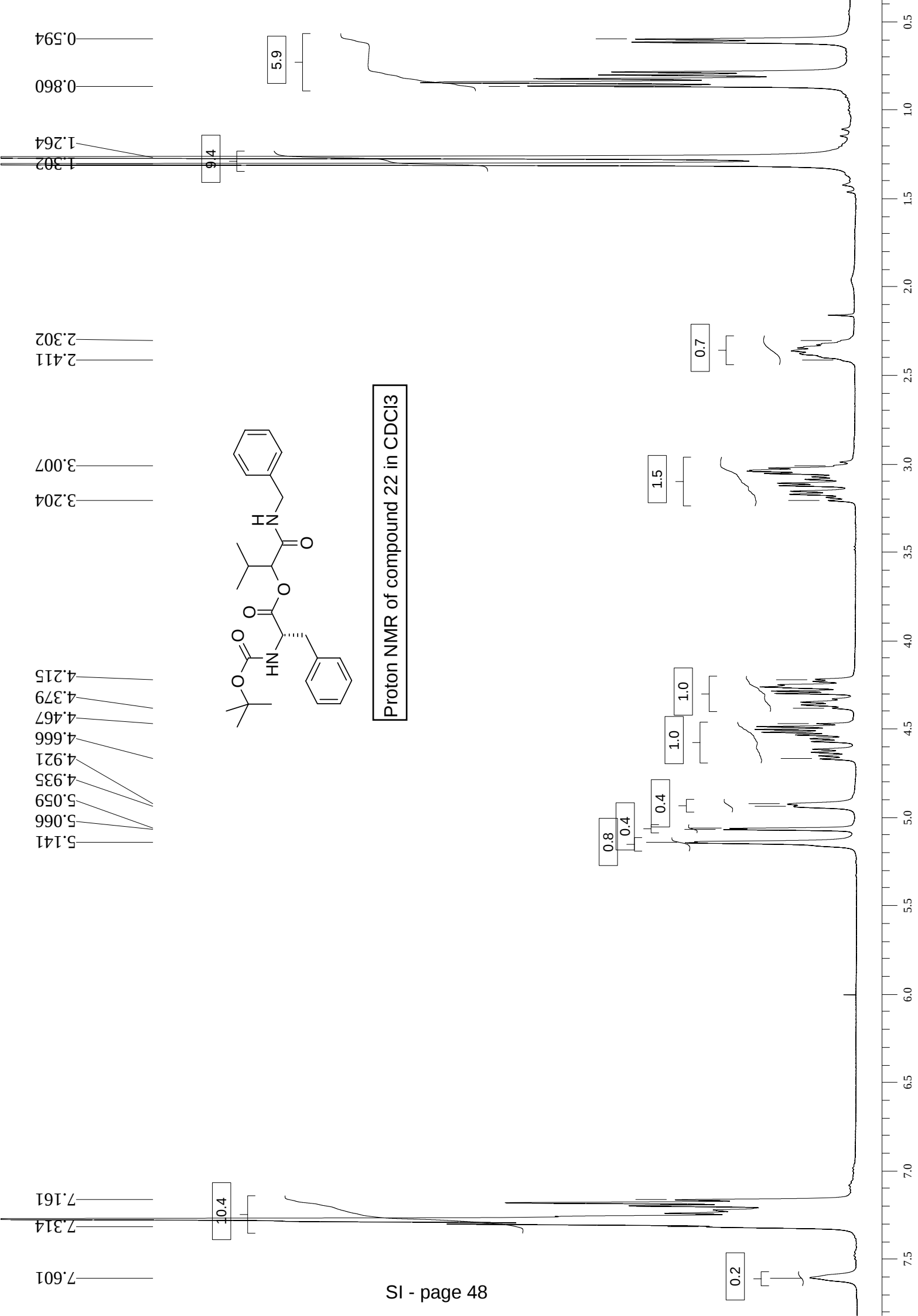
data acquired by:

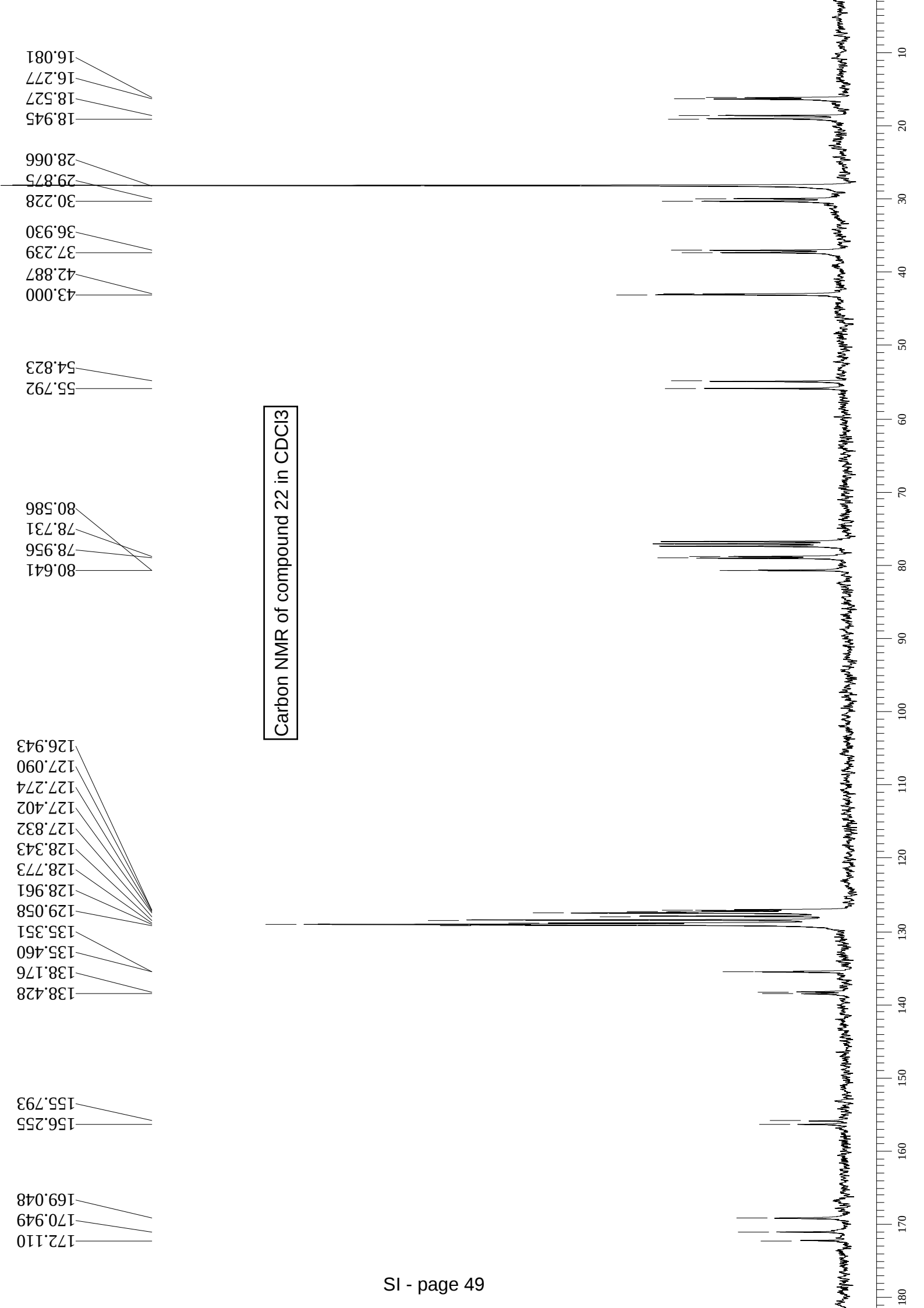
vial: 85

separation of enantiomers of compound 21
with chiral induction of catalyst A7



KU71 - chiral hplc → 36% ee





Standard Report

Data File name: I:\ULRIKE\HPLC4\110602\KU7_10µL.D
 Sample name: KU7
 Act. Inj. Vol.: 10.00

uL

Acquis. Meth.: POS1_3.M

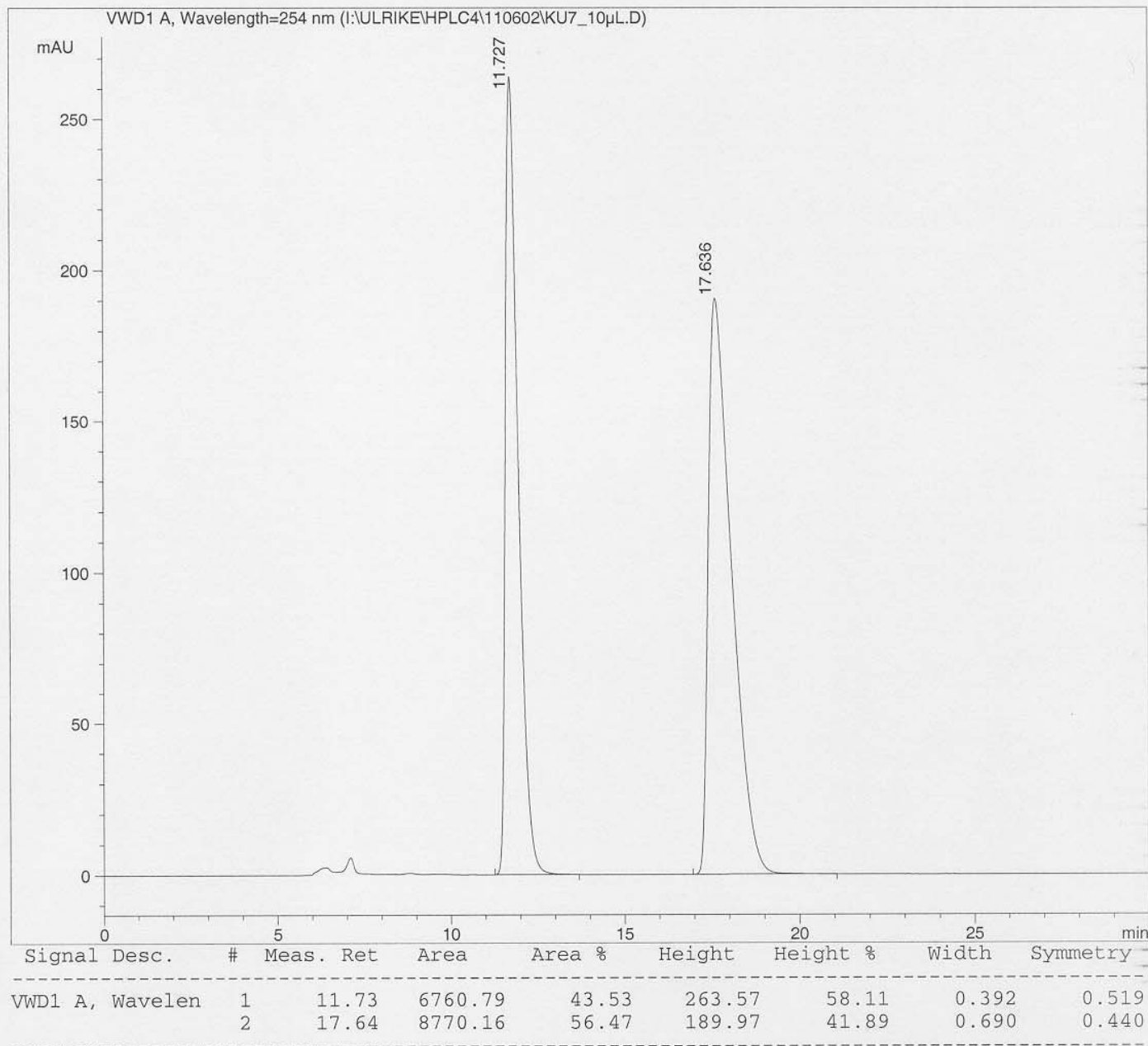
Integr. Method: D:\HPCHEM\1\METHODS\PO51_2.M
 Meth. Inj. Vol.: 10.00 uL

Process Meth. Info:
 Position 5:
 Phenomenex Chirex S-VAL & R-NEA
 4.6 x 50

Injection date: 6/11/02 2:19:20 PM
 Report date: 5/22/03 1:24:15 PM

data acquired by:
 vial: 52

separation of diastereomers of compound 22
 (taken from the supernatant of solution with crystals)



Standard Report

Data File name: I:\ULRIKE\HPLC4\110602\KU7_1482.D
 Sample name: KU7-1482xray
 Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\PO51_2.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

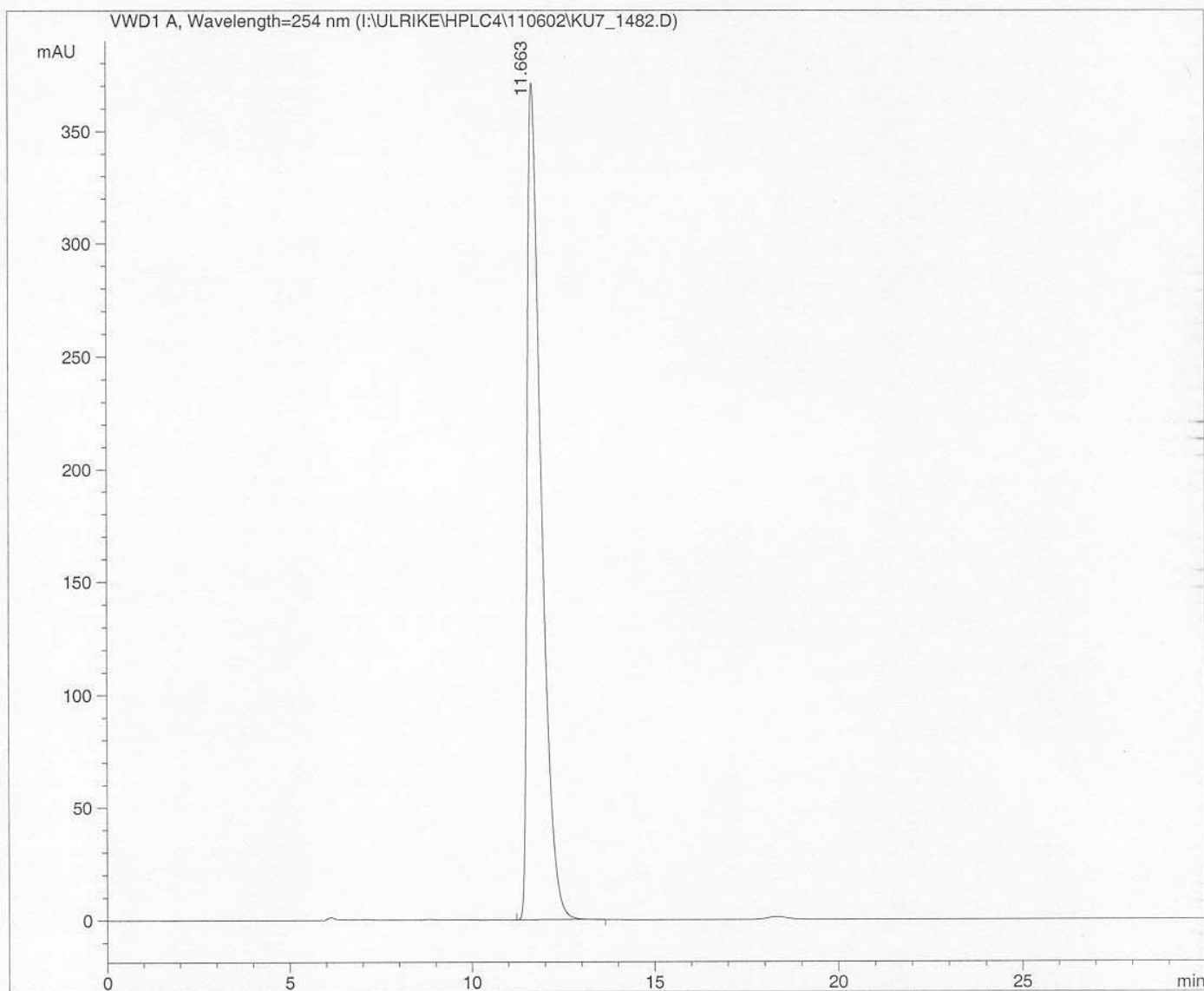
Position 5:

Phenomenex Chirex S-VAL & R-NEA
 4.6 x 50

Injection date: 6/11/02 1:44:57 PM
 Report date: 5/22/03 1:24:47 PM

data acquired by:
 vial: 51

part of the crystal used for single crystal
 X-ray determination of compound 22



Signal Desc.	#	Meas. Ret	Area	Area %	Height	Height %	Width	Symmetry
VWD1 A, Wavelen	1	11.66	9897.53	100.00	371.28	100.00	0.397	0.433

Standard Report

Data File name: I:\ULRIKE\HPLC4\110602\7_4182.D
 Sample name: KU7+KU7-4182
 Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: POS1_3.M

Integr. Method: D:\HPCHEM\1\METHODS\PO51_2.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

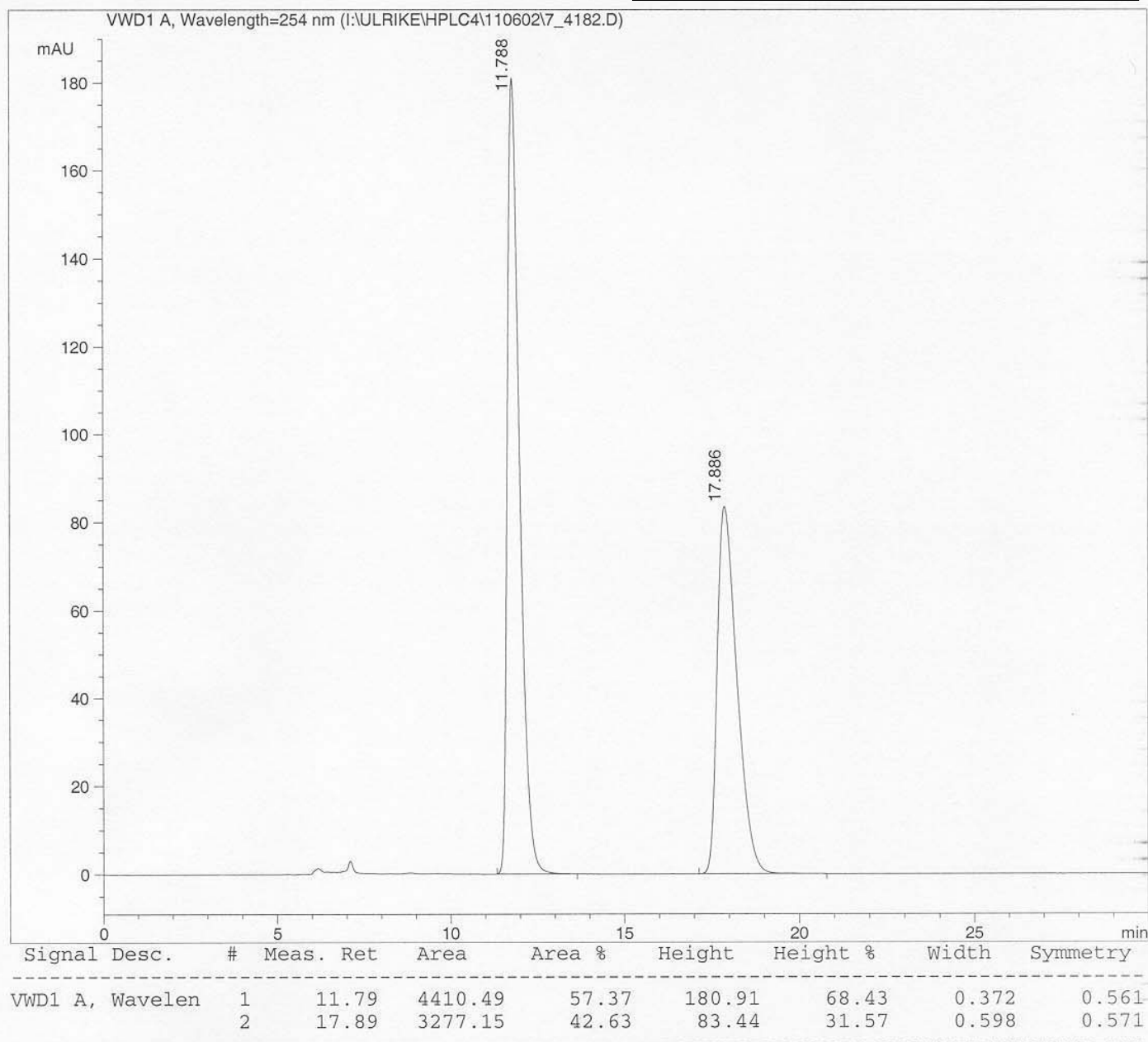
Position 5:

Phenomenex Chirex S-VAL & R-NEA
 4.6 x 50

Injection date: 6/11/02 3:13:44 PM
 Report date: 5/22/03 1:25:40 PM

data acquired by:
 vial: 53

coinjection of single crystal used for X-ray
 determination and solution diastereomeric
 mixture of compound 22



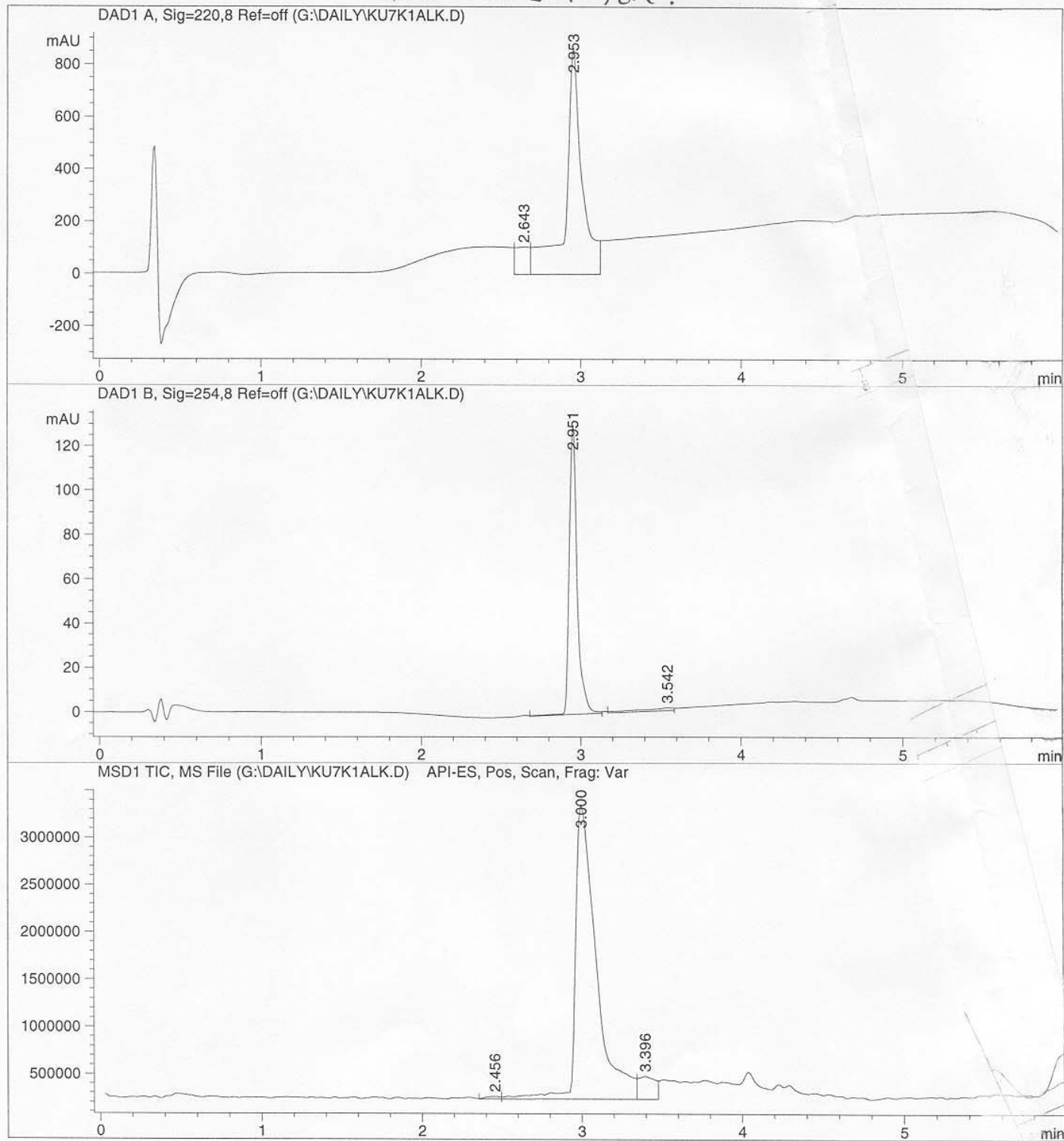
=====

Injection Date	: 3/7/03 7:20:30 PM	Seq. Line	: 79
Sample Name	: KU7K1alk	Location	: Vial 1
Acq. Operator	: Brixner	Inj	: 1
		Inj Volume	: 5 µl
Acq. Method	: D:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 3/7/03 7:18:59 PM by Brixner (modified after loading)		
Analysis Method	: C:\HPCHEM\1\METHODS\ESI_LC6.M		
Last changed	: 3/7/03 10:48:48 AM (modified after loading)		

YMC ODS-A 5cm 2µ
ESI positiv

HPLC-MS spectrum of compound 23

*Kristall - versetzt - abgeheutet
Alkohol auf LC1 → ok!*



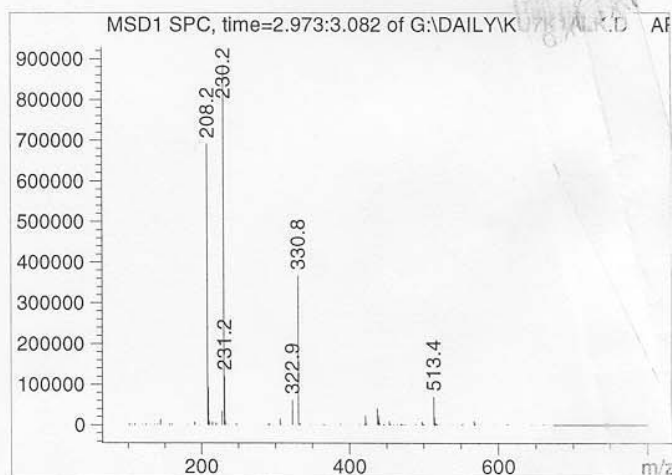
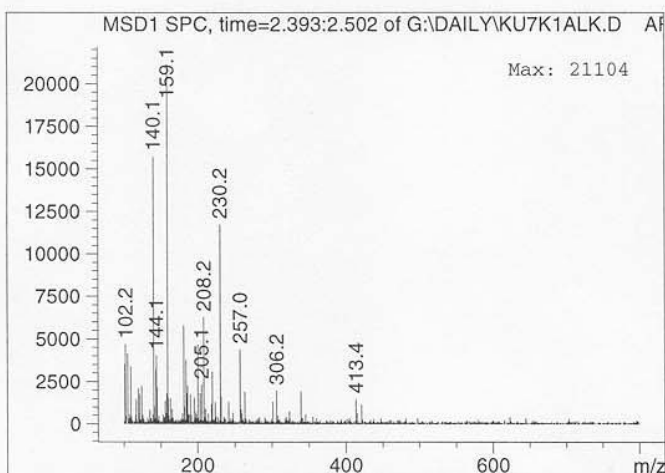
MS Signal: MSD1 TIC, MS File, API-ES, Pos, Scan, Frag: Var

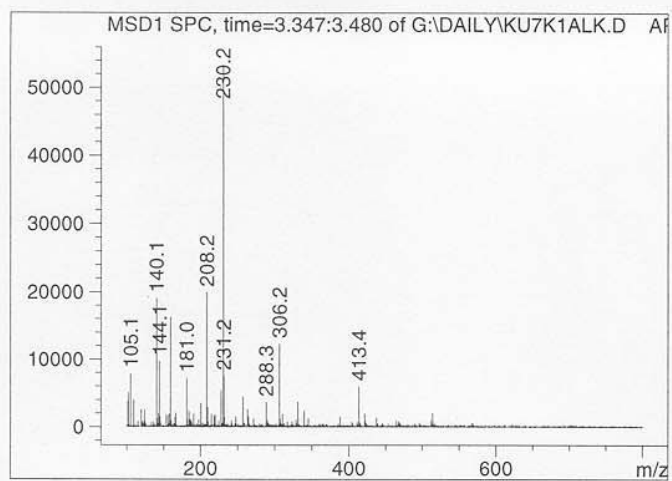
Spectra averaged over upper half of peaks.

Noise Cutoff: 1000 counts.

Reportable Ion Abundance: > 10%.

Retention Time (MS)	MS Area	Mol. Weight or Ion
2.456	184763	256.95 I
		230.20 I
		219.20 I
		208.20 I
		205.10 I
		200.10 I
		186.05 I
		184.10 I
		181.00 I
		159.10 I
		145.05 I
		144.10 I
		143.05 I
		140.10 I
		124.10 I
		109.15 I
		105.10 I
		102.20 I
		101.20 I
3.000	28285604	330.80 I
		231.20 I
		230.20 I
		209.20 I
		208.20 I
3.396	1812192	413.40 I
		306.20 I
		231.20 I
		230.20 I
		227.30 I
		208.20 I
		181.00 I
		159.05 I
		144.15 I
		140.10 I
		105.10 I





*** End of Report ***

Standard Report

Data File name: D:\HPCHEM\1\DATA\DAILY\BBKRI.D

Sample name: bbkri

Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: PO11_5_D.M

Integr. Method: D:\HPCHEM\1\METHODS\PO11_10.M

Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:

Position 1:

Daicel Chiralcel OD-H

4.6 x 250

Injection date: 3/26/03

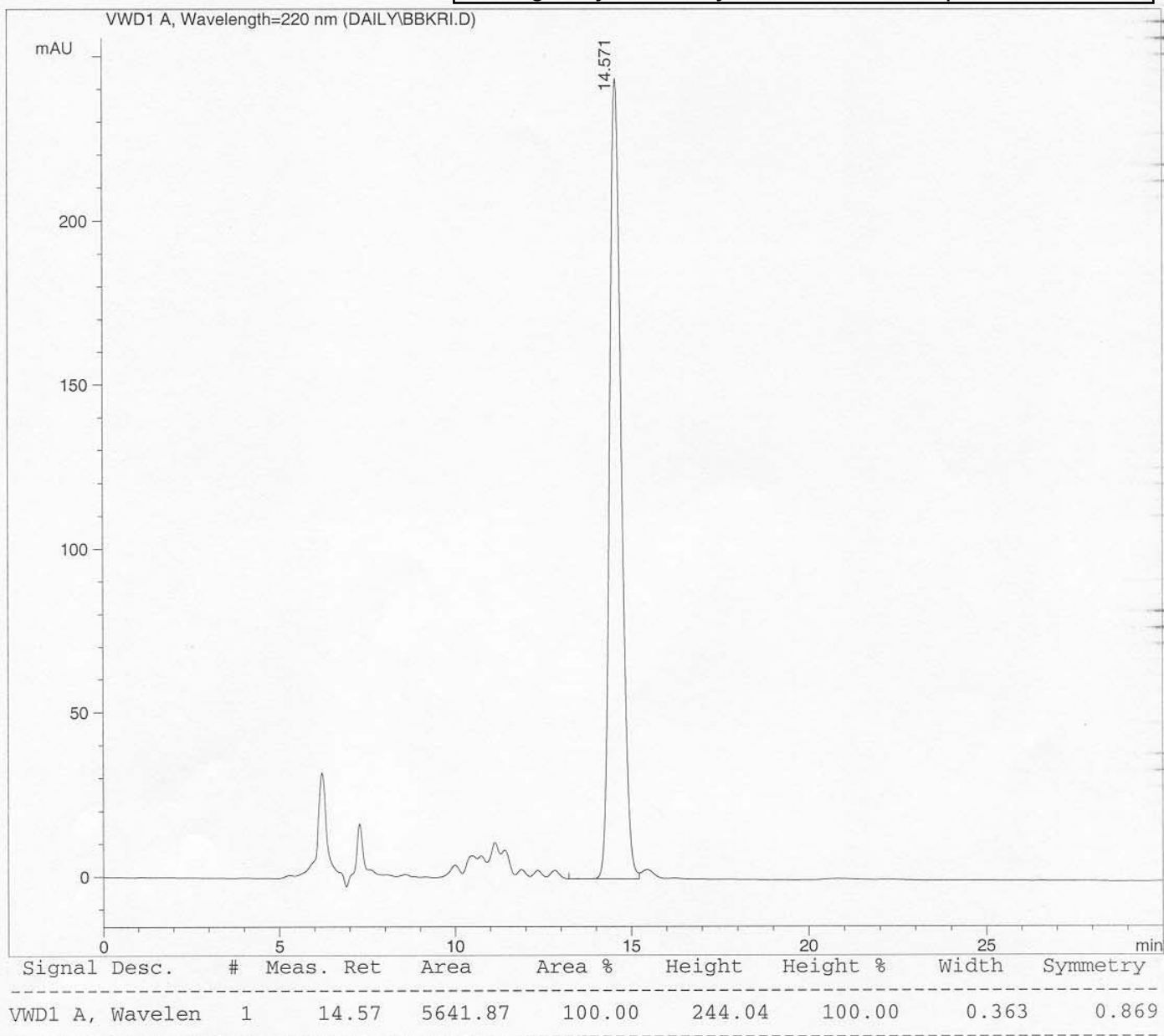
1:58:36 PM

Report date: 5/23/03

7:22:32 PM

data acquired by:
vial: 2

alcohol 23 resulting from saponification of crystal 22 used
for single crystal X-ray determination; Experiment A



Standard Report

Data File name: D:\HPCHEM\1\DATA\BH\KU6IN.D
 Sample name: ku6in
 Act. Inj. Vol.: 10.00

uL

Acquis. Meth.: P011_5_D.M

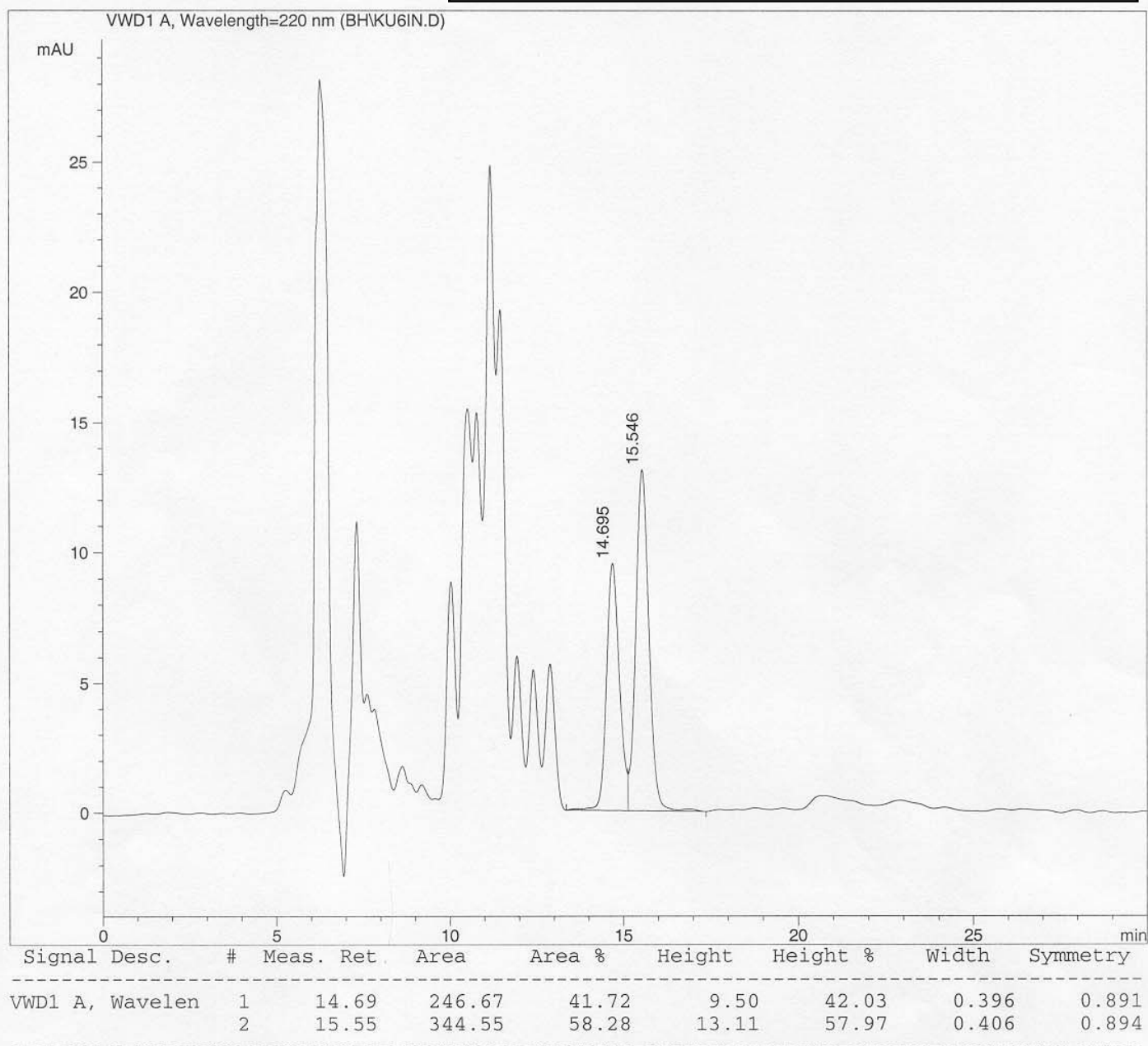
Integr. Method: D:\HPCHEM\1\METHODS\P011_7.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:
 Position 1:
 Daicel Chiralcel OD-H
 4.6 x 250

Injection date: 3/25/03 1:33:38 PM
 Report date: 3/25/03 3:32:30 PM

data acquired by:
 vial: 12

alcohol 23 resulting from saponification of ester 16 with
 chiral induction of catalyst A7; Experiment B



Standard Report

Data File name: D:\HPCHEM\1\DATA\DAILY\BBCOIN.D
 Sample name: bbcoin
 Act. Inj. Vol.: 5.00

uL

Acquis. Meth.: PO11_5_D.M

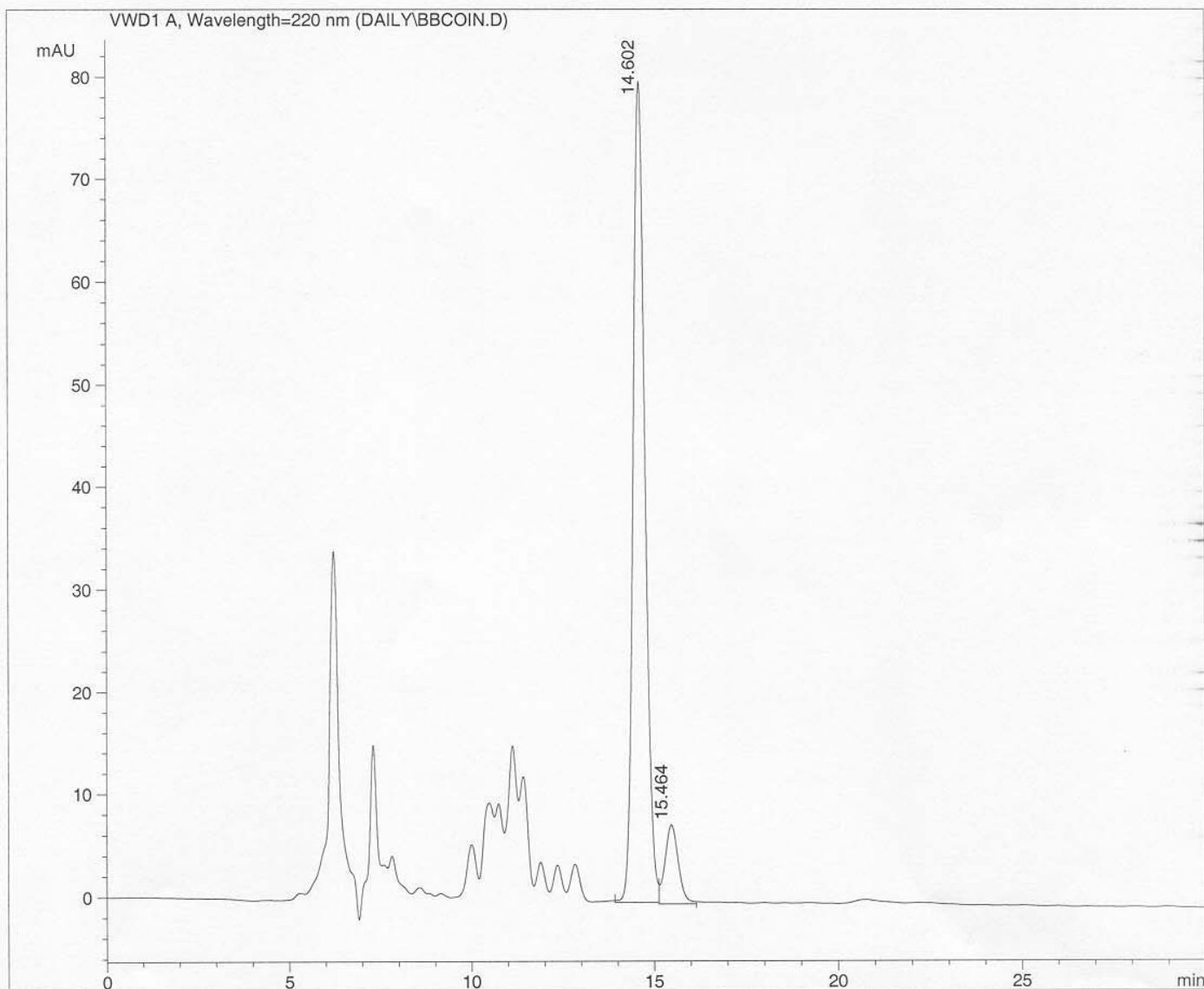
Integr. Method: D:\HPCHEM\1\METHODS\PO11_10.M
 Meth. Inj. Vol.: 5.00 uL

Process Meth. Info:
 Position 1:
 Daicel Chiralcel OD-H
 4.6 x 250

Injection date: 3/26/03 2:29:44 PM
 Report date: 5/23/03 7:21:56 PM

data acquired by:
 vial: 3

alcohol 23 coinjection of hplc solutions
 from experiment A and B



Signal Desc.	#	Meas. Ret	Area	Area %	Height	Height %	Width	Symmetry
VWD1 A, Wavelen	1	14.60	1806.25	90.25	80.12	91.24	0.376	0.906
	2	15.46	195.20	9.75	7.69	8.76	0.423	0.899

=====

SUPPORTING INFORMATION

=====

DOEA 4182-173

Compound 22 (C₂₆H₃₄N₂O₅)

BELONGING TO THE PAPER

Massive Parallel Catalyst Screening: Towards Asymmetric MCRs

by

Ulrike Kusebauch, Barbara Beck, Kim Messer, Eberhardt Herdtweck, Alexander Dömling

Contents

=====

Table S1 - Crystal Data and Details of the Structure Determination
for: DOEA 4182-173

Table S2 - Final Coordinates and Equivalent Isotropic Thermal
Parameters of the non-Hydrogen atoms
for: DOEA 4182-173

Table S3 - Hydrogen Atom Positions and Isotropic Thermal
Parameters
for: DOEA 4182-173

Table S4 - (An)isotropic Thermal Parameters
for: DOEA 4182-173

Table S5 - Bond Distances (Å)
for: DOEA 4182-173

Table S6 - Bond Angles (Degrees)
for: DOEA 4182-173

Table S1 - Crystal Data and Details of the Structure Determination for DOE A 4182-173

Operator:	*** Herdtweck ***
Molecular Formula:	C ₂₆ H ₃₄ N ₂ O ₅
Crystal Colour / Shape	Colorless fragment. Approximate size of crystal used for data collection:
Crystal Size	0.76 × 0.64 × 0.38 mm
Molecular Weight:	454.55 a.m.u.
F ₀₀₀ :	976
Systematic Absences:	h00: h≠2n; 0k0: k≠2n; 00l: l≠2n
Space Group:	Orthorhombic P 2 ₁ 2 ₁ 2 ₁ (I.T.-No.: 19)
Cell Constants:	Least-squares refinement of 2669 reflections with the programs "DENZO / HKL" [1, 2]; Theta range 1.98° < Θ < 25.33°; Mo(K α); λ = 0.71073 Å a = 11.2376(1) Å b = 12.3949(1) Å c = 18.3839(1) Å V = 2560.67(3) Å ³ ; Z = 4; D _{calc} = 1.179 g cm ⁻³ ; Mos. = 0.406(1)
Diffractometer:	Kappa CCD (Area Diffraction System; NONIUS); rotating anode; graphite monochromator; 50 kV; 60 mA; λ = 0.71073 Å; Mo(K α)
Temperature:	(-100±1) °C; (173±1) K
Measurement Range:	1.98°<Θ<25.33°; h: -13/13, k: -14/14, l: -22/22
Measurement Time:	2 × 2.5 s per film
Measurement Mode:	9 sets; 1034 films φ- and Ω-movement; Increment: Δφ/ΔΩ = 1.0°; dx = 40.0 mm
LP - Correction:	Yes [2]
Intensity Correction	No
Absorption Correction:	No; μ = 0.082 mm ⁻¹
Reflection Data:	97013 reflections were used 57025 reflections were integrated 57025 reflections to be merged 4666 independent reflections

	0.037	R_{int} : (basis F_o^2)	
	4666	independent reflections (all) were used in refinements	
	4314	independent reflections with $I_o > 2\sigma(I_o)$	
	100.0%	completeness of the data set	
	435	Parameter full-matrix refinement	
	10.7	reflections per parameter	
Solution:	Direct Methods [3]; Difference Fourier syntheses		
Refinement Parameters:	In the asymmetric unit: 33 Non-hydrogen atoms with anisotropic displacement parameters 34 Hydrogen atoms with isotropic displacement parameters		
Hydrogen Atoms:	All hydrogen atom positions were found in the difference map calculated from the model containing all non-hydrogen atoms. The hydrogen positions were refined with individual isotropic displacement parameters.		
Atomic Form Factors:	For neutral atoms and anomalous dispersion [4]		
Extinction Correction:	$F_c(\text{kor}) = kF_c[1 + 0.001 \cdot \epsilon \cdot F_c^2 \cdot \lambda^3 / \sin(2\Theta)]^{-1/4}$ SHELXL-97 [5] ϵ refined: $\epsilon = 0.0121(15)$		
Weighting Scheme:	$w^{-1} = \sigma^2(F_o^2) + (a \cdot P)^2 + b \cdot P$. with a: 0.0379; b: 0.2620; P: $[\text{Maximum}(0 \text{ or } F_o^2) + 2 \cdot F_c^2] / 3$		
Shift/Err :	Less than 0.001 in the last cycle of refinement:		
Resid. Electron Dens.:	$+0.11 \text{ e}_0; ^{-} / \text{\AA}^3$; $-0.11 \text{ e}_0; ^{-} / \text{\AA}^3$		
R1 :	$\sum(F_o - F_c) / \sum F_o $		
[$F_o > 4\sigma(F_o)$;	N=4314]:		= 0.0277
[all reflctns;	N=4666 :		= 0.0319
wR2 :	$[\sum w (F_o^2 - F_c^2)^2 / \sum w (F_o^2)^2]^{1/2}$		
[$F_o > 4\sigma(F_o)$;	N=4314]:		= 0.0661
[all reflctns;	N=4666]:		= 0.0680
Goodness of fit :	$[\sum w (F_o^2 - F_c^2)^2 / (\text{NO} - \text{NV})]^{1/2}$		
			= 1.045
Flack's Parameter :	$x = 0.1(6)$		
Remarks:	Refinement expression $\sum w(F_o^2 - F_c^2)^2$ The correct enantiomere could not be proved by Flack's Parameter.		

Hydrogen bond exists between:

D - H A	D A	D-H	H · A	D-H A	Σ
N1-H1 ... O5	287.7(1)	84(2)	204(2)	171(1)°	
N2-H2 ... O1	264.8(1)	85(2)	227(2)	107(1)°	
N2-H2 ... O3	281.3(1)	85(2)	201(2)	156(1)°	357(2)°

All tables are calculated with the program PLATON [6] --->
DOEA_FINE.CIF (options round 2 ; table IC)

----->

No match in a CSD-Data-Research [8]; 224400 entries; version 2/00

References:

Measurements and Data Reduction

[1] Data Collection Software for Nonius KappaCCD, Delft, The Netherlands, (1997).

[2] Otwinowski, Z.; Minor, W., *Methods in Enzymology* **1997**, 276, 307.

Solution

[3] Altomare, A.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli M. "SIR92", *J. Appl. Cryst.* **1994**, 27, 435-436.

Refinement

[4] International Tables for Crystallography, Vol. C, Tables 6.1.1.4 (pp. 500-502), 4.2.6.8 (pp. 219-222), and 4.2.4.2 (pp. 193-199), Wilson, A. J. C., Ed., Kluwer Academic Publishers, Dordrecht, The Netherlands, 1992.

[5] Sheldrick, G. M. "SHELXL-97", University of Göttingen, Göttingen, Germany, (1998).

Graphics

[6] Spek, A. L. "PLATON", A Multipurpose Crystallographic Tool, Utrecht University, Utrecht, The Netherlands, (2001).

Miscellaneous

[7] Artus, G.; Scherer, W.; Priermeier, T.; Herdtweck, E. "S T R U X - V", A Program System to Handle X-Ray Data, TU München, Germany, (1997).

[8] 3d Search and research Using the Cambridge Structural Database; Allen, F. H.; Kennard, O. *Chemical Design Automation News* **1993**, 8, 1 & 31-37.

Table S2 - Final Coordinates and Equivalent Isotropic Thermal Parameters of the non-Hydrogen atoms for DOE A 4182-173

Atom	x	y	z	U(eq) [Å ²]
----	---	---	---	-----
O1	0.22428(7)	0.60882(7)	0.18632(4)	0.0248(2)
O2	0.08237(10)	0.49383(9)	0.14754(5)	0.0480(3)
O3	0.36705(8)	0.41904(7)	0.11375(5)	0.0346(3)
O4	0.34362(9)	0.31908(7)	0.01111(5)	0.0356(3)
O5	0.29290(9)	0.53289(8)	0.36883(5)	0.0376(3)
N1	0.24360(10)	0.47190(8)	0.02371(6)	0.0289(3)
N2	0.36813(11)	0.48083(10)	0.26099(6)	0.0351(3)
C1	0.16498(11)	0.55179(10)	0.13533(7)	0.0272(4)
C2	0.21729(11)	0.57365(10)	0.05981(6)	0.0254(4)
C3	0.12780(14)	0.63681(12)	0.01377(7)	0.0343(4)
C4	0.09721(13)	0.74673(11)	0.04402(7)	0.0345(4)
C5	0.18254(18)	0.82578(13)	0.05122(10)	0.0508(6)
C6	0.1520(2)	0.92793(15)	0.07620(12)	0.0739(8)
C7	0.0357(2)	0.95148(16)	0.09396(11)	0.0734(8)
C8	-0.0493(2)	0.87392(17)	0.08787(9)	0.0619(7)
C9	-0.01882(15)	0.77170(14)	0.06342(8)	0.0464(5)
C10	0.32251(12)	0.40409(10)	0.05393(6)	0.0269(3)
C11	0.43015(14)	0.23506(11)	0.03365(8)	0.0404(5)
C12	0.4156(3)	0.15131(17)	-0.02630(11)	0.0693(8)
C13	0.3946(2)	0.18574(14)	0.10561(10)	0.0564(7)
C14	0.55318(17)	0.28322(19)	0.03602(14)	0.0646(7)
C15	0.19182(11)	0.59469(10)	0.26231(6)	0.0266(3)
C16	0.16467(13)	0.70453(11)	0.29602(7)	0.0332(4)
C17	0.26557(18)	0.78366(13)	0.28292(9)	0.0487(6)
C18	0.04610(17)	0.74767(16)	0.26948(10)	0.0528(6)
C19	0.29055(11)	0.53420(10)	0.30167(6)	0.0279(4)
C20	0.45540(13)	0.40772(13)	0.29220(8)	0.0398(4)
C21	0.40711(13)	0.29781(12)	0.31238(7)	0.0357(4)
C22	0.47938(15)	0.22805(13)	0.35195(9)	0.0462(5)
C23	0.43993(17)	0.12742(14)	0.37213(10)	0.0536(6)
C24	0.32814(18)	0.09354(14)	0.35373(10)	0.0569(6)
C25	0.2550(2)	0.16154(15)	0.31446(12)	0.0656(7)
C26	0.29460(16)	0.26320(14)	0.29373(10)	0.0532(6)

=====

U(eq) = 1/3 of the trace of the orthogonalized U

Table S3 - Hydrogen Atom Positions and Isotropic Thermal Parameters for DOEA
4182-173

Atom	x	y	z	U(iso) [Å ²]
----	---	---	---	-----
H1	0.2264(13)	0.4664(12)	-0.0208(9)	0.036(4)
H2	0.3625(14)	0.4816(13)	0.2147(9)	0.041(4)
H21	0.2918(12)	0.6152(10)	0.0675(7)	0.023(3)
H31	0.1625(14)	0.6449(12)	-0.0324(9)	0.041(4)
H32	0.0562(14)	0.5905(13)	0.0096(8)	0.038(4)
H51	0.2634(18)	0.8172(15)	0.0404(10)	0.060(5)
H61	0.212(2)	0.980(2)	0.0812(12)	0.084(7)
H71	0.013(2)	1.0246(19)	0.1120(12)	0.089(7)
H81	-0.138(2)	0.8907(18)	0.1042(11)	0.086(6)
H91	-0.0818(18)	0.7134(17)	0.0602(11)	0.073(6)
H121	0.328(3)	0.130(2)	-0.0313(14)	0.102(9)
H122	0.4680(19)	0.0926(18)	-0.0175(11)	0.073(6)
H123	0.4466(18)	0.1877(18)	-0.0715(12)	0.080(7)
H131	0.308(2)	0.1656(17)	0.1053(11)	0.070(6)
H132	0.4102(17)	0.2350(16)	0.1485(10)	0.066(6)
H133	0.435(2)	0.1189(19)	0.1097(11)	0.081(6)
H141	0.570(2)	0.310(2)	-0.0163(16)	0.116(9)
H142	0.6085(18)	0.2259(16)	0.0482(10)	0.070(6)
H143	0.5602(19)	0.3369(19)	0.0753(13)	0.083(7)
H151	0.1202(13)	0.5494(11)	0.2622(7)	0.030(3)
H161	0.1595(12)	0.6896(12)	0.3495(8)	0.035(4)
H171	0.2470(18)	0.8492(18)	0.3127(11)	0.074(6)
H172	0.2710(14)	0.8002(14)	0.2309(10)	0.049(4)
H173	0.3424(19)	0.7550(16)	0.2998(11)	0.071(6)
H181	0.0256(18)	0.8219(19)	0.2913(11)	0.076(6)
H182	-0.0241(19)	0.6986(18)	0.2856(10)	0.072(6)
H183	0.0476(16)	0.7584(15)	0.2164(11)	0.062(5)
H201	0.4869(14)	0.4406(13)	0.3374(9)	0.044(4)
H202	0.5206(14)	0.3994(13)	0.2565(9)	0.045(4)
H221	0.5568(18)	0.2527(15)	0.3647(10)	0.060(5)
H231	0.4909(16)	0.0811(15)	0.3996(10)	0.060(5)
H241	0.2981(16)	0.0246(17)	0.3667(10)	0.064(5)
H251	0.1775(19)	0.1398(16)	0.3015(11)	0.069(6)
H261	0.2433(15)	0.3084(15)	0.2671(10)	0.054(5)

=====

The Temperature Factor has the Form of Exp(-T) Where
 $T = 8*(\pi^2)*U*(\sin(\theta)/\lambda)^2$ for Isotropic Atoms

Table S4 - (An)isotropic Thermal Parameters for DOEA 4182-173

Atom	U(1,1)	U(2,2)	U(3,3)	U(2,3)	U(1,3)	U(1,2)
O1	0.0308(4)	0.0268(4)	0.0168(4)	-0.0021(3)	-0.0004(3)	-0.0016(4)
O2	0.0535(6)	0.0579(7)	0.0325(5)	-0.0095(5)	0.0042(5)	-0.0287(5)
O3	0.0461(5)	0.0316(5)	0.0262(5)	-0.0056(4)	-0.0108(4)	0.0086(4)
O4	0.0479(6)	0.0310(5)	0.0279(5)	-0.0082(4)	-0.0082(4)	0.0150(4)
O5	0.0511(6)	0.0429(5)	0.0187(4)	0.0028(4)	0.0017(4)	0.0048(4)
N1	0.0412(6)	0.0259(5)	0.0196(5)	-0.0046(4)	-0.0051(5)	0.0080(5)
N2	0.0434(6)	0.0411(6)	0.0209(5)	0.0020(5)	-0.0019(5)	0.0114(5)
C1	0.0329(7)	0.0241(6)	0.0245(6)	-0.0033(5)	-0.0038(5)	0.0003(5)
C2	0.0323(7)	0.0231(6)	0.0208(6)	-0.0027(5)	-0.0018(5)	0.0029(5)
C3	0.0436(8)	0.0343(7)	0.0249(7)	-0.0026(6)	-0.0059(6)	0.0102(6)
C4	0.0475(8)	0.0311(7)	0.0249(6)	0.0042(5)	-0.0005(6)	0.0117(6)
C5	0.0615(11)	0.0345(8)	0.0563(10)	0.0003(7)	0.0174(8)	0.0021(8)
C6	0.1137(19)	0.0313(9)	0.0768(13)	-0.0005(9)	0.0252(13)	-0.0059(11)
C7	0.127(2)	0.0378(10)	0.0554(11)	0.0044(8)	0.0236(12)	0.0379(12)
C8	0.0780(13)	0.0668(13)	0.0409(9)	0.0044(8)	0.0063(9)	0.0449(11)
C9	0.0460(9)	0.0576(10)	0.0355(8)	0.0007(7)	-0.0058(7)	0.0211(8)
C10	0.0330(6)	0.0250(6)	0.0227(6)	-0.0027(5)	0.0001(5)	0.0018(5)
C11	0.0543(9)	0.0344(8)	0.0326(7)	-0.0023(6)	-0.0062(7)	0.0206(7)
C12	0.1084(19)	0.0515(11)	0.0479(11)	-0.0166(9)	-0.0163(11)	0.0474(12)
C13	0.0873(15)	0.0323(8)	0.0495(10)	0.0039(8)	-0.0011(9)	0.0095(9)
C14	0.0489(11)	0.0681(13)	0.0767(14)	0.0080(11)	0.0020(10)	0.0220(10)
C15	0.0314(6)	0.0306(6)	0.0177(5)	-0.0009(5)	0.0028(5)	-0.0034(6)
C16	0.0433(8)	0.0340(7)	0.0224(6)	-0.0049(5)	-0.0005(6)	0.0064(6)
C17	0.0711(12)	0.0338(8)	0.0412(9)	-0.0106(7)	-0.0011(8)	-0.0066(8)
C18	0.0590(10)	0.0612(11)	0.0383(9)	-0.0092(8)	-0.0078(8)	0.0260(10)
C19	0.0364(7)	0.0257(6)	0.0217(6)	0.0004(5)	0.0010(5)	-0.0025(5)
C20	0.0398(8)	0.0460(8)	0.0335(7)	0.0007(7)	-0.0049(6)	0.0123(7)
C21	0.0404(8)	0.0412(8)	0.0254(6)	-0.0057(6)	-0.0010(6)	0.0133(6)
C22	0.0444(9)	0.0498(9)	0.0443(8)	0.0049(7)	-0.0006(7)	0.0151(7)
C23	0.0589(11)	0.0465(10)	0.0554(10)	0.0066(8)	-0.0020(8)	0.0196(8)
C24	0.0734(12)	0.0362(8)	0.0611(11)	-0.0039(8)	-0.0027(9)	0.0058(9)
C25	0.0644(12)	0.0502(10)	0.0821(13)	-0.0049(9)	-0.0259(11)	-0.0022(9)
C26	0.0556(10)	0.0454(9)	0.0585(10)	0.0004(8)	-0.0200(8)	0.0086(8)

=====

The Temperature Factor has the Form of $\exp(-T)$ Where
 $T = 8(\pi^2)U(\sin(\theta)/\lambda)^2$ for Isotropic Atoms
 $T = 2(\pi^2)\sum_i \sum_j h(i)h(j)U(i,j)A^*(i)A^*(j)$, for
Anisotropic Atoms. $A^*(i)$ are Reciprocal Axial Lengths and
 $h(i)$ are the Reflection Indices.

Table S5 - Bond Distances (Å) for **DOEA 4182-173**

O1 -C1	1.3500(15)	N1 -H1	0.844(16)
O1 -C15	1.4544(13)	N2 -H2	0.853(17)
O2 -C1	1.1951(17)	C2 -H21	0.993(13)
O3 -C10	1.2224(15)	C3 -H31	0.939(16)
O4 -C10	1.3365(15)	C3 -H32	0.991(16)
O4 -C11	1.4838(17)	C5 -H51	0.94(2)
O5 -C19	1.2351(14)	C6 -H61	0.94(2)
N1 -C2	1.4555(16)	C7 -H71	1.00(2)
N1 -C10	1.3422(17)	C8 -H81	1.06(2)
N2 -C19	1.3255(17)	C9 -H91	1.01(2)
N2 -C20	1.453(2)	C12 -H121	1.02(3)
C1 -C2	1.5318(17)	C12 -H122	0.95(2)
C2 -C3	1.5299(19)	C12 -H123	1.01(2)
C3 -C4	1.511(2)	C13 -H131	1.00(2)
C4 -C9	1.387(2)	C13 -H132	1.013(19)
C4 -C5	1.377(2)	C13 -H133	0.95(2)
C5 -C6	1.390(3)	C14 -H141	1.03(3)
C6 -C7	1.378(3)	C14 -H142	0.97(2)
C7 -C8	1.360(3)	C14 -H143	0.99(2)
C8 -C9	1.387(3)	C15 -H151	0.981(14)
C11 -C14	1.507(3)	C16 -H161	1.002(15)
C11 -C13	1.511(2)	C17 -H171	1.00(2)
C11 -C12	1.523(3)	C17 -H172	0.980(18)
C15 -C16	1.5267(18)	C17 -H173	0.98(2)
C15 -C19	1.5221(17)	C18 -H181	1.03(2)
C16 -C18	1.516(2)	C18 -H182	1.04(2)
C16 -C17	1.518(2)	C18 -H183	0.98(2)
C20 -C21	1.513(2)	C20 -H201	0.991(17)
C21 -C26	1.378(2)	C20 -H202	0.989(16)
C21 -C22	1.392(2)	C22 -H221	0.95(2)
C22 -C23	1.375(2)	C23 -H231	0.955(18)
C23 -C24	1.367(3)	C24 -H241	0.95(2)
C24 -C25	1.381(3)	C25 -H251	0.94(2)
C25 -C26	1.390(3)	C26 -H261	0.941(18)

Table S6 - Bond Angles (Degrees) for DOEA 4182-173

C1	-O1	-C15	118.69(9)
C10	-O4	-C11	120.34(10)
C2	-N1	-C10	119.22(10)
C19	-N2	-C20	122.16(11)
O1	-C1	-C2	110.32(10)
O2	-C1	-C2	125.07(12)
O1	-C1	-O2	124.60(12)
N1	-C2	-C1	109.75(10)
N1	-C2	-C3	108.95(10)
C1	-C2	-C3	109.86(10)
C2	-C3	-C4	114.03(11)
C3	-C4	-C9	120.65(13)
C5	-C4	-C9	118.11(14)
C3	-C4	-C5	121.22(14)
C4	-C5	-C6	120.52(18)
C5	-C6	-C7	120.35(19)
C6	-C7	-C8	119.79(19)
C7	-C8	-C9	119.9(2)
C4	-C9	-C8	121.28(16)
O3	-C10	-N1	123.23(11)
O4	-C10	-N1	111.54(10)
O3	-C10	-O4	125.22(12)
O4	-C11	-C12	101.89(14)
O4	-C11	-C13	110.80(13)
O4	-C11	-C14	109.35(13)
C12	-C11	-C13	109.23(14)
C12	-C11	-C14	112.93(18)
C13	-C11	-C14	112.18(16)
O1	-C15	-C19	109.46(9)
C16	-C15	-C19	113.08(10)
O1	-C15	-C16	109.43(10)
C17	-C16	-C18	112.17(13)
C15	-C16	-C17	111.25(12)
C15	-C16	-C18	111.07(12)
O5	-C19	-C15	119.82(11)
N2	-C19	-C15	117.20(10)
O5	-C19	-N2	122.92(12)
N2	-C20	-C21	114.60(12)
C20	-C21	-C22	118.59(13)
C20	-C21	-C26	123.25(14)
C22	-C21	-C26	118.16(14)
C21	-C22	-C23	121.11(16)
C22	-C23	-C24	120.55(17)
C23	-C24	-C25	119.26(17)
C24	-C25	-C26	120.41(19)
C21	-C26	-C25	120.51(17)

Table S6 (cont.) - Bond Angles (Degrees) for DOEA 4182-173

C2 -N1 -H1	117.8(10)	O1 -C15 -H151	105.8(8)
C10 -N1 -H1	120.2(10)	C16 -C15 -H151	110.3(8)
C20 -N2 -H2	116.8(11)	C19 -C15 -H151	108.5(8)
C19 -N2 -H2	120.5(11)	C15 -C16 -H161	104.2(9)
N1 -C2 -H21	110.1(7)	C17 -C16 -H161	108.6(8)
C1 -C2 -H21	106.6(7)	C18 -C16 -H161	109.3(8)
C3 -C2 -H21	111.6(7)	C16 -C17 -H171	106.4(12)
C2 -C3 -H31	106.4(10)	C16 -C17 -H172	109.7(10)
C2 -C3 -H32	106.3(9)	C16 -C17 -H173	111.8(12)
C4 -C3 -H31	109.3(9)	H171 -C17 -H172	112.1(16)
C4 -C3 -H32	111.5(9)	H171 -C17 -H173	107.7(17)
H31 -C3 -H32	109.2(13)	H172 -C17 -H173	109.2(15)
C4 -C5 -H51	125.1(12)	C16 -C18 -H181	112.7(11)
C6 -C5 -H51	114.4(12)	C16 -C18 -H182	111.6(12)
C5 -C6 -H61	118.8(14)	C16 -C18 -H183	110.6(11)
C7 -C6 -H61	120.8(14)	H181 -C18 -H182	104.0(17)
C6 -C7 -H71	120.9(13)	H181 -C18 -H183	105.6(16)
C8 -C7 -H71	119.3(13)	H182 -C18 -H183	112.0(15)
C7 -C8 -H81	119.9(12)	N2 -C20 -H201	108.4(9)
C9 -C8 -H81	120.2(12)	N2 -C20 -H202	107.6(10)
C4 -C9 -H91	118.9(12)	C21 -C20 -H201	107.0(9)
C8 -C9 -H91	119.9(12)	C21 -C20 -H202	109.5(9)
C11 -C12 -H121	110.1(15)	H201 -C20 -H202	109.6(13)
C11 -C12 -H122	109.4(13)	C21 -C22 -H221	117.6(11)
C11 -C12 -H123	104.7(13)	C23 -C22 -H221	121.3(11)
H121 -C12 -H122	114(2)	C22 -C23 -H231	119.6(11)
H121 -C12 -H123	112.0(19)	C24 -C23 -H231	119.8(11)
H122 -C12 -H123	105.6(17)	C23 -C24 -H241	122.8(11)
C11 -C13 -H131	110.6(12)	C25 -C24 -H241	118.0(11)
C11 -C13 -H132	113.1(11)	C24 -C25 -H251	120.5(12)
C11 -C13 -H133	107.2(13)	C26 -C25 -H251	119.1(12)
H131 -C13 -H132	108.8(16)	C21 -C26 -H261	120.4(11)
H131 -C13 -H133	104.3(18)	C25 -C26 -H261	119.1(11)
H132 -C13 -H133	112.5(17)		
C11 -C14 -H141	105.5(13)		
C11 -C14 -H142	107.7(12)		
C11 -C14 -H143	111.2(13)		
H141 -C14 -H142	109.4(17)		
H141 -C14 -H143	117(2)		
H142 -C14 -H143	105.9(17)		